### FACULDADE DE ODONTOLOGIA

## AVALIAÇÃO MACRO E MICROSCÓPICA DE LESÕES ORAIS INDUZIDAS POR PROCEDIMENTOS CIRÚRGICOS EM RATOS SOB TERAPIA COM BISFOSFONATOS

ANA CAROLINA UCHOA VASCONCELOS

2012



ANA CAROLINA UCHOA VASCONCELOS

## AVALIAÇÃO MACRO E MICROSCÓPICA DE LESÕES ORAIS INDUZIDAS POR PROCEDIMENTOS CIRÚRGICOS EM RATOS SOB TERAPIA COM BISFOSFONATOS

## MACRO AND MICROSCOPIC EVALUATION OF LESIONS INDUCED BY ORAL SURGICAL PROCEDURES IN RATS UNDER THERAPY WITH BISPHOSPHONATES

Porto Alegre 2012



PONTIFÍCIA UNIVERSIDADE CATÓLICA DO RIO GRANDE DO SUL PROGRAMA DE PÓS-GRADUAÇÃO EM ODONTOLOGIA

### AVALIAÇÃO MACRO E MICROSCÓPICA DE LESÕES ORAIS INDUZIDAS POR PROCEDIMENTOS CIRÚRGICOS EM RATOS SOB TERAPIA COM BISFOSFONATOS

### MACRO AND MICROSCOPIC EVALUATION OF LESIONS INDUCED BY ORAL SURGICAL PROCEDURES IN RATS UNDER THERAPY WITH BISPHOSPHONATES

Tese apresentada como requisito para obtenção do título de Doutor pelo Programa de Pós-Graduação da Faculdade de Odontologia da Pontifícia Universidade Católica do Rio Grande do Sul. Área de Concentração, Estomatologia Clínica

### ANA CAROLINA UCHOA VASCONCELOS

Orientadora: Prof<sup>a</sup> Dr<sup>a</sup> Karen Cherubini

Porto Alegre 2012

DEDICATÓRIA

Dedico este trabalho aos meus pais, Fátima e Paulo, e às minhas irmãs Fabíola e Paula, pelo amor generoso e desprendido. Amo-os profunda e infinitamente.

## AGRADECIMENTOS

À minha orientadora, Dra. Karen Cherubini, exemplo de dedicação, competência e responsabilidade pelo exercício acadêmico. Sou extremamente grata pelo ensinamento generoso e, sobretudo, pela constante (e incansável) disponibilidade a mim dedicada durante todo o período em que convivemos. Por sua grande contribuição ao meu amadurecimento profissional, muito obrigada!

À minha avó Urçula, aos meus tios e cunhado, pela ternura, confiança e carinho constantes.

Às minhas sobrinha e afilhada, lindas e cheias de vida, Beatriz e Clara, por não me deixarem esquecer que a vida, quando colorida de *pink* com brilhos, pode ser muito mais divertida.

Ao meu sobrinho Pedro, por entender que, "por uns tempos", a tia Carol precisou morar no computador.

Às minhas grandes e especiais amigas Cláudia, Carla, Tatiana, Flávia, Sílvia, Mara e Emmanuela. Este trabalho jamais teria acontecido sem o apoio de vocês. Obrigada por fazerem a minha vida valer muito a pena! Orgulho imenso em tê-las presentes comigo. Amo, de uma forma especial, cada uma de vocês!

Às professoras do Doutorado em Estomatologia Clínica, Dra. Liliane Soares Yurgel, Dra. Maria Antonia Zancanaro de Figueiredo e Dra. Fernanda Gonçalves Salum, pelo exemplo de competência e profissionalismo.

Ao Dr. Alan Arrieira Azambuja, pelo apoio atencioso e dedicado durante a pesquisa.

Ao professor Dr. Vinícius Duval da Silva, pelo apoio técnico fornecido durante a etapa de captura e análise das imagens histológicas.

Ao Técnico do Laboratório de Anatomia Patológica e Citopatologia do Hospital São Lucas da PUC-RS, Tiago Giulliani Lopes, por toda a disponibilidade.

Aos colegas do Programa de Pós-graduação, em especial a Maria Noel, Márcia Fava, Wâneza Borges, Soraya Berti, Jonas Dantas e Luiz Arteche (*in memoriam*), pelos bons momentos compartilhados nesses anos de PUC-RS.

A Luciana Giacometti, por todo apoio.

Aos funcionários do Programa de Pós-Graduação em Odontologia, Ana Lúcia, Paulo, Marcos e Davenir, pela atenção e competência.

À Faculdade de Farmácia da PUCRS, por disponibilizar o suporte técnico necessário à realização do experimento.

Ao CNPq, pelo apoio financeiro.

EPÍGRAFE

Não existem fatos, apenas interpretações.

**Friedrich Nietzsche** 

(1844-1900)

## RESUMO

### RESUMO

A presente pesquisa teve por objetivo comparar a influência dos bisfosfonatos ácido zoledrônico e clodronato no reparo de feridas cirúrgicas induzidas por meio de exodontia e lesão de tecido mole oral. Trinta e quatro ratos (Rattus novergicus, linhagem Wistar) foram distribuídos em três grupos: (1) 12 animais tratados com ácido zoledrônico; (2) 12 animais tratados com clodronato; e (3) 10 animais que receberam solução salina. Decorridos 90 dias do início do tratamento, os animais foram submetidos a exodontias e procedimentos cirúrgicos em tecido mole, ambos na maxila. Aos 180 dias de administração dos fármacos, foi realizada a eutanásia. Após avaliação macroscópica, as maxilas foram processadas, e os cortes histológicos corados por hematoxilina e eosina (H&E), bem como submetidos a processamento imunoistoquímico com os marcadores RANKL, OPG, von Willebrand e caspase-3. Nas lâminas coradas por H&E, foram quantificadas as variáveis osso não-vital, osso vital, infiltrado inflamatório, colônias microbianas, tecido epitelial e tecido conjuntivo, sendo que, no sítio das exodontias, os restos radiculares também foram avaliados. A análise histológica foi realizada por meio do programa Image Pro Plus. Na avaliação macroscópica, no sítio das exodontias o grupo ácido zoledrônico exibiu associação com solução de continuidade da mucosa (qui-quadrado, análise de resíduos ajustados, p < 0.001), enquanto na área de lesão de tecido mole, nenhum grupo exibiu essa associação (qui-quadrado, p=0,151). No sítio das exodontias, o grupo ácido zoledrônico exibiu proporção significativamente maior de osso não-vital e colônias microbianas do que os demais grupos. Na lesão de tecido mole, a proporção de osso não-vital e colônias microbianas foi significativamente maior nos grupos ácido zoledrônico e clodronato do que no controle. Não houve diferença significativa das variáveis tecido epitelial, infiltrado inflamatório e resto radicular entre os grupos (Kruskal-Wallis teste de comparações múltiplas, p>0,05). A expressão imunoistoquímica de RANKL, OPG, von Willebrand e caspase-3 também não diferiu significativamente entre os grupos (Kruskal-Wallis, p>0.05). Os resultados permitem concluir que (1) ambos, ácido zoledrônico e clodronato, são capazes de induzir osteonecrose; (2) de acordo com a análise imunoistoquímica, é pouco provável que eventos intrínsecos à mucosa oral sejam os iniciadores da osteonecrose.

Palavras-chave: bisfosfonatos, osteonecrose, marcadores biológicos, ratos

## SUMMARY

#### SUMMARY

The aim of this work was to compare clodronate and zoledronic acid effect on the repair of surgical wounds induced by tooth extraction and oral soft tissue lesion. Thirty-four rats (Rattus novergicus, Wistar) were allocated into 3 groups: (1) 12 animals treated with zoledronic acid; (2) 12 animals treated with clodronate; and (3) 10 animals that were given saline solution. Elapsed 90 days from the beginning of the treatment, the animals were subjected to tooth extractions and surgical-induced soft tissue injury in maxillae. At 180 days of drug administration, they were euthanized. After macroscopic evaluation, maxillae were processed and histological cuts were stained with hematoxylin and eosin (H&E). Immunohistochemical expression of RANKL, OPG, von Willebrand and caspase-3 was also evaluated. Non-vital bone, inflammatory infiltrate, microbial colonies, epithelial tissue, connective tissue and vital bone were quantified at the tissue wound sites. At the tooth extraction site, root fragments were also evaluated. The variables were quantified with Image Pro Plus software. Macroscopic analysis at the tooth extraction site showed that zoledronic acid group was associated with loss of mucosal integrity (chi-square, residual adjusted analysis, p < 0.001), whereas at the soft tissue wound site, no group showed this association (chi-square, p=0.151). At the tooth extraction site, the zoledronic acid group showed greater proportion of non-vital bone and microbial colonies in comparison with the other groups. At the soft tissue wound site, the proportions of nonvital bone and microbial colonies were greater in the zoledronic acid and clodronate groups than in the control group. There was no significant difference for epithelial tissue, inflammatory infiltrate and root fragments between the groups (Kruskal-Wallis test complemented by its multiple comparisons test, p>0.05). Immunohistochemical expression of RANKL, OPG, von Willebrand and caspase-3 at tooth extraction and soft tissue wound sites did not differ significantly between the three groups analyzed (Kruskal-Wallis test, p>0.05). According to the results, (1) both bisphosphonates zoledronic acid and clodronate are capable of inducing osteonecrosis; (2) the immunohistochemical analysis suggests that the involvement of soft tissues as the initiator of osteonecrosis development is less probable than has been pointed out.

Key words: bisphosphonates, osteonecrosis, biological markers, rats

LISTA DE ILUSTRAÇÕES

### LISTA DE ILUSTRAÇÕES

**Figura 1-Artigo 1** Osteoclast (white arrow) in H&E stain (original magnification x 400) (A); Osteoclasts stained by TRAP (original magnification x 400) (B); Immunostaining of RANKL (original magnification x 400) (C); Immunostaining of OPG (original magnification x 200) (D)\_\_\_\_\_\_36

**Figura 1-Artigo 2** Quantification of histological features in one of the 3 fields examined at the site of tooth extraction using Image Pro Plus 4.5.1 software (Media Cybernetics, Bethesda, MD, USA; H&E stain, x 5 objective)\_\_\_\_\_63

 Figura 2-Artigo 2 Quantification of immunohistochemical expression of OPG by means of semi-automated segmentation technique using Image Pro Plus 4.5.1 software (Media Cybernetics, Bethesda, MD, USA; x10 objective)\_\_\_\_\_\_\_64

Figura 3-Artigo 2 Persistent soft tissue defect showing the disruption of epithelium (H&E stain, original magnification x 100) (A); Non-vital bone, microbial colonies, and inflammatory infiltrate (H&E stain, original magnification x 200) (B); Complete tissue repair (hematoxylin-eosin stain, original magnification x 100) (C); Vital bone (H&E stain, original magnification x 400) (D); Immunostaining of RANKL (original magnification x 200) (E); Immunostaining of OPG (original magnification x 400) (F)\_\_\_\_\_\_70

LISTA DE TABELAS

### LISTA DE TABELAS

**Tabela 2** Sample distribution according to presence/absence of non-vital bone at thetooth extraction and soft tissue wound sites on microscopic examination66

**Tabela 3** Quantification of histological features (H&E stain) at the tooth extraction sitein the zoledronic acid, clodronate and control groups67

 Tabela 4 Quantification of histological features (H&E stain) at the soft tissue wound site in the zoledronic acid, clodronate and control groups \_\_\_\_\_\_68

 Tabela 5 Immunohistochemical quantification of RANKL, OPG, von Willebrand factor (vWF) and caspase-3 at tooth extraction site in the zoledronic acid, clodronate and control groups \_\_\_\_\_\_69

 Tabela 6 Immunohistochemical quantification of OPG, RANKL, von Willebrand factor (vWF) and caspase-3 at soft tissue wound site in the zoledronic acid, clodronate and control groups \_\_\_\_\_\_69

LISTA DE SIGLAS

### LISTA DE SIGLAS

BMP	Bone morphogenetic protein
CD 31	Cluster of differentiation 31
CD 34	Cluster of differentiation 34
СТХ	Carboxy terminal collagen cross links
DNA	Deoxyribonucleic acid
ELISA	Enzyme linked immunosorbent assay
H&E	Hematoxylin-eosin
IL-1	Interleukin-1
JPEG	Joint Photographic Expert Group
M-CSF	Macrophage colony-stimulating factor
MMP	Matrix metalloproteinase
NTX	Aminoterminal telopeptide
OCIF	Osteoclastogenesis inhibitory factor
OCN	Osteocalcin
OPG	Osteoprotegerin
OPGL	OPG ligand
OPN	Osteopontin
PCR	Polymerase chain reaction
РТН	Parathyroid hormone
RANK	Receptor activator of nuclear factor-kappa B
RANKL	Receptor activator of nuclear factor- kappa-B ligand
RT-PCR	Reverse transcription-polymerase chain reaction
RUNX2	Runt-related transcription factor 2
TGF-β1	Transforming growth factor beta 1
TNF	Tumor necrosis factor
TRANCE	TNF-related activation-induced cytokine
TRAP	Tartrate resistant acid phosphatase
TUNEL	Terminal deoxynucleotidyl transferase-mediated dUTP nick endlabeling
VEGF	Vascular endothelial growth factor
vWF	Willebrand factor

## SUMÁRIO

## SUMÁRIO

1 INTRODUÇÃO	23
2 ARTIGO 1	25
Introduction	28
Evaluation of bisphosphonate effects on bone cells	29
Evaluation of bisphosphonate effects on angiogenesis	40
Final considerations	42
References	44
3 ARTIGO 2	53
Introduction	57
Material and methods	58
Results	64
Discussion	70
References	79
4 DISCUSSÃO GERAL	84
REFERÊNCIAS	89
ANEXOS	102

## INTRODUÇÃO

### 1 INTRODUÇÃO

Bisfosfonatos são análogos sintéticos do pirofosfato inorgânico, um regulador endógeno do metabolismo ósseo, e são empregados na prevenção e no tratamento de doenças caracterizadas por excessiva reabsorção óssea. Esses fármacos podem ser classificados, de acordo com a presença ou ausência de nitrogênio em sua composição, respectivamente, em nitrogenados e não-nitrogenados. Ácido zoledrônico, alendronato, risedronato, olpadronato e icandronato pertencem ao grupo dos nitrogenados, enquanto clodronato, tiludronato e etidronato classificam-se como não-nitrogenados (MIGLIORATI et al., 2005; RUGGIERO et al., 2009).

A osteonecrose por bisfosfonato é uma condição que se caracteriza por exposição óssea persistente no complexo maxilomandibular de pacientes submetidos ao tratamento com o fármaco, mas sem histórico de radioterapia da região de cabeça e pescoço. O achado clínico habitual consiste em área de mucosa ulcerada com exposição de osso desvitalizado localizada, mais frequentemente, na região póstero-lingual da mandíbula (RUGGIERO; WOO, 2008; RUGGIERO et al., 2009). Os achados radiográficos são inespecíficos e podem assemelhar-se a lesão periapical, osteomielite e neoplasia maligna primária ou metastática. Ao exame histológico, podem-se observar áreas compatíveis com necrose óssea evidenciada por regiões de infiltrado celular inflamatório e osso avascular e acelular (KHOSLA et al., 2007). A condição é de difícil tratamento e este, em muitos casos, visa apenas a preservar a qualidade de vida do paciente por meio do controle da dor e da infecção, bem como prevenir a ocorrência de novas áreas de necrose (RUGGIERO; WOO, 2008).

A hipótese de que a etiopatogenia da osteonecrose por bisfosfonatos tem origem multifatorial é suportada por estudos que apontam os efeitos do fármaco sobre o metabolismo ósseo (KELLINSALMI et al., 2005; SENEL et al., 2010; SONIS et al., 2009) e sobre a vascularização sanguínea (BI et al., 2010; NAIDU et al., 2008; WALTER et al., 2010; YAMADA et al., 2009; WOOD et al., 2002), bem como pela participação de agentes infecciosos (KOBAYASHI et al., 2010). Pesquisas revelam, ainda, que os bisfosfonatos são tóxicos ao epitélio oral, o que suscita dúvidas quanto à origem da lesão ser no tecido ósseo ou na mucosa de revestimento (LANDESBERG et al., 2008). Além disso, a maior parte das pesquisas que avaliam os efeitos do fármaco sobre os tecidos e sua relação com a osteonecrose são realizadas com bisfosfonatos nitrogenados, sendo poucas as investigações sobre o comportamento tecidual mediante o emprego dos não-nitrogenados.

A presente tese é composta por dois trabalhos apresentados sob a forma de artigos científicos. O primeiro teve por objetivo apresentar uma revisão da literatura concernente a métodos laboratoriais e biomarcadores disponíveis para avaliação dos efeitos biológicos dos bisfosfonatos. O segundo descreve o experimento, cujo objetivo foi comparar a influência dos bisfosfonatos nitrogenados (ácido zoledrônico) e não nitrogenados (clodronato) no reparo de feridas cirúrgicas induzidas em osso e em tecido mole da maxila.

## **ARTIGO 1**

### 2 ARTIGO 1

O artigo a seguir intitula-se Laboratory methods and biomarkers in the evaluation of bisphosphonate effects on body tissues - A literature review e foi formatado de acordo com as normas do periódico *Oral Oncology* (Anexo A).

# Laboratory methods and biomarkers in the evaluation of bisphosphonate effects on body tissues - A literature review

Ana Carolina Uchoa Vasconcelos<sup>1</sup> Soraya de Azambuja Berti<sup>1</sup> Maria Antonia Figueiredo<sup>2</sup> Fernanda Gonçalves Salum<sup>2</sup> Tiago Giulliani Lopes<sup>3</sup> Karen Cherubini<sup>2</sup>

<sup>1</sup> Ph.D. Student, Postgraduate Program, Dental College, Pontifical Catholic University of Rio Grande do Sul

<sup>2</sup> Ph.D., Postgraduate Program, Dental College, Pontifical Catholic University of Rio Grande do Sul

<sup>3</sup> AS, Department of Pathology, School of Medicine, Hospital São Lucas, Pontifical Catholic University of Rio Grande do Sul - PUCRS

### **Corresponding author**

Karen Cherubini Serviço de Estomatologia, Hospital São Lucas PUCRS Av Ipiranga, 6690, sala 231 Porto Alegre RS Brazil CEP 90610-000 Telephone/fax: 55(51)33203254 E-mail: karen.cherubini@pucrs.br/ kebini.ez@terra.com.br

Running title: Biomarkers and bisphosphonates

### ABSTRACT

Bisphosphonates are extensively used to treat bone metabolism disorders, but these drugs have an important side effect - jaw osteonecrosis. Therefore, studies have been conducted aimed at better understand their mechanism of action and to determine a course to neutralize this important side effect. We present here a literature review focusing on the laboratory methods available for investigating bisphosphonate effects on body tissues. There are many different methods available for this purpose, but the correct indication of these techniques and the knowledge of their limitations are of crucial importance for the understanding of bisphosphonate effects on body tissues.

Key words: bisphosphonates; biomarkers; tissues

### Introduction

Bisphosphonates are drugs used in the prevention and treatment of bone metabolism diseases with intense resorption activity.<sup>1,2</sup> This is because of their ability to inhibit both bone resorption<sup>3</sup> and angiogenesis.<sup>4</sup> However, some effects of these drugs on non-mineralized tissues have also been reported.<sup>5,6</sup>

These compounds are able to interfere with remodeling of bone tissue by acting on different cells.<sup>4,7-9</sup> Their effect on osteoclasts occurs by inhibiting recruitment and differentiation, decreasing life span<sup>10</sup> and promoting apoptosis.<sup>7,11,12</sup> The mechanism through which bisphosphonates affect osteoblasts is still unclear, but some *in vitro* studies have demonstrated that high concentrations of nitrogen-containing bisphosphonates inhibit osteoblast proliferation, adhesion, and migration<sup>13,14</sup> as well as inducing osteoblasts to produce osteoprotegerin (OPG).<sup>15</sup> Studies have shown that zoledronic acid and, to a lesser extent, clodronate have antiangiogenic effects, tending to inhibit proliferation, migration,

and adhesion of endothelial cells.<sup>16,17</sup> Zoledronic acid inhibits the proliferation and migration of these cells *in vitro* in a dose-dependent way.<sup>18</sup> Studies have also postulated that bisphosphonates, especially the most potent ones, are able to modulate the secretion of specific growth factors, such as transforming growth factor  $\beta 1$  (TGF- $\beta 1$ ).<sup>13</sup>

Adverse effects related to therapy with these drugs are sporadic and depend on the class of the drug, as well as the route and frequency of administration. Bisphosphonate-related ostonecrosis of the jaws was first described in 2003 and is characterized by an area of exposed bone in the maxillofacial region, which persists for at least eight weeks in patients undergoing therapy with these drugs and with no history of head and neck radiation therapy.<sup>19-21</sup> Several theories have been proposed to explain the pathogenesis of this lesion, whose etiology is multifactorial.<sup>22</sup> It is believed that the impact of the drug on different cell types is capable of contributing to the difficulty of tissue healing after the damage, resulting in clinical manifestation of the disease. The challenge of treating osteonecrosis has demanded research on the mechanism of action of bisphosphonate and its effects on biological tissues,<sup>23</sup> aiming to establish criteria to guide the management of patients using bisphosphonate or with osteonecrosis.<sup>24</sup> The present study reviewed the scientific literature with regard to laboratory methods and biomarkers available for the evaluation of biological effects of bisphosphonates, and we discuss here their indication, limitations, advantages and disadvantages.

### Evaluation of bisphosphonate effects on bone cells

### *Hematoxylin-eosin evaluation (H&E)*

On H&E examination, bisphosphonate-related osteonecrosis shows non-vital bone tissue infected by microorganisms, as well as inflammatory areas.<sup>25,26</sup> In bisphosphonate users without osteonecrosis, bone tissue shows some morphologic changes, with

osteoclasts being the focus of attention. In this examination, they appear as acidophilic multinucleated giant-cells (Fig. 1A). The active osteoclasts are found intimately associated with the bone surface through their active pole. At this site, the ruffled border and the clear zone are noted, which is a small area demarking the borders of the region to be reabsorbed. The osteoclast-bone interface is poorly stained and is not easily seen. The inactive osteoclasts, in turn, do not stay in contact with bone structure and tend to assume a rounded shape. The H&E technique is easily performed, has low cost and also permits the visualization of active and inactive osteoclasts, an important feature regarding bisphosphonates. The major limitation lies in the identification of osteoclasts with just one or no nucleus. In these cases, they could be wrongly classified as pre-osteoclasts or not even be identified.<sup>27</sup>

There are some reports about the decrease in osteoclasts in bone under aminobisphosphonate treatment,<sup>28,29</sup> and histomorphometric evaluations in H&E have found increased trabecular density in alveolar bone,<sup>30</sup> tibia and femur.<sup>31,32</sup> Still, there are also reports of the absence of either changes in osteoclast number or significant increase in trabecular density in alveolar bone of rats treated with alendronate.<sup>33</sup>

### Tartrate-resistant acid phosphatase (TRAP)

TRAP is a glycoprotein synthesized by differentiated cells of the monohistiocytic system that can be found in plasma, placenta, macrophages and leukocytes, and is more significantly expressed in osteoclasts.<sup>34</sup> It is one of the proteins secreted in the bone-osteoclast interface, and its increased expression is directly related to bone resorption.<sup>35</sup> TRAP is used as a resorption marker and its activity is inhibited by calcitonin.<sup>36</sup>

TRAP can also be used as a serum marker of osteoclast activity<sup>34</sup> and is identified by a histochemical method, where it shows purple staining, because of the reduction of iron ions located at its active site.<sup>27</sup> The technique has been applied in investigations of bisphosphonate effects using cell cultures<sup>37,38</sup> and animal models,<sup>39,40</sup> showing a decreased number of osteoclasts after bisphosphonate use.

As TRAP staining only detects osteoclast cells (Fig. 1B), it is more specific than H&E. However, the specimen decalcifying process with strong acids such as nitric, formic and trichloroacetic impairs the TRAP technique. These acids cause enzymatic denaturation, which leads to failure in cell staining.<sup>41</sup> Moreover, as it is an enzymatic method, it is highly sensitive to environmental factors such as pH and temperature variation.<sup>35</sup>

### Terminal deoxynucleotidyl transferase-mediated dUTP nick end-labeling (TUNEL)

TUNEL detects apoptotic cells *in situ* by means of specific staining of fragmented DNA. It has already been used to investigate bisphosphonate effects on keratinocytes,<sup>5,42</sup> endothelial cells, fibroblasts, osteoblasts<sup>43,44</sup> and osteoclasts.<sup>45</sup> However, the specificity of this technique has been discussed regarding the fact that the increase in nuclear condensation during apoptosis impairs the access of TUNEL reagents to the target structures (apoptotic nucleus). Also, the method can show false positive results in cases of necrotic or mitotic cells staining.<sup>46</sup>

### Combined TRAP and TUNEL technique

When stained exclusively by TRAP, osteoclasts exhibit notable cytoplasmic activity, which is revealed by a red color. If these cells are subjected to TUNEL, the positivity is identified by vacuoles containing yellowish-brown bodies. Therefore, TUNEL

and TRAP combination shows apoptotic osteoclasts as red cells containing yellowishbrown vacuoles.<sup>47,48</sup>

When compared with caspases, especially with the effector ones, the combined TRAP/TUNEL technique can show lower specificity because of the possible staining of necrotic and mitotic osteoclasts.<sup>47</sup> Still, this combined technique allows the nucleus of apoptotic osteoclasts to be distinguished from that of osteoblasts or osteocytes, which have casually been internalized by osteoclasts during bone resorption.<sup>47,48</sup>

#### Caspases

Apoptosis involves the activation of caspases, which make up a family of cysteine proteases that play a fundamental role in the cleavage of intracellular substrates. These proteins are synthesized as inactive precursors (pro-caspases), where they are activated after receiving a cell death signal.<sup>49,50</sup> Until now, 14 types of caspases have been identified, and they are classified according to their activity as inflammatory (1,4,5,11,12,13,14), initiators (2,8,9,10) and effectors (3,6,7). The activation of apoptosis can occur through two pathways. The extrinsic route is started by external signals such as free-radicals and physical or chemical agents, which stimulate cell surface receptors. The intrinsic route can occur as a result of cellular stress that causes morphofunctional changes in mitochondria. In both pathways, there is triggering of the caspase cascade, which culminates with caspase-3 activation.<sup>51</sup>

Caspases have been used in investigations into the effects of bisphosphonate on various cells, including bone and endothelial cells, fibroblasts and keratinocytes.<sup>6,18</sup> Studies have demonstrated pro- and antiapoptotic effects of bisphosphonates on osteoclasts and osteoblasts/osteocytes, respectively. Benford et al. (2001)<sup>49</sup> observed increased activity of caspase-3 in osteoclasts treated with nitrogen-containing bisphosphonates (alendronate,

zoledronic acid, risedronate, pamidronate) and non-nitrogen containing ones (clodronate, tiludronate, etidronate). According to these authors, high concentrations of clodronate, alendronate and zoledronic acid produce a similar proportion of apoptotic osteoclasts. This suggests that the greater antiresorptive effect of nitrogen-containing bisphosphonates is related not only to apoptosis, but also to effects on osteoclast migration and differentiation. Similarly, using caspase-3, Plotkin et al. (2006)<sup>52</sup> found that nitrogen- and non nitrogen-containing bisphosphonates exert apoptotic effect on osteoclasts. On the other hand, both groups of drugs inhibited osteoblast and osteocyte apoptosis at doses three times lower than that used for osteoclasts.

## Receptor activator of nuclear factor-kappa B (RANK), receptor activator of nuclear factorkappa B ligand (RANKL) and osteoprotegerin (OPG)

Bone remodeling is characterized by the balance between activation and apoptosis of osteoblasts and osteoclasts. Different stages of development of this process are regulated by diverse growth factors, cytokines and hormones, including macrophage colony-stimulating factor (M-CSF), interleukin-1 (IL-1), transforming growth factor  $\beta$  (TGF- $\beta$ ), tumor necrosis factor (TNF), D3-vitamin, calcitonin and parathyroid hormone (PTH).<sup>53</sup>

The discovery of a triad of proteins belonging to the TNF family and capable of regulating maturation, function and survival of osteoclasts has improved the understanding of bone metabolism.<sup>54</sup> Such proteins are RANKL also called TNF-related activation induced cytokine (TRANCE) or OPG ligand (OPGL), RANK, and OPG, also called osteoclastogenesis inhibitory factor (OCIF).<sup>55-57</sup>

Osteoclastogenesis requires osteoclast activation by two molecules characterized by complementary activities, M-CSF and RANKL. M-CSF increases osteoclast precursors, whereas RANKL, which is expressed on the osteoblast surface, exerts its effects when bound to RANK receptor expressed on the osteoclast precursor cell surface, inducing the signaling of the cascade that leads to osteoclast differentiation and maturation.<sup>55,58,59</sup>

RANK is a transmembrane receptor found in osteoclast precursor cells, dendritic cells, fibroblasts and T cells.<sup>60</sup> RANKL, on the other hand, is a protein produced by osteoblasts and T cells, which can be expressed as a cell membrane cytokine or be released as a soluble factor. The two forms of RANKL have different functional properties, where the membrane receptor is known as a more effective osteoclastogenesis mediator. These two proteins (RANK and RANKL) can be quantified by many methods. Nevertheless, immunosorbent assay (ELISA) has the limitation of detecting just the soluble form of RANKL, which is less effective in expressing osteoclast formation.<sup>56</sup>

OPG is a protein expressed by a variety of cell types, including immune cells, osteoblasts and endothelial cells. It is considered an antagonist receptor of RANKL, as it blocks the RANK/RANKL interaction and inhibits the final stage of osteoclast differentiation. All these proteins show cytoplasmic staining (Fig.1 C and D). *In vitro* studies report that OPG effects include inhibition of both differentiation and activation of osteoclasts, therefore reducing bone resorption.<sup>61</sup>

Investigations into the relationship between serum levels of RANKL and OPG in patients undergoing bisphosphonate therapy have shown controversial results. Martini et al. (2006)<sup>56</sup> observed serum OPG levels significantly higher in patients with Paget disease treated with nitrogen-containing bisphosphonates. However, Alvarez et al. (2003)<sup>62</sup> reported a decrease in serum OPG in Paget patients after treatment with tiludronate. This same study, however, did not observe changes in serum levels of RANKL. Mountzios et al. (2010),<sup>63</sup> in turn, did not find any significant change in serum levels of OPG and RANKL in patients with bone metastases treated with zoledronic acid.

*In vitro* studies report that nitrogen-containing bisphosphonates increase the expression of OPG and decrease the expression of RANKL in different cell types. Pan et al. (2004)<sup>64</sup> investigated the effect of zoledronic acid on OPG and RANKL expression in osteoblast cultures using flow cytometry, Western blotting and ELISA. According to this study, bisphosphonates inhibit osteoclast differentiation through decreased expression of RANKL and increased OPG secretion in osteoblasts. This finding corroborates the report of Viereck et al. (2002),<sup>15</sup> who demonstrated by means of reverse transcription-polymerase chain reaction (RT-PCR) and ELISA, that zoledronic acid and sodium pamidronate increase the expression and secretion of OPG in osteoblasts in a dose-dependent manner. According to Tipton et al. (2011),<sup>57</sup> pamidronate and alendronate increased OPG production and decreased RANKL in gingival fibroblasts, changing the OPG/RANKL ratio, which suggests that these cells, located next to resorption sites, can be directly involved in alveolar bone metabolism regulation.

The RANK/RANKL/OPG triad shows a strong potential to be used in diagnosis and follow-up of patients with bone metabolism diseases. However, these markers still have restricted indication in clinical practice, because of their technical limitations, which include interference by physiological factors. Thus, they cannot be indicated without considering the clinical criteria usually applied to the diagnosis of the disease.<sup>56</sup>


**Figure 1**. Osteoclast (white arrow) in H&E stain (original magnification x 400) (A); Osteoclasts stained by TRAP (original magnification x 400) (B); Immunostaining of RANKL (original magnification x 400) (C); Immunostaining of OPG (original magnification x 200) (D).

# Osteocalcin (OCN)

OCN is a peptide secreted by mature osteoblasts, hypertrophic chondrocytes and odontoblasts, classified as the most abundant noncollagenous protein of bone matrix.<sup>65,66</sup> The affinity to hydroxyapatite seems to be sufficient for this protein to play an important role in the regulation and maturation of crystal and preservation of bone matrix.<sup>67</sup> OCN is considered a marker of mature osteoblasts, as it increases with the start of the mineralization process.<sup>68</sup> The lack of specificity is a limitation to the use of this marker because both osteolysis and osteogenesis release this protein into the serum.<sup>66</sup>

Studies on bisphosphonate interference with OCN levels suggest that these compounds cause metabolic changes in osteoblasts.<sup>69,70</sup> Koch et al. (2010)<sup>71</sup> evaluated gene expression of OCN by means of RT-PCR. These authors found that zoledronic acid and

ibandronate increase, in a dose-dependent manner, the expression of OCN in osteoblasts, whereas clodronate does not significantly change its expression, even when administered at high doses. Therefore, it is probable that nitrogen-containing bisphosphonates increase bone density by stimulating osteoblast differentiation.

# Osteopontin (OPN)

OPN is a glycoprotein involved in many physiological and pathological events, including processes of immune response, tumorigenesis, and cell proliferation, migration and differentiation. It exists either as a membrane-bound molecule of the extracellular matrix in mineralized tissues, or as a cytokine in body fluids. In bone tissue, it is found in pre-osteoblasts, osteocytes and osteoclasts.<sup>72</sup> It is expressed not only in bone tissue, but also in macrophages, lymphocytes, dentin, cementum, kidneys and brain, which reflects its multiplicity of functions.<sup>73</sup>

OPN is upregulated by hormones and cytokines, such as calcitonin and fibroblast growth factors, and suppressed by bisphosphonates.<sup>53</sup> Downregulation by bisphosphonates in bone is compatible with osteoclast activity suppression.<sup>73</sup> Increased OPN was observed in cementoblasts treated with zoledronic acid.<sup>74</sup> Also, serum levels of OPN were increased in patients with bone metastases after treatment with zoledronic acid,<sup>63</sup> and higher immunohistochemical expression of this protein in periodontal ligament cells of rats under etidronate therapy has been reported.<sup>75</sup> These reports support the idea of bisphosphonates having the ability to increase bone mass. Nevertheless, the association of OPN with a variety of diseases and physiological situations in response to stress makes it a low-specificity method to evaluate bisphosphonate effects on bone tissue.<sup>72</sup>

#### Runt-related transcription factor 2 (RUNX2)

The RUNX family comprises a group of three transcription factor proteins: Runx1, Runx2 and Runx3. These proteins are expressed in bone and hematopoietic system and are involved in a series of neoplasms either as a tumor promoter or suppressor.<sup>76</sup> RUNX2 exerts known effects on bone development through the regulation of osteoblast differentiation.<sup>76,77</sup>

Few studies have focused on bisphosphonate effects on RUNX2 levels. Koch et al. (2010)<sup>71</sup> observed by means of RT- PCR that zoledronic acid and ibandronate increase the expression of the RUNX2 gene in osteoblasts, whereas clodronate does not cause significant changes in this expression, even when administered at high concentrations.

# Alkaline phosphatase

Alkaline phosphatase is an ectoenzyme expressed in three isoforms, namely placental, intestinal and liver/bone/kidney forms, the latter being a nonspecific one which can be found in almost all tissues. The bone isoform, located on the surface of osteoblasts and chondrocytes, represents a marker of the middle stage of osteogenesis, appearing during the matrix maturation phase.<sup>68</sup>

A study analyzing the effect of zoledronic acid on metabolic activity of osteoblasts showed that the increase in drug concentration matched with a decrease in alkaline phosphatase production.<sup>78</sup> Other studies, on the other hand, found neither an increase in gene expression of alkaline phosphatase in osteoblasts treated with alendronate<sup>79</sup> nor significant changes in serum levels of alkaline phosphatase in patients treated with this drug.<sup>80</sup>

# Carboxy terminal collagen cross links (CTX)/amino-terminal telopeptide (NTX)

CTX and NTX are products from the degradation of type I collagen, the major organic component of extracellular matrix. During collagen degradation, CTX and NTX fragments are released into the blood circulation. As both segments are small enough to be eliminated by the renal route, they can be measured by serum and urine examination. These are not specific markers as they are found in tissues other than bone, and their detection should be interpreted in the clinical context.<sup>68,81</sup>

Urine levels have as a disadvantage the necessity of correcting results for creatinine excretion to compensate variations in urine dilution. This requires a second evaluation, which generates some complications inherent to the technique. According to some authors, as serum measurement does not have this limitation, it is the most reliable method to evaluate CTX and NTX.<sup>81</sup> Moreover, studies comparing urinary and serum results with these markers report that there is still no consensus about their correlation.<sup>82</sup> Regardless of the evaluation method or the marker used, the technique shows great variability according to the sampling time and processing and storage of specimens, as well as the age and sex of the patients. Therefore, it is crucial to consider these limitations to minimize interpretation errors.<sup>81</sup>

Studies report significant reduction of CTX and NTX in patients with bone metastases after 6 months of clodronate therapy<sup>83</sup> and in patients with osteoporosis treated with zoledronic acid (yearly) and alendronate (weekly).<sup>84</sup> These reports reinforce the idea that nitrogen- and non-nitrogen-containing bisphosphonates inhibit osteoclast function.<sup>81</sup>

# Bone morphogenetic proteins (BMPs) and matrix metalloproteinase proteins (MMPs)

BMPs comprise a series of proteins from TGF- $\beta$  family, which are involved in the morphogenesis and organogenesis of different tissues and organs. Until now, nine types of

BMPs have been described, and with exception of BMP-1, all of them seem to be capable of inducing bone formation. MMPs, in turn, are proteins capable of degrading the components of extracellular matrix and are involved in a variety of physiological and pathological events of bone remodeling and inflammatory response regulation. These proteins can be distributed into five subfamilies: collagenases (MMP-1, MMP-8, MMP-13), stromelysins (MMP-3, MMP-10), gelatinases (MMP-2, MMP-9), matrilysines (MMP-7, MMP-26) and membrane-type metalloproteinases (MMP-14, MMP-15, MMP-16, MMP-17, MMP-24, MMP-25), according to their structure and the type of substrate to which they bind.<sup>85</sup>

Studies report that bisphosphonate therapy is related to decreased expression of MMPs in periodontal tissue of rats,<sup>86</sup> as well as to a reduction in their serum levels in humans.<sup>87</sup> There are also reports about the association of these drugs with increased expression of BMPs in human bone marrow and periodontal tissue.<sup>88,89</sup>

#### **Evaluation of bisphosphonate effects on angiogenesis**

# Vascular endothelial growth factor (VEGF)

VEGF is the major mitogenic factor for endothelial cell growth, either in physiological or pathological conditions. VEGF is produced by keratinocytes, macrophages, fibroblasts and mastocytes, and it is capable of potentiating vascular permeability and is directly involved in vascular differentiation and formation, also working as an antiapoptotic factor in endothelial cells of recently formed vessels.<sup>90,91</sup>

It was reported that the immunohistochemical expression of VEGF in tooth extraction area did not significantly differ between rats treated with zoledronic acid and alendronate and controls.<sup>26</sup> Also, a prospective study did not show a significant reduction in serum VEGF levels in cancer patients treated with nitrogen- and non nitrogen-

containing bisphosphonates.<sup>92,93</sup> Nevertheless, there are reports of decreased levels of circulating VEGF in cancer patients after zoledronic acid administration,<sup>94,95</sup> as well as significantly reduced expression of VEGF caused by zoledronic acid *in vivo* and *in vitro*.<sup>96</sup>

#### Cluster of differentiation 34 (CD34)

CD34 is a glycoprotein expressed on the surface of progenitor or primitive cells of the hematopoietic system. Although highly sensitive, it shows poor specificity, a reason for which the morphological identification of the stained structures is an important auxiliary tool for the immunohistochemical method. The technique also has the disadvantage of staining lymphatic vessels, which can cause false-positive results.<sup>97</sup>

Animal models with prostate cancer under alendronate therapy showed significant reduction of CD34 immunohistochemical expression in capillary vessels.<sup>98</sup> Also in flow cytometry, serum levels of CD34 were significantly higher in patients with multiple myeloma treated with nitrogen-containing bisphosphonate who developed osteonecrosis than in those who did not have the lesion. This result suggests an association between osteonecrosis and reduced vascular proliferation.<sup>99</sup>

## Cluster of differentiation 31 (CD31)

CD31 is a protein expressed by hematopoietic and endothelial cells. An *in vivo* study showed no significant change in gene expression of CD31 in the gingival tissue of rats undergoing zoledronic acid therapy and subjected to oral lesion induction.<sup>100</sup> On the other hand, an *in vitro* study using CD31 immunohistochemistry observed reduced vascular density associated with either zoledronic acid or clodronate.<sup>101</sup>

vWF or factor VIII is a glycoprotein exclusively produced by endothelial cells and megakaryocytes. It is routinely used to identify blood vessels in histological samples.<sup>102</sup> When compared to other vascular markers, this is a highly specific protein for the detection of endothelial cells. Also, as it is released in situations of damage to these cells, vWF has been indicated as a marker of endothelial dysfunction.<sup>103</sup>

Freitas et al. (2005)<sup>104</sup> evaluated angiogenic activity in hemangiomas and pyogenic granulomas by immunostaining with anti-CD31, anti-VEGF and anti-vWF. The samples stained with anti-vWF showed a more uniform and more easily interpreted immunostaining pattern. However, CD 31 and vWF are not capable of differentiating new vessels from the preexisting ones, whereas VEGF is capable of overcoming this limitation (SCHOR et al., 1998).<sup>105</sup>

#### **Final considerations**

Bisphosphonates are relatively recent drugs whose effects on body tissues are partially known. Jaw osteonecrosis associated with these drugs has demanded new studies in this field. In this review, we list some markers that can serve this purpose. Actually, there are many other methods available, each one with its specificities, advantages and disadvantages. Most proteins presented here have been assessed through biochemical,<sup>67</sup> immunohistochemical<sup>86</sup> and genetic methods.<sup>88</sup> Moreover, studies using them in the investigation of bisphosphonate effects show controversial results, which seem to be supported by the lack of standardization of methods, type of bone structure evaluated, length of use, type and dose of the drug, as well as presence or absence of previous metabolically induced disease.<sup>33</sup> The correct indication of these techniques and the

understanding of their limitations are of crucial importance for understanding the effects of bisphosphonate on body tissues.

Acknowledgements

We thank Dr. A. Leyva (U.S.A) for English editing the manuscript.

# References

1. Ruggiero SL. Bisphosphonate-related osteonecrosis of the jaw (BRONJ): initial discovery and subsequent development. *J Oral Maxillofac Surg.* 2009;**67**(5):13-8.

2. Ruggiero SL. Bisphosphonate-related osteonecrosis of the jaw: an overview. *Ann N Y Acad Sci.* 2011;**1218**:38-46.

3. Marx RE, Sawatari Y, Fortin M, Broumand V. Bisphosphonate-induced exposed bone (osteonecrosis/osteopetrosis) of the jaws: risk factors, recognition, prevention, and treatment. *J Oral Maxillofac Surg.* 2005;**63**(11):1567-75.

4. Bi Y, Gao Y, Ehirchiou D, Cao C, Kikuiri T, Le A, et al. Bisphosphonates cause osteonecrosis of the jaw-like disease in mice. *Am J Pathol*. 2010;**177**(1):280-90.

5. Landesberg R, Cozin M, Cremers S, Woo V, Kousteni S, Sinha S, et al. Inhibition of oral mucosal cell wound healing by bisphosphonates. *Oral Maxillofac Surg*. 2008;**66**(5):839-47.

6. Cozin M, Pinker BM, Solemani K, Zuniga JM, Dadaian SC, Cremers S, et al. Novel therapy to reverse the cellular effects of bisphosphonates on primary human oral fibroblasts. *J Oral Maxillofac Surg.* 2011;**69**(10):2564-78.

7. Kellinsalmi M, Mönkkönen H, Mönkkönen J, Leskelä HV, Parikka V, Hämäläinen M, et al. In vitro comparison of clodronate, pamidronate and zoledronic acid effects on rat osteoclasts and human stem cell-derived osteoblasts. *Basic Clin Pharmacol Toxicol*. 2005;**97**(6):382-91.

8. Senel FC, Duman MK, Muci E, Cankaya M, Pampu AA, Ersoz S, et al. Jaw bone changes in rats after treatment with zoledronate and pamidronate. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2010;**109**(3):385-91.

9. Sonis ST, Watkins BA, Lyng GD, Lerman MA, Anderson KC. Bony changes in the jaws of rats treated with zoledronic acid and dexamethasone before dental extractions mimic bisphosphonate-related osteonecrosis in cancer patients. *Oral Oncol.* 2009;**45**(2):164-72.

10. Russell RG. Bisphosphonates: mode of action and pharmacology. *Pediatrics*. 2007;**119** (2):S150-62.

11. Biasotto M, Chiandussi S, Zacchigna S, Moimas S, Dore F, Pozzato G, et al. A novel animal model to study non-spontaneous bisphosphonates osteonecrosis of jaw. *J Oral Pathol Med.* 2010;**39**(5):390-6.

12. Senel FC, Saracoglu Tekin U, Durmus A, Bagis B. Severe osteomyelitis of the mandible associated with the use of non-nitrogen-containing bisphosphonate (disodium clodronate): report of a case. *J Oral Maxillofac Surg.* 2007;**65**(3):562-5.

13. Naidu A, Dechow PC, Spears R, Wright JM, Kessler HP, Opperman LA. The effects of bisphosphonates on osteoblasts in vitro. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2008;**106**(1):5-13.

14. Marolt D, Cozin M, Vunjak-Novakovic G, Cremers S, Landesberg R. Effects of Pamidronate on Human Alveolar Osteoblasts In Vitro. *J Oral Maxillofac Surg.* 2011; doi:10.1016/j.joms.2011.05.002.

15. Viereck V, Emons G, Lauck V, Frosch KH, Blaschke S, Gründker C, et al. Bisphosphonates pamidronate and zoledronic acid stimulate osteoprotegerin production by primary human osteoblasts. *Biochem Biophys Res Commun.* 2002 1;**291**(3):680-6.

16. Walter C, Klein MO, Pabst A, Al-Nawas B, Duschner H, Ziebart T. Influence of bisphosphonates on endothelial cells, fibroblasts, and osteogenic cells. *Clin Oral Investig.* 2010;**14**(1):35-41.

17. Yamada J, Tsuno NH, Kitayama J, Tsuchiya T, Yoneyama S, Asakage M, et al. Antiangiogenic property of zoledronic acid by inhibition of endothelial progenitor cell differentiation. *J Surg Res.* 2009;**151**(1):115-20.

18. Kobayashi Y, Hiraga T, Ueda A, Wang L, Matsumoto-Nakano M, Hata K, et al. Zoledronic acid delays wound healing of the tooth extraction socket, inhibits oral epithelial cell migration, and promotes proliferation and adhesion to hydroxyapatite of oral bacteria, without causing osteonecrosis of the jaw, in mice. *J Bone Miner Metab*. 2010;**28**(2):165-75.

19. Khosla S, Burr D, Cauley J, Dempster DW, Ebeling PR, Felsenberg D, et al. Bisphosphonate-associated osteonecrosis of the jaw: report of a task force of the American Society for Bone and Mineral Research. *J Bone Miner Res.* 2007;**22**(10):1479-91.

20. Landesberg R, Woo V, Cremers S, Cozin M, Marolt D, Vunjak-Novakovic G, et al. Potential pathophysiological mechanisms in osteonecrosis of the jaw. *Ann N Y Acad Sci*. 2011;**1218**:62-79. doi: 10.1111/j.1749-6632.2010.05835.x.

21. Rizzoli R, Burlet N, Cahall D, Delmas PD, Eriksen EF, Felsenberg D, et al. Osteonecrosis of the jaw and bisphosphonate treatment for osteoporosis. *Bone*. 2008;**42**(5):841-7.

22. Allen, M. R, Burr DB. The pathogenesis of bisphosphonate-related osteonecrosis of the jaw: so many hypotheses, so few data. *J Oral Maxillofac Surg* 2009; **67**(5):61-70.

23. Pazianas M, Miller P, Blumentals WA, Bernal M, Kothawala P. A review of the literature on osteonecrosis of the jaw in patients with osteoporosis treated with oral

bisphosphonates: prevalence, risk factors, and clinical characteristics. *Clin Ther* 2007; **29**(8):1548-58.

24. Coxon FP, Thompson K, Rogers MJ. Recent advances in understanding the mechanism of action of bisphosphonates. *Curr Opin Pharmacol*. 2006;**6**(3):307-12.

25. Hansen T, Kirkpatrick CJ, Walter C, Kunkel M. Increased numbers of osteoclasts expressing cysteine proteinase cathepsin K in patients with infected osteoradionecrosis and bisphosphonate-associated osteonecrosis-a paradoxical observation? *Virchows Arch*. 2006;**449**(4):448-54.

26. Maahs MP, Azambuja AA, Campos MM, Salum FG, Cherubini K. Association between bisphosphonates and jaw osteonecrosis: a study in Wistar rats. *Head Neck*. 2011; **33**(2):199-207.

27. Baroukh B, Saffar JL. Identification of osteoclasts and their mononuclear precursors. A comparative histological and histochemical study in hamster periodontitis. *J Periodontal Res.* 1991;**26**(3 Pt 1):161-6.

28. Eslami B, Zhou S, Van Eekeren I, LeBoff MS, Glowacki J. Reduced osteoclastogenesis and RANKL expression in marrow from women taking alendronate. *Calcif Tissue Int*. 2011;**88**(4):272-80.

29. Ito M, Amizuka N, Nakajima T, Ozawa H. Ultrastructural and cytochemical studies on cell death of osteoclasts induced by bisphosphonate treatment. *Bone*. 1999;**25**(4):447-52.

30. Fujita Y, Watanabe K, Uchikanbori S, Maki K. Effects of risedronate on cortical and trabecular bone of the mandible in glucocorticoid-treated growing rats. *Am J Orthod Dentofacial Orthop.* 2011;**139**(3):e267-77.

31. Krempien R, Huber PE, Harms W, Treiber M, Wannenmacher M, Krempien B. Combination of early bisphosphonate administration and irradiation leads to improved remineralization and restabilization of osteolytic bone metastases in an animal tumor model. *Cancer*. 2003;**98**(6):1318-24.

32. Ogawa K, Hori M, Takao R, Sakurada T. Effects of combined elcatonin and alendronate treatment on the architecture and strength of bone in ovariectomized rats. *J Bone Miner Metab.* 2005;**23**(5):351-8.

33. Spolidorio LC, Marcantonio E Jr, Spolidorio DM, Nassar CA, Nassar PO, Marcantonio RA, Rossa C Jr. Alendronate therapy in cyclosporine-induced alveolar bone loss in rats. *J Periodontal Res.* 2007;**42**(5):466-73.

34. Igarashi Y, Lee MY, Matsuzaki S. Acid phosphatases as markers of bone metabolism. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2002;**781**(1-2):345-58.

35. Price CP, Kirwan A, Vader C. Tartrate-resistant acid phosphatase as a marker of bone resorption. *Clin Chem.* 1995;**41**(5):641-3.

36. Chambers TJ, Fuller K, Darby JA. Hormonal regulation of acid phosphatase release by osteoclasts disaggregated from neonatal rat bone. *J Cell Physiol*. 1987;**132**(1):90-6.

37. Nicolin V, Bareggi R, Baldini G, Bortul R, Martinelli B, Narducci P. Effects of neridronic acid on osteoclasts derived by physiological dual-cell cultures. *Acta Histochem*. 2007;**109**(5):397-402.

38. Nishida S, Tsubaki M, Hoshino M, Namimatsu A, Uji H, Yoshioka S, et al. Nitrogencontaining bisphosphonate, YM529/ONO-5920 (a novel minodronic acid), inhibits RANKL expression in a cultured bone marrow stromal cell line ST2. *Biochem Biophys Res Commun.* 2005;**328**(1):91-7.

39. Fujimura Y, Kitaura H, Yoshimatsu M, Eguchi T, Kohara H, Morita Y, Yoshida N. Influence of bisphosphonates on orthodontic tooth movement in mice. *Eur J Orthod*. 2009; **31**(6):572-7.

40. Xiong H, Wei L, Hu Y, Zhang C, Peng B. Effect of alendronate on alveolar bone resorption and angiogenesis in rats with experimental periapical lesions. *Int Endod J*. 2010;43(6):485-91.

41. Kovacevic M, Tamarut T, Zoricic S, Beslic S. A method for histological enzyme histochemical and immunohistochemical analysis of periapical diseases on undercalcified bone with teeth. *Acta Estomat Croat*. 2003; **37**(3):269-73.

42. Pabst AM, Ziebart T, Koch FP, Taylor KY, Al-Nawas B, Walter C. The influence of bisphosphonates on viability, migration, and apoptosis of human oral keratinocytes-in vitro study. *Clin Oral Investig.* 2011; doi 10.1007/s00784-010-0507-6.

43. Scheper MA, Badros A, Chaisuparat R, Cullen KJ, Meiller TF. Effect of zoledronic acid on oral fibroblasts and epithelial cells: a potential mechanism of bisphosphonate-associated osteonecrosis. *Br J Haematol*. 2009;**144**(5):667-76.

44. Walter C, Pabst A, Ziebart T, Klein M, Al-Nawas B. Bisphosphonates affect migration ability and cell viability of HUVEC, fibroblasts and osteoblasts in vitro. *Oral Dis.* 2011;**17**(2):194-9.

45. Escudero ND, Lacave M, Ubios AM, Mandalunis PM. Effect of monosodium olpadronate on osteoclasts and megakaryocytes: an in vivo study. *J Musculoskelet Neuronal Interact.* 2009;**9**(2):109-20.

46. Labat-Moleur F, Guillermet C, Lorimier P, Robert C, Lantuejoul S, Brambilla E, et al. TUNEL apoptotic cell detection in tissue sections: critical evaluation and improvement. *J Histochem Cytochem.* 1998;**46**(3):327-34.

47. Cerri PS, Boabaid F, Katchburian E. Combined TUNEL and TRAP methods suggest that apoptotic bone cells are inside vacuoles of alveolar bone osteoclasts in young rats. J *Periodontal Res.* 2003;**38**(2):223-6.

48. Faloni AP, Sasso-Cerri E, Katchburian E, Cerri PS. Decrease in the number and apoptosis of alveolar bone osteoclasts in estrogen-treated rats. *J Periodontal Res.* 2007;**42**(3):193-201.

49. Benford HL, McGowan NW, Helfrich MH, Nuttall ME, Rogers MJ. Visualization of bisphosphonate-induced caspase-3 activity in apoptotic osteoclasts in vitro. *Bone*. 2001;**28**(5):465-73.

50. Chowdhury I, Tharakan B, Bhat GK. Caspases - an update. *Comp Biochem Physiol B Biochem Mol Biol.* 2008;**151**(1):10-27.

51. Chang HY, Yang X. Proteases for cell suicide: functions and regulation of caspases. *Microbiol Mol Biol Rev.* 2000;**64**(4):821-46.

52. Plotkin LI, Manolagas SC, Bellido T. Dissociation of the pro-apoptotic effects of bisphosphonates on osteoclasts from their anti-apoptotic effects on osteoblasts/osteocytes with novel analogs. *Bone*. 2006;**39**(3):443-52.

53. Hill PA. Bone remodelling. *Br J Orthod*. 1998;**25**(2):101-7.

54. Simonet WS, Lacey DL, Dunstan CR, Kelley M, Chang MS, Lüthy R, et al. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. *Cell*. 1997;**89**(2):309-19.

55. Aubin JE, Bonnelye E. Osteoprotegerin and its ligand: a new paradigm for regulation of osteoclastogenesis and bone resorption. *Osteoporos Int*. 2000;**11**(11):905-13.

56. Martini G, Gennari L, Merlotti D, Salvadori S, Franci MB, Campagna S, et al. Serum OPG and RANKL levels before and after intravenous bisphosphonate treatment in Paget's disease of bone. *Bone*. 2007;**40**(2):457-63.

57. Tipton DA, Seshul BA, Dabbous MKh. Effect of bisphosphonates on human gingival fibroblast production of mediators of osteoclastogenesis: RANKL, osteoprotegerin and interleukin-6. *J Periodontal Res.* 2011;**46**(1):39-47.

58. Teitelbaum SL. Bone resorption by osteoclasts. Science. 2000;289(5484):1504-8.

59. Trouvin AP, Goëb V. Receptor activator of nuclear factor- $\kappa$ B ligand and osteoprotegerin: maintaining the balance to prevent bone loss. *Clin Interv Aging*. 2010/**19**(5):345-54.

60. Khosla S. Minireview: the OPG/RANKL/RANK system. *Endocrinology*. 2001;**142**(12):5050-5.

61. Lacey DL, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, et al. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell*. 1998;**93**(2):165-76.

62. Alvarez L, Peris P, Guañabens N, Vidal S, Ros I, Pons F, et al. Serum osteoprotegerin and its ligand in Paget's disease of bone: relationship to disease activity and effect of treatment with bisphosphonates. *Arthritis Rheum*. 2003;**48**(3):824-8.

63. Mountzios G, Terpos E, Syrigos K, Papadimitriou C, Papadopoulos G, Bamias A, et al. Markers of bone remodeling and skeletal morbidity in patients with solid tumors metastatic to the skeleton receiving the bisphosphonate zoledronic acid. *Transl Res.* 2010;**155**(5):247-55.

64. Pan B, Farrugia AN, To LB, Findlay DM, Green J, Lynch K, Zannettino AC. The nitrogen-containing bisphosphonate, zoledronic acid, influences RANKL expression in human osteoblast-like cells by activating TNF-alpha converting enzyme (TACE). *J Bone Miner Res.* 2004;**19**(1):147-54.

65. Chenu C, Colucci S, Grano M, Zigrino P, Barattolo R, Zambonin G, et al. Osteocalcin induces chemotaxis, secretion of matrix proteins, and calcium-mediated intracellular signaling in human osteoclast-like cells. *J Cell Biol*.1994;**127**(4):1149-58.

66. Coleman R, Brown J, Terpos E, Lipton A, Smith MR, Cook R, et al. Bone markers and their prognostic value in metastatic bone disease: clinical evidence and future directions. *Cancer Treat Rev.* 2008;**34**(7):629-39.

67. Seibel MJ. Clinical use of markers of bone turnover in metastatic bone disease. *Nat Clin Pract Oncol.* 2005;**2**(10):504-17.

68. Brown JE, Sim S. Evolving role of bone biomarkers in castration-resistant prostate cancer. *Neoplasia*. 2010;**12**(9):685-96.

69. Aström E, Magnusson P, Eksborg S, Söderhäll S. Biochemical bone markers in the assessment and pamidronate treatment of children and adolescents with osteogenesis imperfecta. Acta Paediatr. 2010;99(12):1834-40.

70. Tsuchimoto M, Azuma Y, Higuchi O, Sugimoto I, Hirata N, Kiyoki M, et al. Alendronate modulates osteogenesis of human osteoblastic cells in vitro. *Jpn J Pharmacol*. 1994;**66**(1):25-33.

71. Koch FP, Merkel C, Al-Nawas B, Smeets R, Ziebart T, Walter C, et al. Zoledronate, ibandronate and clodronate enhance osteoblast differentiation in a dose dependent manner - A quantitative in vitro gene expression analysis of Dlx5, Runx2, OCN, MSX1 and MSX2. *J Craniomaxillofac Surg.* 2010;doi:10.1016/j.jcms.2010.10.007.

72. Zohar R, Lee W, Arora P, Cheifetz S, McCulloch C, Sodek J. Single cell analysis of intracellular osteopontin in osteogenic cultures of fetal rat calvarial cells. *J Cell Physiol*. 1997;**170**(1):88-100.

73. Sodek J, Ganss B, McKee MD. Osteopontin. *Crit Rev Oral Biol Med.* 2000;**11**(3):279-303.

74. Chun YH, Foster BL, Lukasavage PA, Berry JE, Zhao M, Tenenbaum HC, Somerman MJ. Bisphosphonate modulates cementoblast behavior in vitro. *J Periodontol*. 2005;**76**(11):1890-900.

75. Lekic P, Rubbino I, Krasnoshtein F, Cheifetz S, McCulloch CA, Tenenbaum H. Bisphosphonate modulates proliferation and differentiation of rat periodontal ligament cells during wound healing. *Anat Rec*.1997;**247**(3):329-40.

76. Blyth K, Vaillant F, Jenkins A, McDonald L, Pringle MA, Huser C, et al. Runx2 in normal tissues and cancer cells: A developing story. *Blood Cells Mol Dis*. 2010; **45**(2):117-23.

77. Hu H, Djuretic I, Sundrud MS, Rao A. Transcriptional partners in regulatory T cells: Foxp3, Runx and NFAT. *Trends Immunol*. 2007;**28**(8):329-32.

78. Corrado A, Neve A, Maruotti N, Gaudio A, Marucci A, Cantatore FP. Dose-dependent metabolic effect of zoledronate on primary human osteoblastic cell cultures. *Clin Exp Rheumatol.* 2010;**28**(6):873-9.

79. Enjuanes A, Ruiz-Gaspà S, Peris P, Ozalla D, Álvarez L, Combalia A, et al. The effect of the alendronate on OPG/RANKL system in differentiated primary human osteoblasts. *Endocrine*. 2010;**37**(1):180-6.

80. Muñoz-Torres M, Reyes-García R, Mezquita-Raya P, Fernández-García D, Alonso G, Luna Jde D, et al. Serum cathepsin K as a marker of bone metabolism in postmenopausal women treated with alendronate. *Maturitas*. 2009;**64**(3):188-92.

81. Herrmann M, Seibel MJ. The amino- and carboxyterminal cross-linked telopeptides of collagen type I, NTX-I and CTX-I: a comparative review. *Clin Chim Acta*. 2008;**393**(2):57-75.

82. Kawana K, Takahashi M, Hoshino H, Kushida K. Comparison of serum and urinary C-terminal telopeptide of type I collagen in aging, menopause and osteoporosis. *Clin Chim Acta*. 2002;**316**(1-2):109-15.

83. Brown JE, McCloskey EV, Dewar JA, Body JJ, Cameron DA, Harnett AN, et al. The use of bone markers in a 6-week study to assess the efficacy of oral clodronate in patients with metastatic bone disease. *Calcif Tissue Int*. 2007;**81**(5):341-51.

84. Orwoll ES, Miller PD, Adachi JD, Brown J, Adler RA, Kendler D, et al. Efficacy and safety of a once-yearly i.v. Infusion of zoledronic acid 5 mg versus a once-weekly 70 mg oral alendronate in the treatment of male osteoporosis: a randomized, multicenter, double-blind, active-controlled study. *J Bone Miner Res.* 2010;**25**(10):2239-50.

85. Nakaya H, Osawa G, Iwasaki N, Cochran DL, Kamoi K, Oates TW. Effects of bisphosphonate on matrix metalloproteinase enzymes in human periodontal ligament cells. *J Periodontol*. 2000;**71**(7):1158-66.

86. Buduneli E, Vardar-Sengül S, Buduneli N, Atilla G, Wahlgren J, Sorsa T. Matrix metalloproteinases, tissue inhibitor of matrix metalloproteinase-1, and laminin-5 gamma2 chain immunolocalization in gingival tissue of endotoxin-induced periodontitis in rats: effects of low-dose doxycycline and alendronate. *J Periodontol.* 2007;**78**(1):127-34.

87. Facchini G, Caraglia M, Morabito A, Marra M, Piccirillo MC, Bochicchio AM, et al. Metronomic administration of zoledronic acid and taxotere combination in castration resistant prostate cancer patients: phase I ZANTE trial. *Cancer Biol Ther*. 2010;**10**(6):543-8.

88. von Knoch F, Jaquiery C, Kowalsky M, Schaeren S, Alabre C, Martin I, Rubash HE, Shanbhag AS. Effects of bisphosphonates on proliferation and osteoblast differentiation of human bone marrow stromal cells. *Biomaterials*. 2005;**26**(34):6941-9.

89. Wehrhan F, Hyckel P, Ries J, Stockmann P, Nkenke E, Schlegel KA, et al. Expression of Msx-1 is suppressed in bisphosphonate associated osteonecrosis related jaw tissueetiopathology considerations respecting jaw developmental biology-related unique features. *J Transl Med.* 2010;**8**(96):1-9.

90. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. Nat Med. 2003;9(6):669-76.

91. Ferrara N.Vascular endothelial growth factor: basic science and clinical progress. *Endocr Rev.* 2004;**25**(4):581-611.

92. Amir E, Trinkaus M, Simmons CE, Dranitsaris G, Clemons MJ.Vascular endothelial growth factor activity after switching of bisphosphonate treatment for metastatic breast cancer. *J Clin Pathol*. 2009;**62**(5):474-6.

93. Tas F, Duranyildiz D, Oguz H, Camlica H, Yasasever V, Topuz E. Effect of zoledronic acid on serum angiogenic factors in patients with bone metastases. *Med Oncol.* 2008;**25**(3):346-9.

94. Santini D, Vincenzi B, Galluzzo S, Battistoni F, Rocci L, Venditti O, et al. Repeated intermittent low-dose therapy with zoledronic acid induces an early, sustained, and long-lasting decrease of peripheral vascular endothelial growth factor levels in cancer patients. *Clin Cancer Res.* 2007;**13**(15):4482-6.

95. Zhao X, Xu X, Guo L, Ragaz J, Guo H, Wu J, et al. Biomarker alterations with metronomic use of low-dose zoledronic acid for breast cancer patients with bone metastases and potential clinical significance. *Breast Cancer Res Treat*. 2010;**124**(3):733-43.

96. Wypij JM, Fan TM, Fredrickson RL, Barger AM, de Lorimier LP, Charney SC. In vivo and in vitro efficacy of zoledronate for treating oral squamous cell carcinoma in cats. *J Vet Intern Med*. 2008;**22**(1):158-63.

97. Siena S, Bregni M, Brando B, Belli N, Ravagnani F, Gandola L, et al. Flow cytometry for clinical estimation of circulating hematopoietic progenitors for autologous transplantation in cancer patients. *Blood*. 1991;**77**(2):400-9.

98. Tuomela JM, Valta MP, Väänänen K, Härkönen PL. Alendronate decreases orthotopic PC-3 prostate tumor growth and metastasis to prostate-draining lymph nodes in nude mice. BMC *Cancer*. 2008;**8**(81):1-12.

99. Allegra A, Oteri G, Nastro E, Alonci A, Bellomo G, Del Fabro V, Quartarone E, Alati C, De Ponte FS, Cicciù D, Musolino C. Patients with bisphosphonates-associated osteonecrosis of the jaw have reduced circulating endothelial cells. *Hematol Oncol.* 2007;**25**(4):164-9.

100. Yamashita J, Koi K, Yang DY, McCauley LK. Effect of zoledronate on oral wound healing in rats. *Clin Cancer Res.* 2011;**17**(6):1405-14.

101. Soltau J, Zirrgiebel U, Esser N, Schächtele C, Totzke F, Unger C, et al. Antitumoral and antiangiogenic efficacy of bisphosphonates in vitro and in a murine RENCA model. *Anticancer Res.* 2008;**28**(2A):933-41.

102. Zanetta L, Marcus SG, Vasile J, Dobryansky M, Cohen H, Eng K, Shamamian P, Mignatti P. Expression of von Willebrand factor, an endothelial cell marker, is upregulated by angiogenesis factors: a potential method for objective assessment of tumor angiogenesis. *Int J Cancer*. 2000;**85**(2):281-8.

103. Lip GY, Blann A. von Willebrand factor: a marker of endothelial dysfunction in vascular disorders? *Cardiovasc Res.* 1997;**34**(2):255-65.

104. Freitas TM, Miguel MC, Silveira EJ, Freitas RA, Galvão HC. Assessment of angiogenic markers in oral hemangiomas and pyogenic granulomas. *Exp Mol Pathol*. 2005;**79**(1):79-85.

105. Schor AM, Pendleton N, Pazouki S, Smither RL, Morris J, Lessan K, et al. Assessment of vascularity in histological sections: effects of methodology and value as an index of angiogenesis in breast tumours. *Histochem J*. 1998;**30**(12):849-56.

# ARTIGO 2

# 3 ARTIGO 2

O artigo a seguir intitula-se Comparison of effects of clodronate and zoledronic acid on the repair of maxilla surgical wounds - Histomorphometric, RANKL, OPG, von Willebrand factor and caspase-3 evaluation e foi formatado de acordo com as normas do periódico *Journal of Oral Pathology & Medicine* (Anexo B). Comparison of effects of clodronate and zoledronic acid on the repair of maxilla surgical wounds - Histomorphometric, RANKL, OPG, von Willebrand factor and caspase-3 evaluation

Ana Carolina Uchoa Vasconcelos<sup>1</sup> Soraya de Azambuja Berti Couto<sup>1</sup> Alan Arrieira Azambuja<sup>2</sup> Fernanda Gonçalves Salum<sup>3</sup> Maria Antonia Figueiredo<sup>3</sup> Vinícius Duval da Silva<sup>4</sup> Karen Cherubini<sup>3</sup>

<sup>1</sup> Ph.D. Student, Postgraduate Program, Dental College, Pontifical Catholic University of Rio Grande do Sul - PUCRS

<sup>2</sup> M.Sc., Oncology Department, Hospital São Lucas, Pontifical Catholic University of Rio Grande do Sul - PUCRS

<sup>3</sup>Ph.D., Postgraduate Program, Dental College, Pontifical Catholic University of Rio Grande do Sul - PUCRS

<sup>4</sup> Ph.D., Cytometry and Immunohistochemistry Laboratory, Pathology Department,
Hospital São Lucas, Medical College, Pontifical Catholic University of Rio Grande do Sul
PUCRS, Porto Alegre, RS, Brazil

# **Corresponding author**

Karen Cherubini Serviço de Estomatologia, Hospital São Lucas PUCRS Av Ipiranga, 6690, sala 231, Porto Alegre, RS, Brazil CEP 90610-000 Telephone/fax: 55(51)3320 3254 E-mail: karen.cherubini@pucrs.br

Running title: Bisphosphonates and surgical wounds

#### ABSTRACT

**BACKGROUND**. The aim of this study was to compare clodronate and zoledronic acid regarding their influence on the repair of surgical wounds in maxillae (soft tissue wound and tooth extraction) and their relation to osteonecrosis.

**MATERIAL AND METHODS.** Thirty-four Wistar rats were allocated into 3 groups according to the treatment received: (1) 12 animals treated with zoledronic acid; (2) 12 animals treated with clodronate; and (3) 10 animals that were given saline solution. All animals were subjected to tooth extractions and surgically induced soft tissue injury. Histological analysis of the wound sites was performed by means of hematoxylin-eosin (H&E) staining and immunohistochemical staining for RANKL, OPG, von Willebrand factor and caspase-3.

**RESULTS.** The zoledronic acid group showed higher incidence of non-vital bone than did the clodronate group at the tooth extraction site. At the soft tissue wound site, there were no significant differences in non-vital bone between the test groups. RANKL, OPG, von Willebrand factor and caspase-3 did not show significant differences between the groups for both sites of surgical procedures.

**CONCLUSION**. Both of the bisphosphonates zoledronic acid and clodronate are capable of inducing maxillary osteonecrosis. Immunohistochemical analysis suggests that the involvement of soft tissues as the initiator of osteonecrosis development is less probable than has been pointed out.

Key words: bisphosphonates; zoledronic acid; clodronate; osteonecrosis

#### Introduction

Bisphosphonates are classified as nitrogen- and non-nitrogen according, respectively, to the presence or absence of nitrogen in their chemical structure. These drugs are associated with osteonecrosis of the jaws, an important adverse effect first described in 2003. The disease is characterized by exposure of bone to the oral cavity that does not heal within eight weeks, in patients who have received bisphosphonates without history of radiotherapy in the head and neck region (1, 2). In some cases, the condition is refractory to treatment, where it is only possible to preserve the patient's quality of life by controlling pain and infection, as well as preventing the occurrence of new areas of necrosis (3).

Bisphosphonates inhibit bone resorption through direct and indirect effects on osteoclasts, which undergo apoptosis or become unable to differentiate from hematopoietic stem cells (4-6). Other effects of these drugs, such as impairment of both angiogenesis (7-11) and epithelial cell proliferation (12), support the hypothesis that osteonecrosis of the jaws has a multifactorial etiology.

The pathway of receptor activator of nuclear factor-kB ligand (RANKL) and osteoprotegerin (OPG) is one of the main regulators of the molecular mechanisms involved in the development and function of osteoclasts. Studies on bisphosphonates, in vitro and in vivo. have shown controversial results when these proteins used are as immunohistochemical, genetic and serum markers (13, 14). It was also demonstrated that nitrogen-containing bisphosphonates are able to cause toxicity to the cells of oral epithelium, raising concerns whether disease onset could occur in bone tissue or in oral mucosa (15, 16). Moreover, most studies evaluating the effects of the drug on body tissues have been performed with nitrogen-containing bisphosphonates, which generate many concerns about tissue behavior when non-nitrogen-containing ones are used (15, 17).

There are many case reports and some animal model studies in which the use of zoledronic acid is associated with jaw osteonecrosis. Nevertheless, even though clodronate is the most prescribed non-nitrogen-containing bisphosphonate, there are a few case-reports in the literature on osteonecrosis of the jaws induced by this drug (18). In some of them, patients had used nitrogen-containing before the treatment with non-nitrogen ones. Nor has there been a study about clodronate effects using animal models. The aim of this work was to compare the effects of clodronate and zoledronic acid on the repair of surgical wounds of maxillae. Microscopic features of tooth extraction and soft tissue injury areas were evaluated by means of hematoxylin-eosin (H&E) staining and immunohistochemical detection of RANKL, OPG, von Willebrand factor (vWF) and caspase-3.

# Material and methods

#### Animals

The present study was approved by the Ethics Committee for Animal Use of the Pontifical Catholic University of Rio Grande do Sul, and the procedures were carried out in accordance with institutional guidelines for animal care and use. The sample comprised 34 female rats (*Rattus norvegicus*, Wistar strain) from the animal facility of the Federal University of Pelotas, which had a mean age of 120 days and a mean weight of 230 g. Animals were individually identified on the tails and housed in plastic cages (5 per cage) placed in ventilated racks (Alesco, Monte Mor, SP, Brazil) at a temperature of 22°C with a 12-h light/dark cycle (lights on at 7:00 am and off at 7:00 pm). During the experiments, a standard diet of rat chow (Nuvilab, Colombo, PR, Brazil) and filtered water were provided *ad libitum*. The cleaning and changing of the cages were done three times a week. No experimental procedures were carried out in the place where the animals were kept in order to avoid any type of stress behavior. The animals were randomly allocated into 3 groups,

according to the bisphosphonate used: (1) zoledronic acid group: 12 animals that were treated with the nitrogen-containing bisphosphonate zoledronic acid (Novartis Pharma AG, Basel, Switzerland) intraperitoneally (0.6 mg/kg, every 28 days); (2) clodronate group: 12 animals that were treated with the non-nitrogen-containing bisphosphonate clodronate (Jenahexal Pharma GmbH, Thuringia, Germany), intraperitoneally (20 mg/kg, every 28 days); and (3) control group: 10 animals that were given saline solution (0.9% sodium chloride), intraperitoneally every 28 days.

#### Surgical procedures

All animals were subjected to tooth extractions and surgical-induced soft tissue injury as described below. Oroscopy was performed after the anesthesia and before the surgical procedures to certify that there were no previous oral lesions. The animals were given intraperitoneal paracetamol at a dose of 50 mg/kg (Medley S/A, Campinas, SP, Brazil) after the surgery for postoperative analgesia.

## Tooth extractions

Tooth extractions were performed 60 days after the beginning of the experiment. Animals were anesthetized with a single intraperitoneal injection of a mixture of ketamine hydrochloride 5% (100 mg/kg; Vetbrands, Jacareí, SP, Brazil) and xylazine hydrochloride 2% (10 mg/kg; Vetbrands, Jacareí, SP, Brazil). The three upper right molars were extracted using an adapted 3s spatula (SSWhite, Duflex, Rio de Janeiro, RJ, Brazil) for luxation and a pediatric forceps (Edlo, Canoas, RS, Brazil) whose functional portion was adapted to the size of the tooth.

#### Surgically induced soft tissue wound

Immediately after the tooth extractions, a surgical wound was made on the mucosa of the hard palate at the opposite side (left side) of the maxilla, with reference to the second left upper molar, and using a surgical scalpel with no. 3 Bard-Parker handle (Solidor, São Paulo, SP, Brazil) and a no. 15 blade (Solidor, São Paulo, SP, Brazil). The incision was elliptical, 3 mm long and 1 mm deep. The size of the lesion was monitored by a millimeter ruler guide and a periodontal probe.

#### Euthanasia of the animals, macroscopic evaluation and dissection of the maxillae

After completing 102 days of drug administration, the animals were euthanized by deep anesthesia with isoflurane (Cristalia, Porto Alegre, RS, Brazil) in an appropriate anesthesia chamber. The specimens were then examined by means of a no.5 clinical probe (SS White, Duflex, Rio de Janeiro, RJ, Brazil) to determine the presence/absence of oral mucosal lesion. Afterwards, the maxillae were dissected and fixed for 24 h in 10% buffered formalin (TopGlass, Porto Alegre, RS, Brazil).

## Histological processing

The specimen was cut into two fragments in the coronal direction, using a steel-sanding disc at low speed and subjected to decalcification in ethylenediaminetetraacetic acid (EDTA, Biodinâmica, Ibiporã, PR, Brazil) solution for 30 days. Next, they were paraffinembedded, cut into 4-µm sections and stained with hematoxylin and eosin (H&E) as well as immunohistochemically processed.

Antigen retrieval was carried out in a 100°C water bath for 40 min, using Tris/EDTA buffer, pH 9 (20 mM Tris/0.65 mM EDTA). Endogenous peroxidase was

blocked with a 3% solution of hydrogen peroxide in methanol for 30 min, followed by three cycles of washing with PBS. The blocking of nonspecific binding was done with the commercial solution Protein Block Serum-Free (Dako, Carpinteria, CA, USA) for 30 min at room temperature. The immunohistochemical staining method based on capillary action was used (Thermo, Shandon, CA, USA), where the sections were incubated overnight at 2°C with dilutions of the following antibodies: anti-RANKL (Santa Cruz Biotechnology, Santa Cruz, CA, USA) diluted at 1:500; anti-OPG goat polyclonal antibody (SC8468 -Santa Cruz Biotechnology, Santa Cruz, CA, USA) at 1:100; anti-vWF (Biocare, Concord, Massachusetts, USA) and anti-caspase-3 (Novocastra, Newcastle, UK) diluted at 1:500. After incubation with the primary antibody, sections were washed with 3 passages in PBS. For amplification of the antigen-antibody reaction, the Picture Max system (Invitrogen, Carlsbad, CA, USA) was used according to manufacturer's recommendations. The slides were then washed with PBS and incubated with diaminobenzidine solution for 5 min. The detection system used was Dako LSAB Kit (Dako, Carpinteria, CA, USA). Color development was carried out with the chromogen 3'-diaminoabenzidine and phosphate buffer solution containing 0.002% hydrogen peroxide. All markers had external and internal controls.

# Histological analysis

The histological sections were digitized using a Zeiss Axioskop 40 light microscope (Zeiss, Göettingen, Germany), connected to a Roper Scientific video camera (Media Cybernetics, Bethesda, MD, USA) and a Pentium IV 2.2 GHZ computer with 512 MB RAM, 160 GB hard drive, and Image Pro Capture Kit Platform (Media Cybernetics). The images were captured using 5x (H&E stain) and 10x (immunohistochemistry) objectives and stored in Joint Photographic Expert Group (JPEG) format. For H&E analysis, 5 fields

were selected in a standardized manner in each slide to include the whole area of tooth extractions and soft tissue wound. For immunohistochemistry, 2 fields were selected in a standardized manner to include the epithelial and connective tissue at both sites evaluated.

The images were analyzed by a calibrated and blinded examiner. The calibration consisted of evaluating a series of 20 histological images for each technique (H&E and immunohistochemistry) twice, at two different moments. The results of these two evaluations were subjected to a paired *t* test and Pearson's correlation coefficient, showing the absence of a significant difference (p > 0.05) and a strong correlation (r > 0.8).

#### H&E analysis

The H&E images were analyzed using the manual counting technique in the Image Pro Plus 4.5.1 software (Media Cybernetics, Bethesda, MD, USA). A quantitative analysis was made for the variables epithelial tissue, connective tissue, root fragments, microbial colonies, inflammatory infiltrate, non-vital bone, and vital bone. A point-grid of 532 points was superimposed to each image and each point was counted according to the matching morphological structure (Fig. 1).



**Figure 1**. Quantification of histological features in 1 of the 3 fields examined at the site of tooth extraction using Image Pro Plus 4.5.1 software (Media Cybernetics, Bethesda, MD, USA; H&E stain, x 5 objective).

## Immunohistochemical analysis

The immunohistochemical expression of RANKL, OPG, vWF and caspase-3 was quantified, according to Amenábar et al. (19) by means of semi-automated segmentation technique in the Image Pro Plus 4.5.1 software (Media Cybernetics, Bethesda, MD, USA; Fig. 2).



**Figure 2**. Quantification of immunohistochemical expression of OPG by means of semi-automated segmentation technique using Image Pro Plus 4.5.1 software (Media Cybernetics, Bethesda, MD, USA; x10 objective)

### Statistical analysis

The data were analyzed by descriptive statistics, chi-squared test complemented by analysis of adjusted residuals, and Kruskal-Wallis test complemented by its multiple comparisons test, setting the level of significance at 5%. The statistics were processed by the SPSS 17.0 software (Statistical Package for the Social Sciences, Chicago, IL, USA).

# Results

# Clinical and macroscopic evaluation

On oral examination before tooth extractions, no animal exhibited oral mucosal lesions. Table 1 presents the results of macroscopic evaluation (after euthanasia) for tooth extraction and soft tissue wound sites. Zoledronic acid was associated with the loss of mucosal integrity in the tooth extraction site (chi-squared test complemented by analysis of adjusted residuals, p < 0.001). Neither zoledronic acid nor clodronate was associated with loss of mucosal integrity at the soft tissue wound site (chi-squared test, p=0.151).

	Loss of mucosal integrity									
		Tooth extr	action site		Soft tissue wound site					
Group	Pre	esent	At	osent	Pre	sent	Absent			
	n	%	n	%	n	%	n	%		
Zoledronic acid (n=12)	12*	100*	0	0	9	75	3	25		
Clodronate (n=12)	10	83.3	2	16.7	9	75	3	25		
Control (n=10)	1	10	9	90	4	40	6	60		
Total	23	67.6	11	32.3	22	64.7	12	35.2		

**Table 1.** Sample distribution according to presence/absence of loss of mucosal integrity on macroscopic examination of the tooth extraction site and soft tissue wound site

n = number of animals

\*Significant difference; chi-square test; analysis of adjusted residuals; p<0.001

#### H&E Analysis

Table 2 shows the results for the frequency (presence/absence) of non-vital bone in the groups evaluated. By means of the chi-squared test, complemented by analysis of adjusted residuals, it was observed that (1) zoledronic acid was associated with non-vital bone at the tooth extraction site (p<0.001); (2) at the soft tissue wound site, both zoledronic acid and clodronate were associated with non-vital bone (p<0.001).

	Non-vital bone									
		Tooth extr	action site	Soft tissue wound site						
Group	Pre	esent	At	osent	Pre	esent	Absent			
	n	%	n	%	n	%	n	%		
Zoledronic acid (n=12)	12*	100*	0	0	12*	100*	0	0		
Clodronate (n=12)	5	41.6	7	58.3	12*	100*	0	0		
Control (n=10)	0	0	10	100	4	40	6	60		

**Table 2**. Sample distribution according to presence/absence of non-vital bone at the tooth extraction and soft tissue wound sites on microscopic examination

n = number of animals

\*Significant difference; chi-square test; analysis of adjusted residuals; p<0.001

The results for the proportions of the histological variables are presented in Tables 3 and 4, respectively, for the tooth extractions and soft tissue wound sites. At the tooth extraction site, the proportion of non-vital bone and microbial colonies was significantly greater in the zoledronic acid group compared to the clodronate and control groups, but the latter two did not differ significantly from each other. There was no significant difference in vital bone and root fragments between the groups. The proportion of connective tissue was significantly greater in the clodronate compared to the zoledronic acid group, but they did not differ significantly from the control group (Table 3, Kruskal-Wallis test complemented by multiple comparisons test,  $\alpha = 0.05$ ).

At the soft tissue wound site, the proportions of nonvital bone and microbial colonies were significantly greater in the zoledronic acid and clodronate groups than in the control. The proportion of connective tissue was significantly greater in the control group compared to the clodronate and zoledronic acid groups, whereas the proportion of vital bone was significantly greater in the control group compared to the clodronate group, but these two did not differ significantly from the zoledronic acid group. There was no significant difference in epithelial tissue and inflammatory infiltrate between the groups

(Table 4, Kruskal-Wallis test complemented by multiple comparisons test,  $\alpha = 0.05$ , Fig.3-A-D).

Table 3. Quantification of histological features (H&E stain) at the tooth extraction site in the zoledronic
acid, clodronate and control groups

	Group										
Histological	Zoledronic acid (%)			Cl	odronate	(%)	Control (%)				
feature	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median		
Non-vital bone	15.38	10.02	13.78 <sup>A</sup>	1.28	2.12	0 <sup>B</sup>	0	0	0 <sup>B</sup>		
Epithelial tissue	18.74	5.90	18.37	16.20	3.46	16.73	18.47	4.70	18.82		
Connective tissue	28.13	8.23	30.44 <sup>B</sup>	41.73	10.44	42.42 <sup>A</sup>	37.48	8.68	<b>34.49</b> <sup>AB</sup>		
Vital bone	16.39	13.84	13.42	24.97	15.74	20.58	31.13	14.63	31.33		
Inflammatory infiltrate	8.04	6.36	5.74	7.97	7.57	5.37	8.41	7.88	6.17		
Microbial colonies	3.46	2.77	3.32 <sup>A</sup>	0.49	0.60	0.21 <sup>B</sup>	0.38	0.72	0.08 <sup>B</sup>		
Root fragments	9.83	7.89	8.37	7.33	5.04	7.97	3.40	4.79	0.79		

\* Bold printed medians, followed by different letters, indicate features that differed significantly between groups; Kruskal-Wallis test complemented by multiple comparisons test,  $p \le 0.05$ 

SD = Standard deviation

	Group									
Histological	Zoledronic acid (%)			Clo	odronate	(%)	Control (%)			
feature	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median	
Non-vital bone	31.35	8.89	<b>30.79</b> <sup>A</sup>	21.99	6.55	23.26 <sup>A</sup>	3.65	5.61	0 <sup>B</sup>	
Epithelial tissue	12.21	3.65	11.44	13.96	5.6	13.11	12.62	2.10	12.98	
Connective tissue	31.22	5.95	30.43 <sup>C</sup>	41.03	9.10	43.99 <sup>B</sup>	52.11	9.33	51.96 <sup>A</sup>	
Vital bone	11.49	8.73	11.32 <sup>AB</sup>	3.73	3.57	3.51 <sup>B</sup>	22.95	16.44	27.77 <sup>A</sup>	
Inflammatory infiltrate	10.12	8.87	8.47	16.33	7.67	15.89	8.00	7.49	8.92	
Microbial colonies	3.58	3.34	2.35 <sup>A</sup>	2.94	2.60	2.29 <sup>A</sup>	0.34	0.85	0 <sup>B</sup>	

**Table 4**. Quantification of histological features (H&E stain) at the soft tissue wound site in the zoledronic acid, clodronate and control groups.

\*\* Bold printed medians, followed by different letters, indicate features that differed significantly between groups; Kruskal-Wallis test complemented by multiple comparisons test,  $p \le 0.05$  SD = Standard deviation

# Immunohistochemical analysis

Immunohistochemical quantification for RANKL, OPG, vWF and caspase-3 at tooth extraction and soft tissue wound sites did not differ significantly between the three groups analyzed (Tables 5 and 6, Kruskal-Wallis test,  $\alpha$ =0.05, Fig.3-D, E).

caspase-3 at tooth extraction site in the zoledronic acid, crodronate and control groups											
	Group										
	Zoledronic acid (mm <sup>2</sup> )			Clodronate (mm <sup>2</sup> )			Co				
Marker	Mean	SD	MD	Mean	SD	MD	Mean	SD	MD	<i>P</i> *	
RANKL	0.38	0.32	0.35	0.34	0.38	0.25	0.42	0.44	0.38	0.76	
OPG	0.31	0.30	0.22	0.35	0.20	0.31	0.26	0.20	0.21	0.55	
vWF	0.50	0.35	0.39	0.77	0.61	0.58	0.49	0.27	0.41	0.58	
Caspase 3	1.72	2.11	1.10	1.03	0.97	0.89	1.33	1.15	1.11	0.74	

**Table 5**. Immunohistochemical quantification of RANKL, OPG, von Willebrand factor (vWF) and caspase-3 at tooth extraction site in the zoledronic acid, clodronate and control groups

\*Kruskal-Wallis test,  $\alpha$ =0.05

SD = Standard deviation; MD = Median

Table 6. Immunohistochemical quantification of OPG, RANKL, von Willebrand factor (vWF) and
caspase-3 at soft tissue wound site in the zoledronic acid, clodronate and control groups

	Group											
	Zoledr	onic acid	$(mm^2)$	Clod	ronate (1	nm <sup>2</sup> )	Control (mm <sup>2</sup> )					
Marker	Mean	SD	MD	Mean	SD	MD	Mean	SD	MD	<i>P</i> *		
RANKL	0.35	0.40	0.13	0.41	0.34	0.32	0.52	0.55	0.30	0.77		
OPG	0.36	0.33	0.30	0.29	0.26	0.16	0.59	0.32	0.71	0.08		
vWF	0.97	0.93	0.67	0.61	0.58	0.30	0.79	0.71	0.60	0.46		
Caspase 3	1.41	1.18	1.27	1.52	2.20	0.82	1.30	0.92	1.14	0.75		

\*Kruskal-Wallis test,  $\alpha$ =0.05

SD = Standard deviation; MD = Median



**Figure 3.** Persistent soft tissue defect showing the disruption of epithelium (H&E stain, original magnification x 100) (A); Non-vital bone, microbial colonies, and inflammatory infiltrate (H&E stain, original magnification x 200) (B); Complete tissue repair (H&E stain, original magnification x 100) (C); Vital bone (H&E stain, original magnification x 400) (D); Immunostaining of RANKL (original magnification x 200) (E); Immunostaining of OPG (original magnification x 400) (F).

# Discussion

No animal in the three groups evaluated showed oral mucosal lesion prior to the surgical procedures, which indicates the lesions observed afterwards were associated with the

surgical procedure and bisphosphonate use. On macroscopic examination, an association was observed between loss of mucosal integrity at the tooth extraction site and zoledronic acid use. The occurrence of osteonecrosis and root fragments seems to explain this finding in both test groups and control. At the soft wound site, however, this association was not observed in any group, even with all animals in the test groups showing non-vital bone. This finding is in agreement with reports suggesting that bisphosphonate-associated jaw osteonecrosis can occur without bone exposure (20-22). Still, it disagrees with the hypothesis that soft tissues could occupy the first position in the pathophysiology of osteonecrosis (23-26).

The frequency of non-vital bone was significantly higher at the soft tissue wound site than at the tooth extraction site. The anatomical specificities of the sites subjected to the surgical procedures, as well as the type of lesion induced, could have contributed to this result. After a tooth extraction, the alveolar socket is first filled with the clot, which gives rise to a neoformed connective tissue, rich in fibroblasts and capillaries, which in turn promotes healing through the formation of well-organized bone trabeculae (27). The buccal and lingual bone walls remain covered by mucosa, healing occurs by secondary intention (28), and growing epithelial cells restore the epithelial continuity of the mucosa (29). In the case of bisphosphonate users, this process would be impaired by diminished vascularization (7) and reduced bone neoformation (16). Moreover, the hard palate has poor vascularization and its mucosa is firmly adhered to the periosteum (30). The wound at this site seems to have the potential of exposing a wider area of bone tissue, with less blood supply, if compared to the tooth extraction site, which could explain its higher prevalence of osteonecrosis. Therefore, the results of non-vital bone for the soft tissue wound site suggest that oral mucosal lesions, depending on their location, vascularization, submucosal thickness and relationship with the subjacent bone tissue, constitute a sufficient risk factor
for the occurrence of osteonecrosis associated with bisphosphonates, with no necessity of more invasive interventions such as tooth extractions. This is true not only for nitrogen-containing bisphosphonates, but also to non-nitrogen ones.

Nevertheless, it is important to recall that the maxilla, the bone subjected to tooth extractions in the present study, because of its higher vascularization (31) and turnover, is less prone to bisphosphonate osteonecrosis compared to the mandible is (32). Thus, it is plausible to infer that if we had done the tooth extractions in the mandible instead of maxilla, the prevalence of osteonecrosis could have been higher at this wound site than at the soft tissue wound site.

On microscopic examination, the tooth extraction site showed non-vital bone in 100% of the animals in the zoledronic acid group. This finding is in agreement with other studies, according to which the trauma constitutes a sufficient risk factor for osteonecrosis in zoledronic acid users (31, 33, 34). On the other hand, at the same site, the clodronate group did not show an association with non-vital bone, even though 5 (41.6%) out of 12 animals showed this feature. In humans, the lower prevalence of osteonecrosis associated with non-nitrogen bisphosphonates, especially clodronate, could be explained by the fact that these compounds have lower potency, and are less prescribed than nitrogen-containing ones (35). Besides that, the duration of non-nitrogen-containing bisphosphonates use needed for the occurrence of osteonecrosis is significantly longer (18). Nonetheless, considering that there was an association between non-vital bone and clodronate at the soft tissue wound site, it is also plausible to infer that a larger sample size would have given us a different result for the tooth extraction site in the clodronate group.

At the tooth extraction site, the zoledronic acid group had a higher proportion of microbial colonies than the clodronate and control. This is in accordance with the higher proportion of non-vital bone found in the zoledronic acid group, since osteonecrosis is characteristically accompanied by microbial infection, especially by *Actinomyces* sp. (36). Nevertheless, there are reports on the antimicrobial effect of clodronate on *Staphylococcus aureus* and *Pseudomonas aeruginosa* (37). Zoledronic acid effects on microorganisms are also reported, although in an opposite direction, as this drug has been shown to improve, *in vitro*, the adhesion of *S. mutans* to the bone hydroxyapatite, promoting bacterial growth in culture dishes (12). Therefore, the greater occurrence of microbial colonies in the zoledronic acid group when compared to clodronate could be related to the specific interactions of each one of these two bisphosphonates with microorganisms.

At the site of soft tissue wound, the clodronate and zoledronic acid groups showed a lower proportion of fibrous connective tissue when compared to the control group. These findings are in agreement with studies showing that bisphosphonates are able to decrease fibroblast proliferation (38). Curiously, at this same site, vital bone proportion was significantly lower in the clodronate than in the control group, whereas the latter and the zoledronic acid group did not differ from each other. It is known that bisphosphonates promote nearly complete suppression of bone turnover (39, 40), but in this case we do not know why clodronate had a greater effect than did zoledronic acid.

The immunohistochemical quantification of RANKL, OPG, vWF and caspase-3 did not show significant differences between the groups evaluated, either at the soft tissue wound site or the tooth extraction site. The bisphosphonate effects on the epithelial cells of the oral mucosa are poorly understood, and until now, it has not been determined if osteonecrosis starts in bone tissue or in oral mucosa (15, 25, 38).

The present study quantified caspase-3 immunohistochemical expression in epithelial and connective tissues aiming to determine if higher rates of apoptosis in these tissues could favor osteonecrosis development. The results are in accordance with the reports of no increase in caspase-3 expression in oral keratinocytes and gingival fibroblasts treated with nitrogen-containing bisphosphonates (15, 38). The lack of a difference in caspase-3 expression between the groups suggests that if the soft tissues are the target of the onset of osteonecrois (41), this involvement is not related to increased apoptosis of these cells. Moreover, studies report some bisphosphonate effects on keratinocytes and fibroblasts, such as loss of cell adhesion and decreased cell migration and proliferation (15, 38), relating them to the ability of promoting necrosis. Therefore, considering the hypothesis of bisphosphonate-related-osteonecrosis starting in soft tissue, it is possible that cell necrosis and/or impairment of critical events to oral wound healing such as cell proliferation, migration and differentiation would be the factor responsible for the lesion (15, 38).

The lack of significant differences in vWF expression between the groups evaluated in this study suggests that bisphosphonates are not capable of inhibiting vascularization in oral soft tissues. This agrees with the reports of Wehrhan et al. (42), who found that the immunohistochemical expression of CD31 in mucoperiosteal tissue did not differ between bisphosphonate users with and without osteonecrosis. However, studies investigating the bisphosphonate effects on angiogenesis show conflicting results.

According to some *in vitro* studies, zoledronic acid inhibits human endothelial cell differentiation (43), reduces their proliferation (44, 45), induces their apoptosis (11) and decreases the formation of capillary tubes (43). Similarly, other *in vitro* studies report that clodronate inhibits endothelial cell proliferation (45, 46). On the other hand, immunohistochemical expression of VEGF in bone tissue at the tooth extraction site did not differ significantly between rats treated with bisphosphonates and controls (33). There are also reports of a significant reduction in serum levels of VEGF in cancer patients treated with nitrogen-containing and non-nitrogen-containing bisphosphonates (47, 48).

The disagreements between the studies, either *in vitro* or *in vivo*, reflect the difficulties in comparing them because of the different methods applied.

vWF, whose expression was examined in the present work, is an important marker of vascularization, but it does not differentiate new vessels from preexisting ones (49). One could infer that by using a marker capable of indentifying new vessels, such as VEGF, we could have found a different result indicating an effect of bisphoshonate on angiogenesis. Nevertheless, we should consider the phase of healing in which the immunohistochemical analysis was performed here. In rats, the healing process at tooth extraction sites is completed by 40 days and neoformed vessels should be detected at 7 days (27), but not at the time at which we performed the evaluation. At this moment, the differentiation between new and preexisting vessels does not seem to influence the results anymore. Considering the conditions of the present study, as well as the conflicting results with VEGF and also the fact that vWF immunostaining pattern is more uniform if compared with VEGF and CD31 (50), we chose vWF as the marker of vascularization in our study.

The lack of significant differences in the immunohistochemical expression of OPG and RANKL between the zoledronic acid, clodronate and control groups would suggest that bisphosphonates are not capable of increasing the OPG/RANKL ratio. In our study, these proteins, which are also expressed in endothelial cells and fibroblasts, were quantified in connective tissue, 6 weeks after the surgical procedures, corresponding to the time of complete bone formation (51). This site of evaluation was chosen because we wanted to analyze the alterations in soft tissue that could be related to the onset of osteonecrosis and also because connective tissue is the matrix of bone neoformation. At this site, growth factors, cytokines and prostaglandins exert their effects on the recruitment, replication and differentiation of bone precursor cells (52). It is possible that an evaluation at the time corresponding to the highest metabolic activity could have shown different results, that is, at one week after tooth extraction, when there are important changes in the connective tissue. Also, it is worth pointing out that at the soft tissue wound site, the statistical comparison of OPG results between groups showed p=0.08, which suggests that a larger sample size could have shown a significant result.

It is also important to consider that most of the *in vivo* studies evaluating the effects of bisphosphonates on the behavior of these proteins have measured their serum levels (53, 54). Maybe this analysis does not represent what actually occurs in the environment of bone cells, oral keratinocytes and fibroblasts (54). Moreover, we have to consider that the serum levels of RANKL reflect only its soluble form, excluding the major mediator of osteoclastogenesis, which is its form as a cytokine of cell membrane (55). Studies comparing the expression of these two forms of RANKL would be helpful.

It is worth recalling that the immunohistochemical analysis was conducted in epithelial and connective tissues, not in bone tissue. Considering this point, the absence of any significant result for the markers used (vWF, caspase-3, OPG, RANKL) in the test groups suggests that the major bisphosphonate effects occur in bone tissue. The high bone affinity of these drugs (40) corroborates this idea. According to the literature, 70% of the zoledronic acid administered binds to hydroxyapatite crystals, whereas the rest of the dose is eliminated unaltered by renal excretion (56). The situation with clodronate is similar except for the fact that the absence of the hydroxyl group in this compound reduces its ability to bind to hydroxyapatite crystals, compared to zoledronic acid (57).

In conclusion, both bisphosphonates zoledronic acid and clodronate are capable of inducing maxillary osteonecrosis. The findings of immunohistochemical analysis suggest that the involvement of soft tissues as the initiator of osteonecrosis development is less probable than has been pointed out. To elucidate this point, further studies should focus on growth factors such as insulin-like growth factor and transforming growth factor-b, critical to oral wound healing, by means of molecular biology techniques such as PCR.

We thank the Novartis Laboratory for the donation of zoledronic acid and Dr. Fernanda Morrone, Head of Laboratório Farmacologia Aplicada, Faculdade de Farmácia da PUCRS for permitting the use of laboratory facilities. We are also grateful to Dr. A. Leyva (U.S.A.) for English editing the manuscript.

#### References

1. Khosla S, Burr D, Cauley J, et al. American Society for Bone and Mineral Research. Bisphosphonate-associated osteonecrosis of the jaw: report of a task force of the American Society for Bone and Mineral Research. *J Bone Miner Res.* 2007;**22**(10):1479-91.

2. Rizzoli R, Burlet N, Cahall D, et al. Osteonecrosis of the jaw and bisphosphonate treatment for osteoporosis. *Bone*. 2008;**42**(5):841-7.

3. Ruggiero SL, Woo SB. Biophosphonate-related osteonecrosis of the jaws. *Dent Clin North Am.* 2008;**52**(1):111-28.

4. Kellinsalmi M, Mönkkönen H, Mönkkönen J, et al. In vitro comparison of clodronate, pamidronate and zoledronic acid effects on rat osteoclasts and human stem cell-derived osteoblasts. *Basic Clin Pharmacol Toxicol*. 2005;**97**(6):382-91.

5. Senel FC, Duman MK, Muci E, et al. Jaw bone changes in rats after treatment with zoledronate and pamidronate. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2010;109(3):385-91.

6. Sonis ST, Watkins BA, Lyng GD, Lerman MA, Anderson KC. Bony changes in the jaws of rats treated with zoledronic acid and dexamethasone before dental extractions mimic bisphosphonate-related osteonecrosis in cancer patients. *Oral Oncol.* 2009;**45**(2):164-72.

7. Bi Y, Gao Y, Ehirchiou D, et al. Bisphosphonates cause osteonecrosis of the jaw-like disease in mice. *Am J Pathol*. 2010;**177**(1):280-90.

8. Naidu A, Dechow PC, Spears R, Wright JM, Kessler HP, Opperman LA. The effects of bisphosphonates on osteoblasts in vitro. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2008;**106**(1):5-13.

9.Walter C, Klein MO, Pabst A, Al-Nawas B, Duschner H, Ziebart T. Influence of bisphosphonates on endothelial cells, fibroblasts, and osteogenic cells. *Clin Oral Investig*. 2010;**14**(1):35-41.

10. Yamada J, Tsuno NH, Kitayama J, et al. Anti-angiogenic property of zoledronic acid by inhibition of endothelial progenitor of cell differentiation. *J Surg Res.* 2009;**151**(1):115-20.

11. Wood J, Bonjean K, Ruetz S, et al. Novel antiangiogenic effects of the bisphosphonate compound zoledronic acid. *J Pharmacol Exp Ther.* 2002;**302**(3):1055-61.

12. Kobayashi Y, Hiraga T, Ueda A, et al. Zoledronic acid delays wound healing of the tooth extraction socket, inhibits oral epithelial cell migration, and promotes proliferation

and adhesion to hydroxyapatite of oral bacteria, without causing osteonecrosis of the jaw, in mice. *J Bone Miner Metab.* 2010;**28**(2):165-75.

13. Kim YH, Kim GS, Jeong-Hwa B. Inhibitory action of bisphosphonates on bone resorption does not involve the regulation of RANKL and OPG expression. *Exp Mol Med.* 2002;**34**(2):145-51.

14. Mountzios G, Terpos E, Syrigos K, et al. Markers of bone remodeling and skeletal morbidity in patients with solid tumors metastatic to the skeleton receiving the bisphosphonate zoledronic acid. *Transl Res.* 2010;**155**(5):247-55.

15. Landesberg R, Cozin M, Cremers S, et al. Inhibition of oral mucosal cell wound healing by bisphosphonates. *J Oral Maxillofac Surg*. 2008;**66**(5):839-47.

16. Landesberg R, Woo V, Cremers S, et al. Potential pathophysiological mechanisms in osteonecrosis of the jaw. *Ann N Y Acad Sci.* 2011;1218:62-79. doi: 10.1111/j.1749-6632.2010.05835.x.

17. Corrado A, Neve A, Maruotti N, Gaudio A, Marucci A, Cantatore FP. Dose-dependent metabolic effect of zoledronate on primary human osteoblastic cell cultures. *Clin Exp Rheumatol*. 2010;**28**(6):873-9.

18. Crépin S, Laroche ML, Sarry B, Merle L.Osteonecrosis of the jaw induced by clodronate, an alkylbisphosphonate: case report and literature review. *Eur J Clin Pharmacol*. 2010;**66**(6):547-54.

19. Amenábar JM, Martins GB, Cherubini K, Figueiredo MA. Comparison between semiautomated segmentation and manual point-counting methods for quantitative analysis of histological sections. *J Oral Sci.* 2006;**48**(3):139-43.

20. Fedele S, Porter SR, D'Aiuto F, et al. Nonexposed variant of bisphosphonate-associated osteonecrosis of the jaw: a case series. *Am J Med.* 2010;**123**(11):1060-4.

21. Junquera L, Gallego L. Nonexposed bisphosphonate-related osteonecrosis of the jaws: another clinical variant? *J Oral Maxillofac Surg.* 2008;**66**(7):1516-7.

22. Mawardi H, Treister N, Richardson P, et al. Sinus tracts--an early sign of bisphosphonate-associated osteonecrosis of the jaws? *J Oral Maxillofac Surg.* 2009;**67**(3):593-601.

23. Agis H, Blei J, Watzek G, Gruber R. Is zoledronate toxic to human periodontal fibroblasts? *J Dent Res.* 2010;**89**(1):40-5.

24. Allen M R, Burr DB. The pathogenesis of bisphosphonate-related osteonecrosis of the jaw: so many hypotheses, so few data. *J Oral Maxillofac Surg* 2009;**67**(5 Suppl): 61-70.

25. Scheper MA, Badros A, Chaisuparat R, Cullen KJ, Meiller TF. Effect of zoledronic acid on oral fibroblasts and epithelial cells: a potential mechanism of bisphosphonate-associated osteonecrosis. *Br J Haematol*. 2009;**144**(5):667-76.

26. Reid IR, Bolland MJ, Grey AB. Is bisphosphonate-associated osteonecrosis of the jaw caused by soft tissue toxicity? *Bone*. 2007;**41**(3):318-20.

27. Cardaropoli G, Araújo M, Lindhe J. Dynamics of bone tissue formation in tooth extraction sites. An experimental study in dogs. *J Clin Periodontol*. 2003;**30**(9):809-18.

28. Bodner L, Kaffe I, Littner MM, Cohen J. Extraction site healing in rats. A radiologic densitometric study. *Oral Surg Oral Med Oral Pathol* 1993;**75**(3): 367-72.

29. Huebsch RF, Coleman RD, Frandsen AM, Becks H. The healing process following molar extraction. I. Normal male rats (long-evans strain). *Oral Surg Oral Med Oral Pathol* 1952;**5**(8): 864-76.

30. Kahnberg KE, Thilander H. Healing of experimental excisional wounds in the rat palate. (I) Histological study of the interphase in wound healing after sharp dissection. *Int J Oral Surg.* 1982;**11**(1):44-51.

31. Migliorati CA, Schubert MM, Peterson DE, Seneda LM. Bisphosphonate-associated osteonecrosis of mandibular and maxillary bone: an emerging oral complication of supportive cancer therapy. Cancer. 2005;104(1):83-93.

32. Ruggiero SL, Dodson TB, Assael LA, Landesberg R, Marx RE, Mehrotra B. American Association of Oral and Maxillofacial Surgeons position paper on bisphosphonate-related osteonecrosis of the jaw - 2009 update. *Aust Endod J* 2009;**35**(3): 119-30.

33. Maahs MP, Azambuja AA, Campos MM, Salum FG, Cherubini K. Association between bisphosphonates and jaw osteonecrosis: a study in Wistar rats. *Head Neck*. 2011;**33**(2):199-207.

34. Cavanna L, Bertè R, Arcari A, Mordenti P, Pagani R, Vallisa D. Osteonecrosis of the jaw. A newly emerging site-specific osseous pathology in patients with cancer treated with bisphosphonates. Report of five cases and review of the literature. *Eur J Intern Med.* 2007;**18**(5):417-22.

35. Marx RE, Sawatari Y, Fortin M, Broumand V. Bisphosphonate-induced exposed bone (osteonecrosis/osteopetrosis) of the jaws: risk factors, recognition, prevention, and treatment. *J Oral Maxillofac Surg.* 2005;**63**(11):1567-75.

36. Ruggiero SL, Mehrotra B, Rosenberg TJ, Engroff SL. Osteonecrosis of the jaws associated with the use of bisphosphonates: a review of 63 cases. *J Oral Maxillofac Surg.* 2004;**62**(5):527-34.

37. Kruszewska H, Zareba T, Tyski S. Search of antimicrobial activity of selected non-antibiotic drugs. *Acta Pol Pharm.* 2002;**59**(6):436-9.

38. Cozin M, Pinker BM, Solemani K, et al. Novel therapy to reverse the cellular effects of bisphosphonates on primary human oral fibroblasts. *J Oral Maxillofac* Surg. 2011;**69**(10):2564-78.

39. Allen MR, Burr DB. Mandible matrix necrosis in beagle dogs after 3 years of daily oral bisphosphonate treatment. *J Oral Maxillofac Surg* 2008;**66**(5): 987-94.

40. Allen MR, Kubek DJ, Burr DB. Cancer treatment dosing regimens of zoledronic acid result in near-complete suppression of mandible intracortical bone remodeling in beagle dogs. *J Bone Miner Res.* 2010;**25**(1):98-105.

41. Reid IR. Osteonecrosis of the jaw: who gets it, and why? Bone 2009; 44(1): 4-10.

42. Wehrhan F, Stockmann P, Nkenke E, et al. Differential impairment of vascularization and angiogenesis in bisphosphonate-associated osteonecrosis of the jaw-related mucoperiosteal tissue. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2011;**112**(2):216-21.

43. Yamada J, Tsuno NH, Kitayama J, et al. Anti-angiogenic property of zoledronic acid by inhibition of endothelial progenitor cell differentiation. *J Surg Res* 2009;**151**(1): 115-20.

44. Ziebart T, Pabst A, Klein MO, et al. Bisphosphonates: restrictions for vasculogenesis and angiogenesis: inhibition of cell function of endothelial progenitor cells and mature endothelial cells in vitro. *Clin Oral Investig.* 2011;**15**(1):105-11.

45. Walter C, Pabst A, Ziebart T, Klein M, Al-Nawas B. Bisphosphonates affect migration ability and cell viability of HUVEC, fibroblasts and osteoblasts in vitro. *Oral Dis.* 2011;**17**(2):194-9.

46. Ribatti D, Maruotti N, Nico B, et al. Clodronate inhibits angiogenesis in vitro and in vivo. *Oncol Rep.* 2008;**19**(5):1109-12.

47. Santini D, Vincenzi B, Dicuonzo G, et al. Zoledronic acid induces significant and longlasting modifications of circulating angiogenic factors in cancer patients. *Clin Cancer Res.* 2003;9(8):2893-7.

48. Vincenzi B, Santini D, Dicuonzo G, et al. Zoledronic acid-related angiogenesis modifications and survival in advanced breast cancer patients. *J Interferon Cytokine Res.* 2005;**25**(3):144-51.

49. Schor AM, Pendleton N, Pazouki S, et al. Assessment of vascularity in histological sections: effects of methodology and value as an index of angiogenesis in breast tumours. *Histochem J.* 1998;**30**(12):849-56.

50. Freitas TM, Miguel MC, Silveira EJ, Freitas RA, Galvão HC. Assessment of angiogenic markers in oral hemangiomas and pyogenic granulomas. *Exp Mol Pathol.* 2005;**79**(1):79-85.

51. Carvalho TL, Bombonato KF, Brentegani LG. Histometric analysis of rat alveolar wound healing. *Braz Dent J.* 1997;**8**(1):9-12.

52. Hill, P. A. Bone remodelling. Br J Orthod. 1998; 25(2):101-7.

53. Alvarez L, Peris P, Guañabens N, et al. Serum osteoprotegerin and its ligand in Paget's disease of bone: relationship to disease activity and effect of treatment with bisphosphonates. *Arthritis Rheum.* 2003;**48**(3):824-8.

54. Martini G, Gennari L, Merlotti D et al. Serum OPG and RANKL levels before and after intravenous bisphosphonate treatment in Paget's disease of bone. *Bone*. 2007;**40**(2):457-63.

55. Nakashima T, Kobayashi Y, Yamasaki S, et al. Protein expression and functional difference of membrane-bound and soluble receptor activator of NF-kappaB ligand: modulation of the expression by osteotropic factors and cytokines. *Biochem Biophys Res Commun.* 2000;**275**(3):768-75.

56. Li B, Ling Chau JF, Wang X, Leong WF. Bisphosphonates, specific inhibitors of osteoclast function and a class of drugs for osteoporosis therapy. *J Cell Biochem*. 2011;**112**(5):1229-42.

57. Conrad KA, Lee SM. Clodronate kinetics and dynamics. *Clin Pharmacol Ther*. 1981;**30**(1):114-20.

DISCUSSÃO GERAL

#### 4 DISCUSSÃO GERAL

A eficácia dos bisfosfonatos na prevenção e no tratamento de doenças caracterizadas por excessiva reabsorção óssea foi o fator responsável pela ampla difusão do uso desses fármacos a partir de sua introdução no mercado farmacêutico na década de 60 (RUSSELL, 2006). Entretanto, a ocorrência de casos de osteonecrose maxilar passou a constituir um importante efeito adverso, que trouxe algumas restrições ao uso desses medicamentos, bem como a necessidade de condutas preventivas e curativas para a condição (RUGGIERO, 2011).

A literatura comporta uma série de pesquisas que investigam os efeitos dos bisfosfonatos por meio da aplicação de diversos marcadores e métodos distintos. A associação com a osteonecrose é comprovada (MAAHS et al., 2011), existindo, entretanto, variações da intensidade do risco de acordo com as condições do paciente e especificidades do fármaco (RUGGIERO, 2011). Por outro lado, o mecanismo exato pelo qual a osteonecrose é induzida não está claramente definido. Algumas publicações levantam a possibilidade de a lesão iniciar na mucosa e, nesse contexto, têm explorado o efeito tóxico sobre células epiteliais incluindo as gastrointestinais, as renais e as da mucosa oral (LANDESBERG et al., 2008, PERAZELLA, 2003, TWISS et al., 1994). O desenvolvimento de úlceras e erosões gástricas é um reconhecido efeito colateral associado à administração oral de bisfosfonatos nitrogenados (WALLACE et al., 1999). No entanto, não está definido se o efeito que exercem sobre o epitélio oral seria capaz de iniciar a osteonecrose dos maxilares.

No presente estudo, verificou-se que, a despeito da ocorrência de osso não-vital, algumas lesões de tecido mole exibiram regeneração completa da mucosa, não tendo sido observada, nesse sítio, associação da variável solução de continuidade da mucosa com o uso de bisfosfonato. Tal achado, somado aos resultados observados para os marcadores imunoistoquímicos caspase-3, von Willebrand, OPG, RANK e RANKL indica pouca repercussão dos efeitos dos bisfosfonatos sobre a mucosa oral. A ausência de associação entre a expressão de caspase-3 e os bisfosfonatos empregados sugere que não haja aumento das taxas de apoptose em epitélio e conjuntivo. Assim, o importante efeito que esses fármacos exercem no tecido ósseo ao interferirem na via do mevalonato ou transformarem-se em análogos de ATP e, consequentemente, levarem os osteoclastos à apoptose (LI et al., 2011; RUSSELL, 2006) parece não ser um evento significativo nesses outros tecidos. Isso, provavelmente, pela distribuição significativamente menor do fármaco em sítios extra-ósseos (RUSSELL, 2007). Ainda, é provável que seja a diferença de sua biodisponibilidade o fator responsável pelos resultados divergentes ao compararem-se pesquisas *in vitro* e *in vivo*. A maior parte das pesquisas que relatam efeitos significativos dos bisfosfonatos fora do tecido ósseo é composta por estudos com cultura de células, nos quais, por vezes, as concentrações do fármaco empregadas são superiores àquelas aplicadas *in vivo* e, mesmo quando em concentrações equivalentes às doses terapêuticas, sua biodisponibilidade é maior.

A análise realizada em apenas um período após as intervenções cirúrgicas, no entanto, não permitiu considerar o efeito dos fármacos de acordo com a cronologia do processo de reparo. É possível que os bisfosfonatos interfiram em fases iniciais desse processo, quando há intensa proliferação de fibroblastos e queratinócitos além da deposição de colágeno (RAVOSA et al., 2011). Assim, estudos que permitam avaliar a dinâmica do reparo tecidual, considerando a sequência de eventos que o constituem, podem ser úteis para determinar se alterações intrínsecas aos tecidos moles orais contribuem para a iniciação e/ou progressão da osteonecrose associada ao uso desses fármacos.

Os resultados são sugestivos, entretanto, de que os efeitos preponderantes ocorram, de fato, no tecido ósseo. Recentemente, estudos clínicos têm relatado casos de osteonecrose maxilar em pacientes usuários de denosumab, um anticorpo monoclonal empregado na prevenção e no tratamento de doenças caracterizadas por excessiva reabsorção óssea (HENRY et al., 2011). Essa droga, cuja farmacocinética difere da dos bisfosfonatos, exerce seus efeitos ao ligar-se ao RANKL, inibindo a atividade osteoclástica (STOPECK et al., 2010). Tais achados reforçam a hipótese de que o prejuízo da função osteoclástica constitui evento primário na patogênese da condição.

Ao compararem-se os efeitos de bisfosfonatos nitrogenados e não-nitrogenados, o clodronato, importante representante dos não-nitrogenados, foi capaz de induzir a ocorrência de osso não-vital, embora em menor prevalência. É preciso lembrar que esse medicamento foi administrado por via parenteral, o que resulta em doses cumulativas significativamente mais elevadas que as resultantes da administração oral. Assim, considerando-se a meia-vida dos bisfosfonatos, sua afinidade pelo tecido ósseo e o fato de não serem metabolizados neste sítio (LI et al., 2011), é plausível inferir que o risco de osteonecrose seja proporcional à dose cumulativa no tecido ósseo. Esta, por sua vez, estará na dependência da estrutura química do fármaco e da via de administração. A ausência do radical hidroxila confere ao clodronato menor afinidade à estrutura óssea, se comparado ao ácido zoledrônico (LI et al., 2011). Entretanto, o menor risco de osteonecrose que estaria associado à menor afinidade ao tecido ósseo poderá ser compensado pelo maior tempo de uso.

Os achados do presente estudo permitem inferir que tanto bisfosfonatos nitrogenados quanto não-nitrogenados são capazes de induzir a ocorrência de osteonecrose, e seu efeito sobre a mucosa oral não parece constituir fator iniciador da lesão. A alta afinidade desses compostos pela hidroxiapatita (RUSSEL, 2006) e os relatos de sua

potente atividade na supressão da remodelação óssea (ALLEN et al., 2010) corroboram a ideia de que seus efeitos ocorram, primordialmente, em estrutura óssea. Ainda, a maior parte dos estudos que avaliam os efeitos biológicos dos bisfosfonatos o faz por meio de ensaios *in vitro*, com dosagens e condições que podem não representar as da rotina clínica. Assim, muitos aspectos relacionados à etiopatogenia da doença permanecem pouco esclarecidos.

A presente pesquisa, ao testar o efeito do ácido zoledrônico e do clodronato em feridas induzidas em modelo animal, obteve alguns resultados divergentes dos relatados na literatura. Novas pesquisas que investiguem a distribuição desses fármacos nos diferentes tecidos, correlacionando-a aos possíveis efeitos farmacológicos, poderão corroborar tais achados. A melhor compreensão dos eventos biofarmacológicos que envolvem os bisfosfonatos no organismo humano poderá nortear condutas e fornecer critérios para tomada de decisão mediante pacientes usuários desses fármacos, sejam eles portadores ou não de osteonecrose maxilar.

# REFERÊNCIAS

### REFERÊNCIAS

Agis H, Blei J, Watzek G, Gruber R. Is zoledronate toxic to human periodontal fibroblasts? J Dent Res. 2010;89(1):40-5.

Allegra A, Oteri G, Nastro E, Alonci A, Bellomo G, Del Fabro V, Quartarone E, Alati C, De Ponte FS, Cicciù D, Musolino C.Patients with bisphosphonates-associated osteonecrosis of the jaw have reduced circulating endothelial cells. Hematol Oncol. 2007;25(4):164-9.

Allen MR, Burr DB. Mandible matrix necrosis in beagle dogs after 3 years of daily oral bisphosphonate treatment. J Oral Maxillofac Surg 2008;66(5): 987-94.

Allen, M. R, Burr DB. The pathogenesis of bisphosphonate-related osteonecrosis of the jaw: so many hypotheses, so few data. J Oral Maxillofac Surg 2009; 67(5):61-70.

Allen MR, Kubek DJ, Burr DB. Cancer treatment dosing regimens of zoledronic acid result in near-complete suppression of mandible intracortical bone remodeling in beagle dogs. J Bone Miner Res. 2010;25(1):98-105.

Altundal H, Güvener O. The effect of alendronate on resorption of the alveolar bone following tooth extraction. Int J Oral Maxillofac Surg. 2004 Apr;33(3):286-93.

Alvarez L, Peris P, Guañabens N, Vidal S, Ros I, Pons F, et al. Serum osteoprotegerin and its ligand in Paget's disease of bone: relationship to disease activity and effect of treatment with bisphosphonates. Arthritis Rheum. 2003;48(3):824-8.

Amenábar JM, Martins GB, Cherubini K, Figueiredo MA. Comparison between semiautomated segmentation and manual point-counting methods for quantitative analysis of histological sections. J Oral Sci. 2006;48(3):139-43.

Amir E, Trinkaus M, Simmons CE, Dranitsaris G, Clemons MJ.Vascular endothelial growth factor activity after switching of bisphosphonate treatment for metastatic breast cancer. J Clin Pathol. 2009;62(5):474-6.

Aström E, Magnusson P, Eksborg S, Söderhäll S. Biochemical bone markers in the assessment and pamidronate treatment of children and adolescents with osteogenesis imperfecta. Acta Paediatr. 2010;99(12):1834-40.

Aubin JE, Bonnelye E. Osteoprotegerin and its ligand: a new paradigm for regulation of osteoclastogenesis and bone resorption. Osteoporos Int. 2000;11(11):905-13.

Baroukh B, Saffar JL. Identification of osteoclasts and their mononuclear precursors. A comparative histological and histochemical study in hamster periodontitis. J Periodontal Res. 1991;26(3 Pt 1):161-6.

Benford HL, McGowan NW, Helfrich MH, Nuttall ME, Rogers MJ. Visualization of bisphosphonate-induced caspase-3 activity in apoptotic osteoclasts in vitro. Bone. 2001;28(5):465-73

Bi Y, Gao Y, Ehirchiou D, Cao C, Kikuiri T, Le A, et al. Bisphosphonates cause osteonecrosis of the jaw-like disease in mice. Am J Pathol. 2010;177(1):280-90.

Biasotto M, Chiandussi S, Zacchigna S, Moimas S, Dore F, Pozzato G, et al. A novel animal model to study non-spontaneous bisphosphonates osteonecrosis of jaw. J Oral Pathol Med. 2010;39(5):390-6.

Blyth K, Vaillant F, Jenkins A, McDonald L, Pringle MA, Huser C, et al. Runx2 in normal tissues and cancer cells: A developing story. Blood Cells Mol Dis. 2010; 45(2):117-23.

Bodner L, Kaffe I, Littner MM, Cohen J. Extraction site healing in rats. A radiologic densitometric study. Oral Surg Oral Med Oral Pathol 1993;75(3): 367-72.

Brown JE, McCloskey EV, Dewar JA, Body JJ, Cameron DA, Harnett AN, et al. The use of bone markers in a 6-week study to assess the efficacy of oral clodronate in patients with metastatic bone disease. Calcif Tissue Int. 2007;81(5):341-51.

Brown JE, Sim S. Evolving role of bone biomarkers in castration-resistant prostate cancer. Neoplasia. 2010;12(9):685-96.

Buduneli E, Vardar-Sengül S, Buduneli N, Atilla G, Wahlgren J, Sorsa T.Matrix metalloproteinases, tissue inhibitor of matrix metalloproteinase-1, and laminin-5 gamma2 chain immunolocalization in gingival tissue of endotoxin-induced periodontitis in rats: effects of low-dose doxycycline and alendronate. J Periodontol. 2007;78(1):127-34.

Cardaropoli G, Araújo M, Lindhe J. Dynamics of bone tissue formation in tooth extraction sites. An experimental study in dogs. J Clin Periodontol. 2003;30(9):809-18.

Carvalho TL, Bombonato KF, Brentegani LG. Histometric analysis of rat alveolar wound healing. Braz Dent J. 1997;8(1):9-12.

Cavanna L, Bertè R, Arcari A, Mordenti P, Pagani R, Vallisa D. Osteonecrosis of the jaw. A newly emerging site-specific osseous pathology in patients with cancer treated with bisphosphonates. Report of five cases and review of the literature. Eur J Intern Med. 2007;18(5):417-22.

Cerri PS, Boabaid F, Katchburian E. Combined TUNEL and TRAP methods suggest that apoptotic bone cells are inside vacuoles of alveolar bone osteoclasts in young rats. J Periodontal Res. 2003;38(2):223-6.

Chambers TJ, Fuller K, Darby JA. Hormonal regulation of acid phosphatase release by osteoclasts disaggregated from neonatal rat bone. J Cell Physiol. 1987;132(1):90-6.

Chang HY, Yang X. Proteases for cell suicide: functions and regulation of caspases. Microbiol Mol Biol Rev. 2000;64(4):821-46.

Chenu C, Colucci S, Grano M, Zigrino P, Barattolo R, Zambonin G, et al. Osteocalcin induces chemotaxis, secretion of matrix proteins, and calcium-mediated intracellular signaling in human osteoclast-like cells. J Cell Biol.1994;127(4):1149-58.

Chowdhury I, Tharakan B, Bhat GK. Caspases - an update. Comp Biochem Physiol B Biochem Mol Biol. 2008;151(1):10-27.

Chun YH, Foster BL, Lukasavage PA, Berry JE, Zhao M, Tenenbaum HC, Somerman MJ. Bisphosphonate modulates cementoblast behavior in vitro. J Periodontol. 2005;76(11):1890-900.

Coleman R, Brown J, Terpos E, Lipton A, Smith MR, Cook R, et al. Bone markers and their prognostic value in metastatic bone disease: clinical evidence and future directions. Cancer Treat Rev. 2008;34(7):629-39.

Conrad KA, Lee SM. Clodronate kinetics and dynamics. Clin Pharmacol Ther. 1981;30(1):114-20.

Corrado A, Neve A, Maruotti N, Gaudio A, Marucci A, Cantatore FP. Dose-dependent metabolic effect of zoledronate on primary human osteoblastic cell cultures. Clin Exp Rheumatol. 2010;28(6):873-9.

Coxon FP, Thompson K, Rogers MJ. Recent advances in understanding the mechanism of action of bisphosphonates. Curr Opin Pharmacol. 2006;6(3):307-12.

Cozin M, Pinker BM, Solemani K, Zuniga JM, Dadaian SC, Cremers S, et al. Novel therapy to reverse the cellular effects of bisphosphonates on primary human oral fibroblasts. J Oral Maxillofac Surg. 2011;69(10):2564-78.

Crépin S, Laroche ML, Sarry B, Merle L.Osteonecrosis of the jaw induced by clodronate, an alkylbisphosphonate: case report and literature review. Eur J Clin Pharmacol. 2010;66(6):547-54.

Enjuanes A, Ruiz-Gaspà S, Peris P, Ozalla D, Álvarez L, Combalia A, et al. The effect of the alendronate on OPG/RANKL system in differentiated primary human osteoblasts. Endocrine. 2010;37(1):180-6.

Escudero ND, Lacave M, Ubios AM, Mandalunis PM. Effect of monosodium olpadronate on osteoclasts and megakaryocytes: an in vivo study. J Musculoskelet Neuronal Interact. 2009;9(2):109-20.

Eslami B, Zhou S, Van Eekeren I, LeBoff MS, Glowacki J. Reduced osteoclastogenesis and RANKL expression in marrow from women taking alendronate. Calcif Tissue Int. 2011;88(4):272-80.

Facchini G, Caraglia M, Morabito A, Marra M, Piccirillo MC, Bochicchio AM, et al. Metronomic administration of zoledronic acid and taxotere combination in castration resistant prostate cancer patients: phase I ZANTE trial. Cancer Biol Ther. 2010;10(6):543-8.

Faloni AP, Sasso-Cerri E, Katchburian E, Cerri PS. Decrease in the number and apoptosis of alveolar bone osteoclasts in estrogen-treated rats. J Periodontal Res. 2007;42(3):193-201.

Fedele S, Porter SR, D'Aiuto F, Aljohani S, Vescovi P, Manfredi M, et al. Nonexposed variant of bisphosphonate-associated osteonecrosis of the jaw: a case series. Am J Med. 2010;123(11):1060-4.

Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. Nat Med. 2003;9(6):669-76.

Ferrara N.Vascular endothelial growth factor: basic science and clinical progress. Endocr Rev. 2004;25(4):581-611.

Freitas TM, Miguel MC, Silveira EJ, Freitas RA, Galvão HC. Assessment of angiogenic markers in oral hemangiomas and pyogenic granulomas. Exp Mol Pathol. 2005;79(1):79-85.

Fujimura Y, Kitaura H, Yoshimatsu M, Eguchi T, Kohara H, Morita Y, Yoshida N. Influence of bisphosphonates on orthodontic tooth movement in mice. Eur J Orthod. 2009; (6):572-7.

Fujita Y, Watanabe K, Uchikanbori S, Maki K. Effects of risedronate on cortical and trabecular bone of the mandible in glucocorticoid-treated growing rats. Am J Orthod Dentofacial Orthop. 2011;139(3):e267-77.

Hansen T, Kirkpatrick CJ, Walter C, Kunkel M. Increased numbers of osteoclasts expressing cysteine proteinase cathepsin K in patients with infected osteoradionecrosis and bisphosphonate-associated osteonecrosis--a paradoxical observation? Virchows Arch. 2006;449(4):448-54.

Henry DH, Costa L, Goldwasser F, Hirsh V, Hungria V, Prausova J, et al. Randomized, double-blind study of denosumab versus zoledronic acid in the treatment of bone metastases in patients with advanced cancer (excluding breast and prostate cancer) or multiple myeloma. J Clin Oncol. 2011; 29(9):1125-32.

Herrmann M, Seibel MJ. The amino- and carboxyterminal cross-linked telopeptides of collagen type I, NTX-I and CTX-I: a comparative review. Clin Chim Acta. 2008;393(2):57-75.

Hill, P. A. Bone remodelling. Br J Orthod 1998; 25(2): 101-7.

Hu H, Djuretic I, Sundrud MS, Rao A. Transcriptional partners in regulatory T cells: Foxp3, Runx and NFAT. Trends Immunol. 2007;28(8):329-32.

Huebsch RF, Coleman RD, Frandsen AM, Becks H. The healing process following molar extraction. I. Normal male rats (long-evans strain). Oral Surg Oral Med Oral Pathol 1952;5(8): 864-76.

Igarashi Y, Lee MY, Matsuzaki S. Acid phosphatases as markers of bone metabolism. J Chromatogr B Analyt Technol Biomed Life Sci. 2002;781(1-2):345-58.

Ito M, Amizuka N, Nakajima T, Ozawa H. Ultrastructural and cytochemical studies on cell death of osteoclasts induced by bisphosphonate treatment. Bone. 1999;25(4):447-52.

Junquera L, Gallego L. Nonexposed bisphosphonate-related osteonecrosis of the jaws: another clinical variant? J Oral Maxillofac Surg. 2008;66(7):1516-7.

Kahnberg KE, Thilander H. Healing of experimental excisional wounds in the rat palate. (I) Histological study of the interphase in wound healing after sharp dissection. Int J Oral Surg. 1982;11(1):44-51.

Kawana K, Takahashi M, Hoshino H, Kushida K. Comparison of serum and urinary C-terminal telopeptide of type I collagen in aging, menopause and osteoporosis. Clin Chim Acta. 2002;316(1-2):109-15.

Kellinsalmi M, Mönkkönen H, Mönkkönen J, Leskelä HV, Parikka V, Hämäläinen M, et al. In vitro comparison of clodronate, pamidronate and zoledronic acid effects on rat osteoclasts and human stem cell-derived osteoblasts. Basic Clin Pharmacol Toxicol. 2005;97(6):382-91.

Khosla S. Minireview: the OPG/RANKL/RANK system. Endocrinology. 2001;142(12):5050-5.

Khosla S, Burr D, Cauley J, Dempster DW, Ebeling PR, Felsenberg D, et al. Bisphosphonate-associated osteonecrosis of the jaw: report of a task force of the American Society for Bone and Mineral Research. J Bone Miner Res. 2007;22(10):1479-91.

Kobayashi Y, Hiraga T, Ueda A, Wang L, Matsumoto-Nakano M, Hata K, et al. Zoledronic acid delays wound healing of the tooth extraction socket, inhibits oral epithelial cell migration, and promotes proliferation and adhesion to hydroxyapatite of oral bacteria, without causing osteonecrosis of the jaw, in mice. J Bone Miner Metab. 2010;28(2):165-75.

Koch FP, Merkel C, Al-Nawas B, Smeets R, Ziebart T, Walter C, et al. Zoledronate, ibandronate and clodronate enhance osteoblast differentiation in a dose dependent manner - A quantitative in vitro gene expression analysis of Dlx5, Runx2, OCN, MSX1 and MSX2. J Craniomaxillofac Surg. 2011;doi:10.1016/j.jcms.2010.10.007.

Kovacevic M, Tamarut T, Zoricic S, Beslic S. A method for histological enzyme histochemical and immunohistochemical analysis of periapical diseases on undercalcified bone with teeth. Acta Estomat Croat. 2003; 37(3):269-273.

Krempien R, Huber PE, Harms W, Treiber M, Wannenmacher M, Krempien B. Combination of early bisphosphonate administration and irradiation leads to improved remineralization and restabilization of osteolytic bone metastases in an animal tumor model. Cancer. 2003;98(6):1318-24.

Kruszewska H, Zareba T, Tyski S. Search of antimicrobial activity of selected nonantibiotic drugs. Acta Pol Pharm. 2002;59(6):436-9.

Labat-Moleur F, Guillermet C, Lorimier P, Robert C, Lantuejoul S, Brambilla E, et al. TUNEL apoptotic cell detection in tissue sections: critical evaluation and improvement. J Histochem Cytochem. 1998;46(3):327-34.

Lacey DL, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, et al. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. Cell. 1998;93(2):165-76.

Landesberg R, Cozin M, Cremers S, Woo V, Kousteni S, Sinha S, et al. Inhibition of oral mucosal cell wound healing by bisphosphonates. J Oral Maxillofac Surg. 2008;66(5):839-47.

Landesberg R, Woo V, Cremers S, Cozin M, Marolt D, Vunjak-Novakovic G, et al. Potential pathophysiological mechanisms in osteonecrosis of the jaw. Ann N Y Acad Sci. 2011;1218:62-79. doi: 10.1111/j.1749-6632.2010.05835.x.

Lekic P, Rubbino I, Krasnoshtein F, Cheifetz S, McCulloch CA, Tenenbaum H. Bisphosphonate modulates proliferation and differentiation of rat periodontal ligament cells during wound healing. Anat Rec.1997;247(3):329-40.

Li B, Ling Chau JF, Wang X, Leong WF. Bisphosphonates, specific inhibitors of osteoclast function and a class of drugs for osteoporosis therapy. J Cell Biochem. 2011; 112(5):1229-42.

Lip GY, Blann A. von Willebrand factor: a marker of endothelial dysfunction in vascular disorders? Cardiovasc Res. 1997;34(2):255-65.

Maahs MP, Azambuja AA, Campos MM, Salum FG, Cherubini K. Association between bisphosphonates and jaw osteonecrosis: a study in Wistar rats. Head Neck. 2011; 33(2):199-207.

Marolt D, Cozin M, Vunjak-Novakovic G, Cremers S, Landesberg R. Effects of Pamidronate on Human Alveolar Osteoblasts In Vitro. J Oral Maxillofac Surg. 2011; doi:10.1016/j.joms.2011.05.002.

Martini G, Gennari L, Merlotti D, Salvadori S, Franci MB, Campagna S, et al. Serum OPG and RANKL levels before and after intravenous bisphosphonate treatment in Paget's disease of bone. Bone. 2007;40(2):457-63.

Marx RE, Sawatari Y, Fortin M, Broumand V. Bisphosphonate-induced exposed bone (osteonecrosis/osteopetrosis) of the jaws: risk factors, recognition, prevention, and treatment. J Oral Maxillofac Surg. 2005;63(11):1567-75.

Mawardi H, Treister N, Richardson P, et al. Sinus tracts--an early sign of bisphosphonateassociated osteonecrosis of the jaws? J Oral Maxillofac Surg. 2009;67(3):593-601.

Migliorati CA, Schubert MM, Peterson DE, Seneda LM. Bisphosphonate-associated osteonecrosis of mandibular and maxillary bone: an emerging oral complication of supportive cancer therapy. Cancer. 2005;104(1):83-93.

Mountzios G, Terpos E, Syrigos K, Papadimitriou C, Papadopoulos G, Bamias A, et al. Markers of bone remodeling and skeletal morbidity in patients with solid tumors metastatic to the skeleton receiving the biphosphonate zoledronic acid. Transl Res. 2010;155(5):247-55.

Muñoz-Torres M, Reyes-García R, Mezquita-Raya P, Fernández-García D, Alonso G, Luna Jde D, et al. Serum cathepsin K as a marker of bone metabolism in postmenopausal women treated with alendronate. Maturitas. 2009;64(3):188-92.

Naidu A, Dechow PC, Spears R, Wright JM, Kessler HP, Opperman LA. The effects of bisphosphonates on osteoblasts in vitro. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2008;106(1):5-13.

Nakaya H, Osawa G, Iwasaki N, Cochran DL, Kamoi K, Oates TW. Effects of bisphosphonate on matrix metalloproteinase enzymes in human periodontal ligament cells. J Periodontol. 2000;71(7):1158-66.

Nakashima T, Kobayashi Y, Yamasaki S, Kawakami A, Eguchi K, Sasaki H, et al.Protein expression and functional difference of membrane-bound and soluble receptor activator of NF-kappaB ligand: modulation of the expression by osteotropic factors and cytokines. Biochem Biophys Res Commun. 2000;275(3):768-75.

Nicolin V, Bareggi R, Baldini G, Bortul R, Martinelli B, Narducci P. Effects of neridronic acid on osteoclasts derived by physiological dual-cell cultures. Acta Histochem. 2007;109(5):397-402.

Nishida S, Tsubaki M, Hoshino M, Namimatsu A, Uji H, Yoshioka S, et al. Nitrogencontaining bisphosphonate, YM529/ONO-5920 (a novel minodronic acid), inhibits RANKL expression in a cultured bone marrow stromal cell line ST2. Biochem Biophys Res Commun. 2005;328(1):91-7. Ogawa K, Hori M, Takao R, Sakurada T. Effects of combined elcatonin and alendronate treatment on the architecture and strength of bone in ovariectomized rats. J Bone Miner Metab. 2005;23(5):351-8.

Orwoll ES, Miller PD, Adachi JD, Brown J, Adler RA, Kendler D, et al. Efficacy and safety of a once-yearly i.v. Infusion of zoledronic acid 5mg versus a once-weekly 70mg oral alendronate in the treatment of male osteoporosis: a randomized, multicenter, double-blind, active-controlled study. J Bone Miner Res. 2010;25(10):2239-50.

Pabst AM, Ziebart T, Koch FP, Taylor KY, Al-Nawas B, Walter C. The influence of bisphosphonates on viability, migration, and apoptosis of human oral keratinocytes-in vitro study. Clin Oral Investig. 2011; doi 10.1007/s00784-010-0507-6

Pan B, Farrugia AN, To LB, Findlay DM, Green J, Lynch K, Zannettino AC. The nitrogencontaining bisphosphonate, zoledronic acid, influences RANKL expression in human osteoblast-like cells by activating TNF-alpha converting enzyme (TACE). J Bone Miner Res. 2004;19(1):147-54.

Pazianas M, Miller P, Blumentals WA, Bernal M, Kothawala P. A review of the literature on osteonecrosis of the jaw in patients with osteoporosis treated with oral bisphosphonates: prevalence, risk factors, and clinical characteristics. Clin Ther 2007; 29(8):1548-58.

Perazella MA. Drug-induced renal failure: update on new medications and unique mechanisms of nephrotoxicity. Am J Med Sci. 2003; 325(6):349-62.

Plotkin LI, Manolagas SC, Bellido T. Dissociation of the pro-apoptotic effects of bisphosphonates on osteoclasts from their anti-apoptotic effects on osteoblasts/osteocytes with novel analogs. Bone. 2006;39(3):443-52.

Price CP, Kirwan A, Vader C. Tartrate-resistant acid phosphatase as a marker of bone resorption. Clin Chem. 1995;41(5):641-3.

Ravosa MJ, Ning J, Liu Y, Stack MS. Bisphosphonate effects on the behaviour of oral epithelial cells and oral fibroblasts. Arch Oral Biol. 2011; 56(5):491-8.

Reid IR. Osteonecrosis of the jaw: who gets it, and why? Bone 2009; 44(1): 4-10.

Reid IR, Bolland MJ, Grey AB. Is bisphosphonate-associated osteonecrosis of the jaw caused by soft tissue toxicity? Bone. 2007;41(3):318-20.

Ribatti D, Maruotti N, Nico B, Longo V, Mangieri D, Vacca A, et al. Clodronate inhibits angiogenesis in vitro and in vivo. Oncol Rep. 2008;19(5):1109-12.

Rizzoli R, Burlet N, Cahall D, Delmas PD, Eriksen EF, Felsenberg D, et al. Osteonecrosis of the jaw and bisphosphonate treatment for osteoporosis. Bone. 2008;42(5):841-7.

Ruggiero SL. Bisphosphonate-related osteonecrosis of the jaw (BRONJ): initial discovery and subsequent development. J Oral Maxillofac Surg. 2009;67(5):13-8.

Ruggiero SL. Bisphosphonate-related osteonecrosis of the jaw: an overview. Ann N Y Acad Sci. 2011;1218:38-46.

Ruggiero SL, Dodson TB, Assael LA, Landesberg R, Marx RE, Mehrotra B. American Association of Oral and Maxillofacial Surgeons position paper on bisphosphonate-related osteonecrosis of the jaw - 2009 update. Aust Endod J 2009;35(3): 119-30.

Ruggiero SL, Mehrotra B, Rosenberg TJ, Engroff SL. Osteonecrosis of the jaws associated with the use of bisphosphonates: a review of 63 cases. J Oral Maxillofac Surg. 2004;62(5):527-34.

Ruggiero SL, Woo SB. Biophosphonate-related osteonecrosis of the jaws. Dent Clin North Am. 2008;52(1):111-28.

Russell, R. G. Bisphosphonates: from bench to bedside. Ann N Y Acad Sci. 2006; 1068: 367-401.

Russell RG. Bisphosphonates: mode of action and pharmacology. Pediatrics. 2007;119 (2):S150-62.

Santini D, Vincenzi B, Galluzzo S, Battistoni F, Rocci L, Venditti O, et al. Repeated intermittent low-dose therapy with zoledronic acid induces an early, sustained, and long-lasting decrease of peripheral vascular endothelial growth factor levels in cancer patients. Clin Cancer Res. 2007;13(15):4482-6.

Santini D, Vincenzi B, Dicuonzo G, Avvisati G, Massacesi C, Battistoni F, et al. Zoledronic acid induces significant and long-lasting modifications of circulating angiogenic factors in cancer patients. Clin Cancer Res. 2003;9(8):2893-7.

Scheper MA, Badros A, Chaisuparat R, Cullen KJ, Meiller TF. Effect of zoledronic acid on oral fibroblasts and epithelial cells: a potential mechanism of bisphosphonate-associated osteonecrosis. Br J Haematol. 2009;144(5):667-76.

Schor AM, Pendleton N, Pazouki S, Smither RL, Morris J, Lessan K, et al. Assessment of vascularity in histological sections: effects of methodology and value as an index of angiogenesis in breast tumours. Histochem J. 1998;30(12):849-56.

Seibel MJ. Clinical use of markers of bone turnover in metastatic bone disease. Nat Clin Pract Oncol. 2005;2(10):504-17.

Senel FC, Duman MK, Muci E, Cankaya M, Pampu AA, Ersoz S, et al. Jaw bone changes in rats after treatment with zoledronate and pamidronate. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2010;109(3):385-91.

Senel FC, Saracoglu Tekin U, Durmus A, Bagis B. Severe osteomyelitis of the mandible associated with the use of non-nitrogen-containing bisphosphonate (disodium clodronate): report of a case. J Oral Maxillofac Surg. 2007;65(3):562-5.

Siena S, Bregni M, Brando B, Belli N, Ravagnani F, Gandola L, et al. Flow cytometry for clinical estimation of circulating hematopoietic progenitors for autologous transplantation in cancer patients. Blood. 1991;77(2):400-9.

Simonet WS, Lacey DL, Dunstan CR, Kelley M, Chang MS, Lüthy R, et al. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. Cell. 1997;89(2):309-19.

Sodek J, Ganss B, McKee MD. Osteopontin. Crit Rev Oral Biol Med. 2000;11(3):279-303.

Soltau J, Zirrgiebel U, Esser N, Schächtele C, Totzke F, Unger C, et al. Antitumoral and antiangiogenic efficacy of bisphosphonates in vitro and in a murine RENCA model. Anticancer Res. 2008;28(2A):933-41.

Sonis ST, Watkins BA, Lyng GD, Lerman MA, Anderson KC. Bony changes in the jaws of rats treated with zoledronic acid and dexamethasone before dental extractions mimic bisphosphonate-related osteonecrosis in cancer patients. Oral Oncol. 2009;45(2):164-72.

Spolidorio LC, Marcantonio E Jr, Spolidorio DM, Nassar CA, Nassar PO, Marcantonio RA, Rossa C Jr. Alendronate therapy in cyclosporine-induced alveolar bone loss in rats. J Periodontal Res. 2007;42(5):466-73.

Stopeck AT, Lipton A, Body JJ, Steger GG, Tonkin K, de Boer RH, et al. Denosumab compared with zoledronic acid for the treatment of bone metastases in patients with advanced breast cancer: a randomized, double-blind study. J Clin Oncol. 2010; 28(35):5132-9.

Tannehill-Gregg SH, Levine AL, Nadella MV, Iguchi H, Rosol TJ. The effect of zoledronic acid and osteoprotegerin on growth of human lung cancer in the tibias of nude mice. Clin Exp Metastasis. 2006;23(1):19-31.

Tas F, Duranyildiz D, Oguz H, Camlica H, Yasasever V, Topuz E. Effect of zoledronic acid on serum angiogenic factors in patients with bone metastases. Med Oncol. 2008;25(3):346-9.

Teitelbaum SL. Bone resorption by osteoclasts. Science. 2000;289(5484):1504-8.

Tipton DA, Seshul BA, Dabbous MKh. Effect of bisphosphonates on human gingival fibroblast production of mediators of osteoclastogenesis: RANKL, osteoprotegerin and interleukin-6. J Periodontal Res. 2011;46(1):39-47.

Trouvin AP, Goëb V. Receptor activator of nuclear factor-κB ligand and osteoprotegerin: maintaining the balance to prevent bone loss. Clin Interv Aging. 2010 19;5:345-54

Tsuchimoto M, Azuma Y, Higuchi O, Sugimoto I, Hirata N, Kiyoki M, et al. Alendronate modulates osteogenesis of human osteoblastic cells in vitro. Jpn J Pharmacol. 1994;66(1):25-33.

Tuomela JM, Valta MP, Väänänen K, Härkönen PL. Alendronate decreases orthotopic PC-3 prostate tumor growth and metastasis to prostate-draining lymph nodes in nude mice. BMC Cancer. 2008;8(81):1-12.

Twiss IM, de Water R, den Hartigh J, Sparidans R, Ramp-Koopmanschap W, Brill H, et al. Cytotoxic effects of pamidronate on monolayers of human intestinal epithelial (Caco-2) cells and its epithelial transport. J Pharm Sci. 1994; 83(5):699-703.

Viereck V, Emons G, Lauck V, Frosch KH, Blaschke S, Gründker C, et al. Bisphosphonates pamidronate and zoledronic acid stimulate osteoprotegerin production by primary human osteoblasts. Biochem Biophys Res Commun. 2002 1;291(3):680-6.

Vincenzi B, Santini D, Dicuonzo G, et al. Zoledronic acid-related angiogenesis modifications and survival in advanced breast cancer patients. J Interferon Cytokine Res. 2005;(3):144-51.

von Knoch F, Jaquiery C, Kowalsky M, Schaeren S, Alabre C, Martin I, Rubash HE, Shanbhag AS. Effects of bisphosphonates on proliferation and osteoblast differentiation of human bone marrow stromal cells. Biomaterials. 2005;26(34):6941-9.

Wallace JL, Dicay M, McKnight W, Bastaki S, Blank MA. N-bisphosphonates cause gastric epithelial injury independent of effects on the microcirculation. Aliment Pharmacol Ther. 1999;13(12):1675-82.

Walter C, Klein MO, Pabst A, Al-Nawas B, Duschner H, Ziebart T. Influence of bisphosphonates on endothelial cells, fibroblasts, and osteogenic cells. Clin Oral Investig. 2010;14(1):35-41.

Walter C, Pabst A, Ziebart T, Klein M, Al-Nawas B. Bisphosphonates affect migration ability and cell viability of HUVEC, fibroblasts and osteoblasts in vitro. Oral Dis. 2011;17(2):194-9.

Wehrhan F, Hyckel P, Ries J, Stockmann P, Nkenke E, Schlegel KA, et al. Expression of Msx-1 is suppressed in bisphosphonate associated osteonecrosis related jaw tissueetiopathology considerations respecting jaw developmental biology-related unique features. J Transl Med. 2010;8(96):1-9.

Wehrhan F, Stockmann P, Nkenke E, Schlegel KA, Guentsch A, Wehrhan T, et al. Differential impairment of vascularization and angiogenesis in bisphosphonate-associated osteonecrosis of the jaw-related mucoperiosteal tissue. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2011;112(2):216-21.

Wood J, Bonjean K, Ruetz S, et al. Novel antiangiogenic effects of the bisphosphonate compound zoledronic acid. J Pharmacol Exp Ther. 2002;302(3):1055-61.

Wypij JM, Fan TM, Fredrickson RL, Barger AM, de Lorimier LP, Charney SC. In vivo and in vitro efficacy of zoledronate for treating oral squamous cell carcinoma in cats. J Vet Intern Med. 2008;22(1):158-63.

Xiong H, Wei L, Hu Y, Zhang C, Peng B Effect of alendronate on alveolar bone resorption and angiogenesis in rats with experimental periapical lesions. Int Endod J. 2010;43(6):485-91.

Yamada J, Tsuno NH, Kitayama J, Tsuchiya T, Yoneyama S, Asakage M, et al. Antiangiogenic property of zoledronic acid by inhibition of endothelial progenitor cell differentiation. J Surg Res. 2009 Jan;151(1):115-20.

Yamashita J, Koi K, Yang DY, McCauley LK. Effect of zoledronate on oral wound healing in rats. Clin Cancer Res. 2011;17(6):1405-14.

Zanetta L, Marcus SG, Vasile J, Dobryansky M, Cohen H, Eng K, Shamamian P, Mignatti P.Expression of Von Willebrand factor, an endothelial cell marker, is up-regulated by angiogenesis factors: a potential method for objective assessment of tumor angiogenesis. Int J Cancer. 2000;85(2):281-8.

Zhao X, Xu X, Guo L, Ragaz J, Guo H, Wu J, et al. Biomarker alterations with metronomic use of low-dose zoledronic acid for breast cancer patients with bone metastases and potential clinical significance. Breast Cancer Res Treat. 2010;124(3):733-43.

Ziebart T, Pabst A, Klein MO, Kämmerer P, Gauss L, Brüllmann D, et al. Bisphosphonates: restrictions for vasculogenesis and angiogenesis: inhibition of cell function of endothelial progenitor cells and mature endothelial cells in vitro. Clin Oral Investig. 2011;15(1):105-11.

Zohar R, Lee W, Arora P, Cheifetz S, McCulloch C, Sodek J. Single cell analysis of intracellular osteopontin in osteogenic cultures of fetal rat calvarial cells. J Cell Physiol. 1997;170(1):88-100.

# ANEXOS

# ANEXO A

De	:	ees.oo.0.14e04a.20a76bee@eesmail.elsevier.com em nome Enviada: seg 31/10/2011 1 de Oral Oncology Editorial Office	.4:03
Par	ra:	Karen Cherubini	
CC: Ass	sunto:	Submission Confirmation	
Ane	exos:		
Dea	ar Profe	essor Cherubini,	
You on b	Your submission entitled "Laboratory methods and biomarkers in the evaluation of bisphosphonate effects on body tissues - A literature review" has been received by journal Oral Oncology		
You autł	You will be able to check on the progress of your paper by logging on to Elsevier Editorial Systems as an author. The URL is <a href="http://ees.elsevier.com/oo/">http://ees.elsevier.com/oo/</a> .		
You If yo	Your username is: karen.cherubini@pucrs.br If you need to retrieve password details please go to: <u>http://ees.elsevier.com/oo/automail_query.asp</u>		
You	our manuscript will be given a reference number once an Editor has been assigned.		
Tha	ink you	I for submitting your work to this journal.	
Kinc	d rega	·ds,	
Else Ora	evier Ed I Onco	ditorial System logy	
		•	
		* · · · · · · · · · · · · · · · · · · ·	
			19 <sup>(</sup> )
https	://corr	eio.pucrs.br/exchange/karen.cherubini/Caixa%20de%20entrada/Submissi	0 31/10/2011



### **Guide for Authors**

#### Submission checklist

It is hoped that this list will be useful during the final checking of an article prior to sending it to the journal's Editor for review.

#### Ensure that the following items are present:

•One Author designated as corresponding Author E-mail address Full postal address Telephone(s) and fax numbers

- All necessary files have been uploaded
- Keywords (as comprehensive as possible)
- All figure captions
- All tables (including title, description, footnotes)
- The Author Form has been completed and uploaded to EES

Further considerations:

- Manuscript has been "spellchecked" and is written in good English
- Title is clear and unambiguous

• If the manuscript is an original research article it should contain a structured abstract, if the manuscript is a review article it should contain an unstructured abstract • References are in the correct format for this journal

• All references mentioned in the Reference list are cited in the text, and vice versa

• Permission has been obtained for use of copyrighted material from other sources (including the Web)

• Colour figures are clearly marked as being intended for colour reproduction on the Web (free of charge) and in print or to be reproduced in colour on the Web (free of charge) and in black-and-white in print

• If only colour on the Web is required, black and white versions of the figures are also supplied for printing purposes

• The manuscript conforms to the limits imposed on original research and review articles (3,000 words for original research articles and 5,000 words for review articles, excluding the abstract, keywords and references)

For any further information please contact the Author Support Department at authorsupport@elsevier.com

#### **Prior to Submission**

*Oral Oncology* will consider manuscripts prepared according to the guidelines adopted by the International Committee of Medical Journal Editors ("Uniform requirements for manuscripts submitted to biomedical journals", available as a PDF from http://www.icmje.org). Authors are advised to read these guidelines.

#### Previous Publication

Submission of an article implies that the work described has not been published previously (except in the form of an abstract or as part of a published lecture or academic thesis), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, without the written consent of the Publisher.

#### **Online-only Publication**

*Oral Oncology* offers authors the opportunity to select online-only publication as their preferred option for publishing original research papers in the journal, rather than print publication. Letters to the Editor which are accepted for publication and errata and corrigenda will be published online-only and will not appear in print.

Any material which is published online-only will be published online on ScienceDirect as paginated and fully citable electronic article. It will be listed in the contents page of a printed issue and the full citation and abstract will be published in print. The citation and abstract of the paper will also still appear in the usual abstracting and indexing databases, including PubMed/Medline, Current Contents/Clinical Medicine and the Science Citation Index.

Authors will be asked to select which publication option they would prefer when submitting their paper to the Editorial Office.

#### Randomised Controlled Trials

All randomised controlled trials submitted for publication in *Oral Oncology* should include a completed Consolidated Standards of Reporting Trials (CONSORT) flow chart. Please refer to the CONSORT statement website at http://www.consort-statement.org for more information. *Oral Oncology* has adopted the proposal from the International Committee of Medical Journal Editors (ICMJE) which require, as a condition of consideration for publication of clinical trials, registration in a public trials registry. Trials must register at or before the onset of patient enrolment. The clinical trial registration number should be included at the end of the abstract of the article. For this purpose, a clinical trial is defined as any research project that prospectively assigns human subjects to intervention or comparison groups to study the cause-and-effect relationship between a medical intervention and a health outcome. Studies designed for other purposes, such as to study pharmacokinetics or major toxicity (e.g. phase I trials) would be exempt. Further information can be found at www.icmje.org.

#### Ethics

Work on human beings that is submitted to *Oral Oncology* should comply with the principles laid down in the Declaration of Helsinki; Recommendations guiding physicians in biomedical research involving human subjects. Adopted by the 18th World Medical Assembly, Helsinki, Finland, June 1964, amended by the 29th World Medical Assembly,

Tokyo, Japan, October 1975, the 35th World Medical Assembly, Venice, Italy, October 1983, and the 41st World Medical Assembly, Hong Kong, September 1989. The manuscript should contain a statement that the work has been approved by the appropriate ethical committees related to the institution(s) in which it was performed and that subjects gave informed consent to the work. Studies involving experiments with animals must state that their care was in accordance with institution guidelines. Patients' and volunteers' names, initials, and hospital numbers should not be used.

#### Patient Consent Guidelines

Studies on patients or volunteers require ethics committee approval and informed consent which should be documented in your paper.

Patients have a right to privacy. Therefore, identifying information, including patients' images, names, initials, or hospital numbers, should not be included in videos, recordings, written descriptions, photographs, and pedigrees unless the information is essential for scientific purposes and you have obtained written informed consent for publication in print and electronic form from the patient (or parent, guardian or next of kin where applicable). If such consent is made subject to any conditions, Elsevier must be made aware of all such conditions. Written consents must be provided to Elsevier on request.

Even where consent has been given, identifying details should be omitted if they are not essential. If identifying characteristics are altered to protect anonymity, such as in genetic pedigrees, authors should provide assurance that alterations do not distort scientific meaning and Editors should so note.

If such consent has not been obtained, personal details of patients included in any part of the paper and in any supplementary materials (including all illustrations and videos) must be removed before submission.

#### Conflict of Interest

By means of a "Conflict of interest statement", all authors must disclose any financial and personal relationships with other people or organisations that could inappropriately influence (bias) their work. If there are no conflicts of interest, please state "None declared". This document should be uploaded as a separate file alongside the submitted manuscript.

#### Role of the Funding Source

All sources of funding should be declared as an acknowledgment at the end of the text.

### Authorship and Acknowledgments

All authors must be accredited on the paper and all must submit a completed Author Form with their submission. The form must be signed by the corresponding author on behalf of all authors and can be scanned and uploaded to EES. If you are unable to upload your Author Form to EES, please contact the Editorial Office (oraloncology@elsevier.com) for further information. No subsequent change in authorship will be possible.

#### Copyright

Upon acceptance of an article, Authors will be asked to transfer copyright (for more information on copyright see http://authors.elsevier.com). This transfer will ensure the widest possible dissemination of information. A letter will be sent to the corresponding Author confirming receipt of the manuscript. A form facilitating transfer of copyright will be provided.

If excerpts from other copyrighted works are included, the Author(s) must obtain written permission from the copyright owners and credit the source(s) in the article. Elsevier has preprinted forms for use by Authors in these cases: contact Elsevier's Rights Department, Philadelphia, PA, USA: phone (+1) 215 239 3804, fax (+1) 215 239 3805, e-mail healthpermissions@elsevier.com. Requests may also be completed on-line via the Elsevier homepage (http://www.elsevier.com/locate/permissions).

#### Authors' Rights

As an author you (or your employer or institution) retain certain rights; for details you are referred to: http://www.elsevier.com/authorsrights.

### Manuscript Submission

Submission to Oral Oncology proceeds totally online. Use the following guidelines to your article. Via the "Author Gateway" page of this prepare iournal ( http://authors.elsevier.com/) you will be guided stepwise through the creation and uploading of the various files. The system automatically converts source files to a single Adobe Acrobat PDF version of the article, which is used in the peer-review process. Please note that even though manuscript source files are converted to PDF at submission for the review process, these source files are needed for further processing after acceptance. All correspondence, including notification of the Editor's decision and requests for revision, takes place by e-mail and via the Author's homepage, removing the need for a hard-copy paper trail.

### **General Points**

We accept most wordprocessing formats, but Word, WordPerfect or LaTeX is preferred. Always keep a backup copy of the electronic file for reference and safety. Save your files using the default extension of the program used.

It is important that the file be saved in the native format of the wordprocessor used. The text should be in single-column format. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing the article. In particular, do not use the wordprocessor's options to justify text or to hyphenate words. However, do use bold face, italics, subscripts, superscripts etc. Do not embed "graphically designed" equations or tables, but prepare these using the wordprocessor's facility. When preparing tables, if you are using a table grid, use only one grid for each individual table and not a grid for each row. If no grid is used, use tabs, not spaces, to align columns. The electronic text should be prepared in a way very similar to that of conventional manuscripts Author Gateway's Guide to Publishing with (see also the Elsevier: http://authors.elsevier.com). Do not import the figures into the text file but, instead, indicate their approximate locations directly in the electronic text and on the manuscript. See also the section on Preparation of electronic illustrations.

To avoid unnecessary errors you are strongly advised to use the "spellchecker" function of your wordprocessor.

## Word Limits
Original research articles submitted to the journal must be 3,000 words in length or less (excluding the abstract, keywords and references). Review articles submitted to the journal must be 5,000 words or less in length (excluding the abstract, keywords and references).

#### **Presentation of Manuscript**

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Italics are not to be used for expressions of Latin origin, for example, in vivo, et al., per se. Use decimal points (not commas); use a space for thousands (10 000 and above).

## Language Polishing

Authors who require information about language editing and copyediting services pre- and post-submission please visit http://www.elsevier.com/wps/find/authorshome.authors/languagepolishing or contact authorsupport@elsevier.com for more information. Please note Elsevier neither endorses nor takes responsibility for any products, goods or services offered by outside vendors through our services or in any advertising. For more information please refer to our Terms and Conditions:

http://www.elsevier.com/wps/find/termsconditions.cws\_home/termsconditions

Provide the following data on the title page:

*Title*: Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.

Author names and affiliations: Where the family name may be ambiguous (e.g., a double name), please indicate this clearly. Present the Authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lower-case superscript letter immediately after the Author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name, and, if available, the e-mail address of each Author.

*Corresponding Author*: Clearly indicate who is willing to handle correspondence at all stages of refereeing and publication, also post-publication. Ensure that telephone and fax numbers (with country and area code) are provided in addition to the e-mail address and the complete postal address.

*Present/permanent address*: If an Author has moved since the work described in the article was done, or was visiting at the time, a "Present address" (or "Permanent address") may be indicated as a footnote to that Author's name. The address at which the Author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes.

*Suggestions for reviewers*: Please supply the names of up to three potential reviewers for your manuscript. Please do not suggest reviewers from your own institution, previous or current collaborators. Please provide full names, addresses and email addresses of suggested reviewers. Please note: the final choice of reviewers is that of the Editor and the journal reserves the right not to use reviewers which have been suggested by the authors.

Abstract: A concise and factual abstract of no more than 250 words is required. The abstract must be structured for original research articles and articles reporting the results of

clinical trials. The abstract should be divided by subheadings as follows: Objectives, Materials and Methods, Results and Conclusion.

The abstract should not be structured for review articles. The abstract should state briefly the purpose of the research, the principal results and major conclusions. An abstract is often presented separate from the article, so it must be able to stand alone.

*Keywords*: Immediately after the abstract provide a maximum of ten keywords, to be chosen from the Medical Subject Headings from Index Medicus. These keywords will be used for indexing purposes. It is usually necessary to include keywords such as *Oral Cancer*, or *Head and Neck cancer* 

*Abbreviations*: Define abbreviations or acronyms that are not standard in this field at their first occurrence in the article: in the abstract but also in the main text after it. Ensure consistency of abbreviations throughout the article.

*Text*: This should start on the third page and should be subdivided into the following sections: Introduction, Patients (or Materials) and Methods, Results, and Discussion.

*References*: Responsibility for the accuracy of bibliographic citations lies entirely with the authors.

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. "Unpublished data" and "Personal communications" are not allowed. As an alternative, say in the text, for example, '(data not shown)' or '(Dr F.G. Tomlin, Karolinska Institute)'. Citation of a reference as "in press" implies that the item has been accepted for publication and a copy of the title page of the relevant article must be submitted.

Indicate references by superscript numbers in the text. The actual authors can be referred to, but the reference numbers must always be given. Number the references in the reference list in the order in which they appear in the text.

Examples:

1. Llewellyn CD, Johnson NW, Warnakulasuriya KAAS. Risk factors for squamous cell carcinoma of the oral cavity in young people - comprehensive literature review. *Oral Oncol* 2001;**37**(5):401-418.

2. Gullick WJ, Venter DJ. The c-erbB2 and its expression in human tumors. In: Waxman J, Sikora K, editors. The molecular biology of cancer. Oxford: Blackwell Scientific, 1989. p. 38-53.

3. Scully C, Cawson RA. Medical Problems in Dentistry. 5th edition Oxford: Butterworth-Heinemann. 2004

For more than 6 authors that first 6 should be listed followed by "et al". For further details you are referred to "Uniform Requirements for Manuscripts submitted to Biomedical Journals" (J Am Med Assoc 1997; 277: 927-934) (see also http://www.nlm.nih.gov/tsd/serials/terms\_cond.html).

#### **Figure Captions, Tables, Figures and Schemes**

Present these, in this order, at the end of the article. They are described in more detail below. High-resolution graphics files must always be provided separate from the main text file (see Preparation of illustrations).

#### Footnotes

Footnotes should be used sparingly. Number them consecutively throughout the article, using superscript Arabic numbers. Many wordprocessors build footnotes into the text, and this feature may be used. Should this not be the case, indicate the position of footnotes in the text and present the footnotes themselves on a separate sheet at the end of the article. Do not include footnotes in the Reference list.

#### Table footnotes

Indicate each footnote in a table with a superscript lowercase letter.

#### Tables

Number tables consecutively in accordance with their appearance in the text. Place footnotes to tables below the table body and indicate them with superscript lowercase letters. Avoid vertical rules. Be sparing in the use of tables and ensure that the data presented in tables do not duplicate results described elsewhere in the article.

#### Nomenclature and Units

Follow internationally accepted rules and conventions: use the international system of units (SI). If other quantities are mentioned, give their equivalent in SI.

#### Preparation of Electronic Illustrations

• Make sure you use uniform lettering and sizing of your original artwork.

• Save text in illustrations as "graphics" or enclose the font.

• Only use the following fonts in your illustrations: Arial, Courier, Helvetica, Times, Symbol.

• Number the illustrations according to their sequence in the text.

• Use a logical naming convention for your artwork files.

• Provide all illustrations as separate files and as hardcopy printouts on separate sheets.• Provide captions to illustrations separately.

• Produce images near to the desired size of the printed version.

• A detailed guide on electronic artwork is available on our website: http://authors.elsevier.com/artwork

You are urged to visit this site; some excerpts from the detailed information are given here.

#### Formats

Regardless of the application used, when your electronic artwork is finalised, please "save as" or convert the images to one of the following formats (Note the resolution requirements for line drawings, halftones, and line/halftone combinations given below.):

EPS: Vector drawings. Embed the font or save the text as "graphics".

TIFF: Colour or greyscale photographs (halftones): always use a minimum of 300 dpi.

TIFF: Bitmapped line drawings: use a minimum of 1000 dpi.

TIFF: Combinations bitmapped line/half-tone (colour or greyscale): a minimum of 500 dpi is required.

DOC, XLS or PPT: If your electronic artwork is created in any of these Microsoft Office applications please supply "as is".

Please do not:

• Supply embedded graphics in your wordprocessor (spreadsheet, presentation) document;

• Supply files that are optimised for screen use (like GIF, BMP, PICT, WPG); the resolution is too low;

• Supply files that are too low in resolution;

• Submit graphics that are disproportionately large for the content.

If, together with your accepted article, you submit usable colour figures then Elsevier will ensure, at no additional charge, that these figures will appear in colour on the Web (e.g., ScienceDirect and other sites) in addition to colour reproduction in print.

#### Captions

Ensure that each illustration has a caption. Supply captions separately, not attached to the figure. A caption should comprise a brief title (not on the figure itself) and a description of the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

#### Line drawings

The lettering and symbols, as well as other details, should have proportionate dimensions, so as not to become illegible or unclear after possible reduction; in general, the figures should be designed for a reduction factor of two to three. The degree of reduction will be determined by the Publisher. Illustrations will not be enlarged. Consider the page format of the journal when designing the illustrations.

Do not use any type of shading on computer-generated illustrations.

#### Photographs (halftones)

Remove non-essential areas of a photograph. Do not mount photographs unless they form part of a composite figure. Where necessary, insert a scale bar in the illustration (not below it), as opposed to giving a magnification factor in the caption.

Note that photocopies of photographs are not acceptable.

## Preparation of supplementary data

Elsevier accepts electronic supplementary material to support and enhance your scientific research. Supplementary files offer the Author additional possibilities to publish supporting applications, movies, animation sequences, high-resolution images, background datasets, sound clips and more. Supplementary files supplied will be published online alongside the electronic version of your article in Elsevier Web products, including ScienceDirect: http://www.sciencedirect.com. In order to ensure that your submitted material is directly usable, please ensure that data is provided in one of our recommended file formats. Authors should submit the material in electronic format together with the article and supply a concise and descriptive caption for each file. For more detailed instructions please visit our artwork instruction pages at the Author Gateway at http://authors.elsevier.com/artwork.

#### **Special Subject Repositories**

Elsevier has established agreements and developed policies to allow authors who publish in Elsevier journals to comply with potential manuscript archiving requirements as specified as conditions of their grant awards. To learn more about existing agreements and policies please visit http://www.elsevier.com/fundingbodies.

#### **Sponsored Articles**

*Oral Oncology* now offers authors the option to sponsor non-subscriber access to individual articles. The access sponsorship contribution fee per article is \$3,000. This contribution is necessary to offset publishing costs – from managing article submission and peer review, to typesetting, tagging and indexing of articles, hosting articles on dedicated servers, supporting sales and marketing costs to ensure global dissemination via ScienceDirect, and permanently preserving the published journal article. The sponsorship fee excludes taxes and other potential author fees such as color charges which are additional.

Authors can specify that they would like to select this option after receiving notification that their article has been accepted for publication, but not before. This eliminates a potential conflict of interest by ensuring that the journal does not have a financial incentive to accept an article for publication.

#### Proofs

When your manuscript is received by the Publisher it is considered to be in its final form. Proofs are not to be regarded as "drafts".

One set of page proofs in PDF format will be sent by e-mail to the corresponding Author, to be checked for typesetting/editing. No changes in, or additions to, the accepted (and subsequently edited) manuscript will be allowed at this stage. Proofreading is solely your responsibility.

Elsevier will do everything possible to get your article corrected and published as quickly and accurately as possible. In order to do this we need your help. When you receive the (PDF) proof of your article for correction, it is important to ensure that all of your corrections are sent back to us in one communication. Subsequent corrections will not be possible, so please ensure your first sending is complete. Note that this does not mean you have any less time to make your corrections, just that only one set of corrections will be accepted.

#### **Author Enquiries**

Visit the Author Gateway from Elsevier http://authors.elsevier.com for the facility to track accepted articles and set up e-mail alerts to inform you when an article's status changes. The Author Gateway also provides detailed artwork guidelines, copyright information, and answers to frequently asked questions.

# ANEXO B

De:	onbehalfof+ame+dadlnet.dk@manuscriptcentral.com em Enviada: seg 31/10/2011 19:16
Para:	Karen Cherubini; carolinauv@gmail.com
Cc:	Journal of Oral Pathology and Medicine Menumint TO JODM 40.44 of 40.04
Anexos:	Journal of Oral Pathology and Medicine - Manuscript ID JOPM-10-11-OA-1894
31-Oct-20	11
Dear Prof.	Karen Cherubini,
Your man surgical w been succ Journal of requireme changes to Medicine.	uscript entitled "Comparison of effects of clodronate and zoledronic acid on the repair of maxilla ounds - Histomorphometric, RANKL, OPG, von Willebrand factor and caspase-3 evaluation" has essfully submitted online and is presently being given full consideration for publication in the Oral Pathology and Medicine. Should your manuscript not comply with the Journal's nts, however, the Journal's administrator will notify you via email that you need to make specific o your manuscript before it can be considered for publication in the Journal of Oral Pathology and
Your man	uscript ID is JOPM-10-11-OA-1894.
Please me questions. Central at	ntion the above manuscript ID in all future correspondence or when calling the office for If there are any changes in your street address or e-mail address, please log in to Manuscript http://mc.manuscriptcentral.com/jopm and edit your user information as appropriate.
You can al in to <u>http:</u> ,	so view the status of your manuscript at any time by checking your Author Center after logging //mc.manuscriptcentral.com/jopm .
Thank you	for submitting your manuscript to the Journal of Oral Pathology and Medicine.
Sincerely, Anne-Mari Administra	e Engel tor, Journal of Oral Pathology and Medicine



## Author Guidelines

**Content of Author Guidelines**: 1. General, 2. Ethical Guidelines, 3. Manuscript Submission Procedure, 4. Manuscript Types Accepted, 5. Manuscript Format and Structure, 6. After Acceptance

Relevant Documents: Copyright Transfer Agreement

**Useful Websites**: Submission Site, Articles published in Journal of Oral Pathology & Medicine, Author Services, Blackwell Publishing's Ethical Guidelines, Guidelines for Figures

The journal to which you are submitting your manuscript employs a plagiarism detection system. By submitting your manuscript to this journal you accept that your manuscript may be screened for plagiarism against previously published works.

#### **1. GENERAL**

Journal of Oral Pathology & Medicine publishes manuscripts of high scientific quality representing original clinical, diagnostic or experimental work in oral pathology and oral medicine. Papers advancing the science or practice of these disciplines will be welcomed, especially those which bring new knowledge and observations from the application of techniques within the spheres of light and electron microscopy, tissue and organ culture, immunology, histochemistry, immunocytochemistry and molecular biology. Review papers on topical and relevant subjects will receive a high priority and articles requiring rapid publication because of their significance and timeliness will be included as brief reports not exceeding three printed pages. All submitted manuscripts falling within the overall scope of the Journal will be assessed by suitably qualified reviewers, but manuscripts in an incorrect format will be returned to the author without review.

Please read the instructions below carefully for details on the submission of manuscripts, the journal's requirements and standards as well as information concerning the procedure after a manuscript has been accepted for publication in *Journal of Oral Pathology & Medicine*. Authors are encouraged to visit Wiley-Blackwell Publishing Author Services for further information on the preparation and submission of articles and figures. Note to NIH Grantees

Pursuant to NIH mandate, Wiley-Blackwell will post the accepted version of contributions authored by NIH grant-holders to PubMed Central upon acceptance. This accepted version

will be made publicly available 12 months after publication. For further information, see www.wiley.com/go/nihmandate.

# 2. ETHICAL GUIDELINES

*Journal of Oral Pathology & Medicine* adheres to the below ethical guidelines for publication and research.

# 2.1. Authorship and Acknowledgements

Authors submitting a paper do so on the understanding that the work has not been published before, is not being considered for publication elsewhere and has been read and approved by all authors.

*Journal of Oral Pathology & Medicine* adheres to the definition of authorship set up by The International Committee of Medical Journal Editors (ICMJE). According to the ICMJE authorship criteria should be based on substantial contributions to conception and design of, or acquisition of data or analysis and interpretation of data, and drafting the article or revising it critically for important intellectual content.

It is a requirement that all authors have been accredited as appropriate upon submission of the manuscript. Contributors who do not qualify as authors should be mentioned under Acknowledgements.

Acknowledgements: Under acknowledgements please specify contributors to the article other than the authors accredited. Acknowledge only persons who have made substantive contributions to the study. Authors are responsible for obtaining written permission from everyone acknowledged by name because readers may infer their endorsement of the data and conclusions.

## **2.2. Ethical Approvals**

Experimentation involving human subjects will only be published if such research has been conducted in full accordance with ethical principles, including the World Medical Association Declaration of Helsinki (version, 2002 www.wma.net/e/policy/b3.htm) and the additional requirements, if any, of the country where the research has been carried out. Manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and written consent of each subject and according to the above mentioned principles. A statement regarding the fact that the study has been independently reviewed and approved by an ethical board should also be included. Editors reserve the right to reject papers if there are doubts as to whether appropriate procedures have been used.

When experimental animals are used the methods section must clearly indicate that adequate measures were taken to minimize pain or discomfort. Experiments should be carried out in accordance with the Guidelines laid down by the National Institute of Health (NIH) in the USA regarding the care and use of animals for experimental procedures or with the European Communities Council Directive of 24 November 1986 (86/609/EEC) and in accordance with local laws and regulations.

## **2.3 Clinical Trials**

Clinical trials should be reported using the CONSORT guidelines available at www.consort-statement.org. A CONSORT checklist should also be included in the submission material.

*Journal of Oral Pathology & Medicine* encourages authors submitting manuscripts reporting from a clinical trial to register the trials in any of the following free, public clinical trials registries: www.clinicaltrials.gov, http://clinicaltrials-dev.ifpma.org/, http://isrctn.org/. The clinical trial registration number and name of the trial register will then be published with the paper.

## 2.4 Conflict of Interest

All sources of institutional, private and corporate financial support for the work within the manuscript must be fully acknowledged, and any potential grant holders should be listed. Please see Conflicts of Interest for generally accepted definitions on conflict of interest? Please enclose this information under the heading 'Conflict of Interest Statement'.

## 2.5 Appeal of Decision

Authors who wish to appeal the decision on their submitted paper may do so by emailing the editor with a detailed explanation for why they find reasons to appeal the decision.

#### **2.6 Permissions**

If all or parts of previously published illustrations are used, permission must be obtained from the copyright holder concerned. It is the author's responsibility to obtain these in writing and provide copies to the Publishers.

## 2.7 Copyright Assignment

Authors submitting a paper do so on the understanding that the work and its essential substance have not been published before and is not being considered for publication elsewhere. The submission of the manuscript by the authors means that the authors automatically agree to assign exclusive copyright to Wiley-Blackwell if and when the manuscript is accepted for publication. The work shall not be published elsewhere in any language without the written consent of the publisher. The articles published in this journal are protected by copyright, which covers translation rights and the exclusive right to reproduce and distribute all of the articles printed in the journal. No material published in the journal may be stored on microfilm or videocassettes or in electronic database and the like or reproduced photographically without the prior written permission of the publisher.

Authors will be required to sign a Copyright Transfer Agreement (CTA) for all papers accepted for publication. Signature of the CTA is a condition of publication and papers will not be passed for production unless a signed form has been received. Please note that signature of the Copyright Transfer Agreement does not affect ownership of copyright in the material. (Government employees need to complete the Author Warranty sections, although copyright in such cases does not need to be assigned). After submission authors will retain the right to publish their paper in various medium/circumstances (please see the form for further details). To assist authors, an appropriate form will be supplied by the editorial office. Alternatively, authors may like to download a copy of the form from www.wiley.com/go/ctaaglobal.

Authors must send the completed CTA upon receiving notice of manuscript acceptance, i.e., do not send the form at submission. Please email or fax the completed form back to the Production Editor (contact details below).

Melody Tan Production Editor Journal Content Management Department fax +65 6643 8599 email jop@wiley.com

## For questions concerning copyright, please visit Copyright FAQ

#### 2.8 OnlineOpen

Journal of Oral Pathology & Medicine offers authors the opportunity to publish their paper OnlineOpen. OnlineOpen is available to authors of primary research articles who wish to make their article available to non-subscribers on publication, or whose funding agency requires grantees to archive the final version of their article. With OnlineOpen, the author, the author's funding agency, or the author's institution pays a fee to ensure that the article is made available to non-subscribers upon publication via Wiley Online Library, as well as deposited in the funding agency's preferred archive. For the full list of terms and conditions, see http://wileyonlinelibrary.com/onlineopen#OnlineOpen\_Terms. Any authors wishing to send their paper OnlineOpen will be required to complete the payment form available from our website at: https://onlinelibrary.wiley.com/onlineOpenOrder. Prior to acceptance there is no requirement to inform an Editorial Office that you intend to publish your paper OnlineOpen if you do not wish to. All OnlineOpen articles are treated in the same way as any other article. They go through the journal's standard peer-review process and will be accepted or rejected based on their own merit.

For questions concerning copyright, please visit Copyright FAQ

# 3. MANUSCRIPT SUBMISSION PROCEDURE

Manuscripts should be submitted electronically via the online submission site http://mc.manuscriptcentral.com/jopm. The use of an online submission and peer review site enables immediate distribution of manuscripts and consequentially speeds up the review process. It also allows authors to track the status of their own manuscripts. Complete instructions for submitting a paper is available online and below. For further instructions, please contact Editorial Assistant Anne-Marie Engel at ame@dadlnet.dk

#### 3.1. Getting Started

• Launch your web browser (supported browsers include Internet Explorer 5.5 or higher, Safari 1.2.4, or Firefox 1.0.4 or higher) and go to the journal's online Submission Site: http://mc.manuscriptcentral.com/jopm.

• Log-in or, if you are a new user, click on 'register here'.

• If you are registering as new user.

- After clicking on 'register here', enter your name and e-mail information and click 'Next'. Your e-mail information is very important.

- Enter your institution and address information as appropriate, and then click 'Next.'

- Enter a user ID and password of your choice (we recommend using your e-mail address as your user ID), and then select your areas of expertise. Click 'Finish'.

• If you are registered as user, but have forgotten your log in details, enter your e-mail address under 'Password Help'. The system will send you an automatic user ID and a new temporary password.

• Log-in and select 'Author Centre'.

#### **3.2. Submitting Your Manuscript**

• After you have logged into your 'Author Centre', submit your manuscript by clicking the submission link under 'Author Resources'.

• Enter data and answer questions as appropriate. You may copy and paste directly from your manuscript and you may upload your pre-prepared covering letter.

- Click the 'Next' button on each screen to save your work and advance to the next screen.
- You are required to upload your files.
- Click on the 'Browse' button and locate the file on your computer.
- Select the designation of each file in the drop down next to the Browse button.

- When you have selected all files you wish to upload, click the 'Upload Files' button.

• Review your submission (in HTML and PDF format) before completing your submission

by sending it to the Journal. Click the 'Submit' button when you are finished reviewing.

## **3.3. Manuscript Files Accepted**

Manuscripts should be uploaded as Word (.doc) or Rich Text Format (.rft) files (not writeprotected) plus separate figure files. GIF, JPEG, PICT or Bitmap files are acceptable for submission, but only high-resolution TIF or EPS files are suitable for printing. The files will be automatically converted to HTML and PDF on upload and will be used for the review process. The text file must contain the entire manuscript including title page, abstract, text, references, acknowledgements and conflict of interest statement, tables, and figure legends, but *no* embedded figures. In the text, please reference figures as for instance 'Figure 1', 'Figure 2' etc to match the tag name you choose for the individual figure files uploaded. Manuscripts should be formatted as described in the Author Guidelines below. Please note that any manuscripts uploaded as Word 2007 (.docx) will be automatically rejected. Please save any .docx file as .doc before uploading.

## 3.4. Blinded Review

All manuscripts submitted to *Journal of Oral Pathology & Medicine* will be reviewed by two experts in the field. *Journal of Oral Pathology & Medicine* uses single blinded review. The names of the reviewers will thus not be disclosed to the author submitting a paper.

#### 3.5. Suggest a Reviewer

*Journal of Oral Pathology & Medicine* attempts to keep the review process as short as possible to enable rapid publication of new scientific data. In order to facilitate this process, the name and current email address of a potential international reviewer whom you consider capable of reviewing your manuscript is requested. Additionally, you may mention non-preferred reviewers as well.

#### 3.6. Suspension of Submission Mid-way in the Submission Process

You may suspend a submission at any phase before clicking the 'Submit' button and save it to submit later. The manuscript can then be located under 'Unsubmitted Manuscripts' and you can click on 'Continue Submission' to continue your submission when you choose to.

#### 3.7. E-mail Confirmation of Submission

After submission you will receive an e-mail to confirm receipt of your manuscript. If you do not receive the confirmation e-mail after 24 hours, please check your e-mail address carefully in the system. If the e-mail address is correct please contact your IT department. The error may be caused by some sort of spam filtering on your e-mail server. Also, the e-mails should be received if the IT department adds our e-mail server (uranus.scholarone.com) to their whitelist.

#### **3.8. Manuscript Status**

You can access ScholarOne Manuscripts (formerly known as Manuscript Central) any time to check your 'Author Centre' for the status of your manuscript. The Journal will inform you by e-mail once a decision has been made.

#### 3.9. Submission of Revised Manuscripts

To submit a revised manuscripts please locate your manuscript under 'Manuscripts with Decisions' and click on 'Submit a Revision'. Please remember to delete any old files uploaded when you upload your revised manuscript.

## 4. MANUSCRIPT TYPES ACCEPTED

**Original Research Articles**: of high scientific quality representing original clinical, diagnostic or experimental work in oral pathology and oral medicine. Papers advancing the science or practice of these disciplines will be welcomed, especially those which bring new knowledge and observations from the application of techniques within the spheres of light and electron microscopy, tissue and organ culture, immunology, histochemistry, immunocytochemistry and molecular biology.

**Review Papers**: *Journal of Oral Pathology & Medicine* commissions review papers and also welcomes uninvited reviews. Reviews should be submitted via the online submission site: http://mc.manuscriptcentral.com/jopm and are subject to peer-review.

# Case Reports: Please note that *Journal of Oral Pathology & Medicine* no longer accepts submissions of case reports.

**Brief Reports**: Original research material requiring rapid publication because of their significance and timeliness will be included as Brief Reports. They should not exceed three pages.

**Letters to the Editor**: Letters, if of broad interest, are encouraged. Letters should not be confused with Brief Reports. Letters may deal with material in papers published in *Journal* of Oral Pathology & Medicine or they may raise new issues, but should have important implications.

## 5. MANUSCRIPT FORMAT AND STRUCTURE

#### 5.1. Page Charge

Articles exceeding 6 published pages are subject to a charge of USD 163 per additional page. One published page amounts approximately to 5,500 characters (excluding figures and tables).

#### 5.2. Format

**Language**: The language of publication is English. Authors for whom English is a second language may choose to have their manuscript professionally edited before submission to improve the English. A list of independent suppliers of editing services can be found at http://authorservices.wiley.com/bauthor/english\_language.asp. All services are paid for and arranged by the author, and use of one of these services does not guarantee acceptance or preference for publication.

**Abbreviations, Symbols and Nomenclature**: Use only standard abbreviations (Vancouver System). All units will be metric. Use no roman numerals in the text. In decimals, a decimal point, and not a comma, will be used. Avoid abbreviations in the title. The full term for which an abbreviation stands should precede its first use in the text unless it is a standard unit of measurement. Useful is Baren DN, ed. Units, symbols, and

abbreviations. A guide for biological and medical editors and authors. 4. ed. London: Royal Society of Medicine.

**Font**: When preparing your file, please use only standard fonts such as Times, Times New Roman or Arial for text, and Symbol font for Greek letters, to avoid inadvertent character substitutions. In particular, please do not use Japanese or other Asian fonts. Do not use automated or manual hyphenation.

## 5.3. Structure

All papers submitted to *Journal of Oral Pathology & Medicine* should include: title page, abstract, main text, references and tables, figures, figure legends and conflict of interest statement where appropriate. Manuscripts must conform to the journal style. Manuscripts not complying with the journal format will be returned to the author(s).

**Title Page**: Should be part of the manuscript document uploaded for review and include: The title of the article, a running title of no more than 50 letters and spaces, 2-5 keywords, complete names and institution for each author, corresponding author's name, address, email address and fax number.

**Abstract**: is limited to 250 words in length and should contain no abbreviations. The abstract should be included in the manuscript document uploaded for review as well as inserted separately where specified in the submission process. The abstract should convey the essential purpose and message of the paper in an abbreviated form. For original articles the abstract should be structured with the following headings in accordance with Index Medicus (Medical Subject Headings): background, methods, results and conclusions. For other article types, please choose headings appropriate for the article.

Main Text of Original Articles: should be divided into introduction, material and methods, results and discussion.

**Introduction**: should clearly state the purpose of the article. Give only strictly pertinent references. Exhaustive literature reviews are inappropriate.

**Materials and Methods**: must contain sufficient detail such that, in combination with the references cited, all clinical trials and experiments reported can be fully reproduced. As a condition of publication, authors are required to make materials and methods used freely available to academic researchers for their own use. This may for example include antibodies etc. Other supporting data sets must be made available on the publication date from the authors directly.

(*i*) *Clinical trials*: Clinical trials should be reported using the CONSORT guidelines available at www.consort-statement.org. A CONSORT checklist should also be included in the submission material.

*Journal of Oral Pathology & Medicine* encourages authors submitting manuscripts reporting from a clinical trial to register the trials in any of the following free, public clinical trials registries: www.clinicaltrials.gov, http://clinicaltrials-dev.ifpma.org/, http://isrctn.org/. The clinical trial registration number and name of the trial register will then be published with the paper. .

(*ii*)*Experimental subjects*: Experimentation involving human subjects will only be published if such research has been conducted in full accordance with ethical principles, including the World Medical Association Declaration of Helsinki (version, 2002)

www.wma.net/e/policy/b3.htm) and the additional requirements, if any, of the country where the research has been carried out. Manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and written consent of each subject and according to the above mentioned principles. A statement regarding the fact that the study has been independently reviewed and approved by an ethical board should also be included. Editors reserve the right to reject papers if there are doubts as to whether appropriate procedures have been used.

When experimental animals are used the methods section must clearly indicate that adequate measures were taken to minimize pain or discomfort. Experiments should be carried out in accordance with the Guidelines laid down by the National Institute of Health (NIH) in the USA regarding the care and use of animals for experimental procedures or with the European Communities Council Directive of 24 November 1986 (86/609/EEC) and in accordance with local laws and regulations.

(*iii*) Suppliers: Suppliers of materials should be named and their location (town, state/county, country) included.

**Results**: Present your results in logical sequence in the text, tables, and illustrations. Do not repeat in the text all the data in the tables, illustrations, or both: emphasize or summarize only important observations.

**Discussion**: Emphasize the new and important aspects of the study and conclusions that follow from them. Do not repeat in detail data given in the Results section. Include in the Discussion the implications of the findings and their limitations and relate the observations to other relevant studies.

**Main Text of Review Articles** comprise an introduction and a running text structured in a suitable way according to the subject treated. A final section with conclusions may be added.

Acknowledgements: Under acknowledgements please specify contributors to the article other than the authors accredited. Acknowledge only persons who have made substantive contributions to the study. Authors are responsible for obtaining written permission from everyone acknowledged by name because readers may infer their endorsement of the data and conclusions. See also above under Ethical Guidelines.

**Conflict of Interest Statement**: All sources of institutional, private and corporate financial support for the work within the manuscript must be fully acknowledged, and any potential grant holders should be listed. Please see Conflicts of Interest for generally accepted definitions on conflict of interest? See also above under Ethical Guidelines.

#### **5.4. References**

References should be kept to the pertinent minimum and numbered consecutively in the order in which they appear in the text. Identify references in text, tables, and legends by Arabic numerals (in parentheses). References cited only in the tables or figure legends should be numbered in accordance with a sequence established by the first identification of that figure or table in the text. Use the style of the examples below, which are based on the formats used in Index Medicus. Try to avoid using abstracts as references. Include manuscripts accepted, but not published; designate the abbreviated title of the journal

followed by (in press). Information from manuscripts not yet accepted, should be cited in the text as personal communication. The references must be verified by the author(s) against the original documents. Titles should be abbreviated in accordance with the style used in Index Medicus and the Vancouver System.

We recommend the use of a tool such as EndNote or Reference Manager for reference management and formatting. EndNote reference styles can be searched for here: www.endnote.com/support/enstyles.asp . Reference Manager reference styles can be searched for here: www.refman.com/support/rmstyles.asp

Examples of the Journal's reference style:

## (1) Standard journal article

(List all authors when 6 or less; when 7 or more, list only the first 3 and add et al.) BUCHNER A, SCIUBBA JJ. Peripheral epithelial odontogenic tumors: a review. Oral Surg Oral Med Oral Pathol 1987; 63: 688-97.

HEINIC GS, GREENSPAN D, MACPHAIL LA, et al. Oral Histoplasma capsulatum infection in association with HIV infection: a case report. J Oral Pathol Med 1992; 21: 85-9.

## (2) Corporate author

European Collaborative Study. Risk factors for mother-to-child transmission of HIV-1. Lancet 1992; 339: 1007-12.

## (3) No author given

Anonymous. 'The importance of being early' [leader]. Br Dent J 1991; 170: 167.

## (4) Journal supplement

MØLLER-PETERSEN J. Evaluation of diagnostic tests. Design and phases. Scand J Clin Lab Invest 1992; 52: suppl. (208): 35-50.

CROSS SS, SCHOLFIELD JH, KENNEDY A, COTTON DWK. Measuring the fractal dimension of tumour borders. J Pathol 1992; 168: 117A (abstr).

#### (5) Journal paginated by issue

HILLAM C. Dentistry in Europe in the 1790's. Dent Historian 1992; 22: (May): 31-4.

## (6) Book

PINDBORG JJ. Atlas of diseases of the oral mucosa. Copenhagen: Munksgaard, 1992: 50-66.

(7) Chapter in a bookVAN DER WAAL I. Salivary gland neoplasms. In: PRABHU SR, WILSON DF, DAFTARY DK, JOHNSON NW, eds. Oral diseases in the tropics. Oxford: Oxford Medical, 1992; 478-86.

#### (8) Published proceedings paper

DRINNAN AJ. Review of the literature: educational aspects of oral medicine. In: MILLARD HD, MASON DK, eds. World workshop on oral medicine. Chicago: Year Book Medical, 1989; 5-11.

## (9) Agency publication

MUIR C, WATERHOUSE J, MACK T, POWELL J, WHELAN S. Cancer incidence in five continents: Vol. 5. Lyon: International Agency for Research on Cancer, 1987; IARC Scientific Publications No. 88.

## (10) Dissertation or thesis

CHUNGPANICH S. The diagnostic and prognostic potential of nucleolar organizer regions in oral epithelial dysplasia. MMedSci Thesis, University of Sheffield, 1989.

## 5.5. Tables, Figures and Figure Legends

**Tables**: should be numbered consecutively with Arabic numerals. Type each table on a separate sheet, with titles making them self-explanatory. Due regard should be given to the proportions of the printed page.

**Figures**: All figures should clarify the text and their number be kept to a minimum. Text on figures should be in CAPITALS. Line drawings should be professionally drawn; half-tones should exhibit high contrast.

All figures and artwork must be provided in electronic format. Figure legends should be a separate section of the manuscript, and should begin with a brief title for the whole figure and continue with a short description of each panel and the symbols used: they should not contain any details of methods.

Submit your figures as EPS, TIFF or PDF files. Use 300 dpi resolution for photographic images and 600 dpi resolution for line art.Full details of the submission of artwork are available at http://authorservices.wiley.com/bauthor/illustration.asp.

## 6. AFTER ACCEPTANCE

#### 6.1. Copyright

A completed Copyright Transfer Agreement (CTA), found at www.wiley.com/go/ctaaglobal must be received by Production Editor before any manuscript can be published. Authors must send the completed original CTA by regular mail upon receiving notice of manuscript acceptance, i.e. do not send the CTA at submission.

## 6.2 Proofs

Proofs will be sent via e-mail as an Acrobat PDF (portable document format) file. The e-mail server must be able to accept attachments up to 4 MB in size. Acrobat Reader will be required in order to read this file.

#### 6.3 Early View

*Journal of Oral Pathology & Medicine* is covered by Wiley-Blackwell Publishing's Early View service. Early View articles are complete full-text articles published online in advance of their publication in a printed issue. Early View articles are complete and final. They have been fully reviewed, revised and edited for publication, and the authors' final corrections have been incorporated. Because they are in final form, no changes can be made after online publication. Early View articles are given a Digital Object Identifier (DOI), which allows the article to be cited and tracked before it is allocated to an issue. After print publication, the DOI remains valid and can continue to be used to cite and access the article.

#### **6.4 Offprints**

The corresponding author will receive a free PDF offprint that can be downloaded via Author Services. Please sign up for the service if you would like to access your free article PDF offprint and enjoy the many other benefits the service offers. Visit http://authorservices.wiley.com/bauthor for more information.

## 6.5 Author Services

Online production tracking through Wiley-Blackwell's Author Services Author Services enables authors to track their article - once it has been accepted - through the production process to publication online and in print. Authors can check the status of their articles online and choose to receive automated e-mails at key stages of production. The author will receive an e-mail with a unique link that enables them to register and have their article automatically added to the system. Please ensure that a complete e-mail address is provided when submitting the manuscript. Visit http://authorservices.wiley.com/bauthor for more details on online production tracking and for a wealth of resources including FAQs and tips on article preparation, submission and more.

ANEXO C

Comissão Científica e de Ética Faculdade da Odontologia da PUCRS Porto Alegre 08 de de 2009 janeiro O Projeto de: Tese Protocolado sob nº: 0105/08 Avaliação macro e microscópica de lesões orais induzidas por Intitulado: procedimentos cirúrgicos em ratos sob terapia com bisfosfonatos Pesquisador Responsável: Profa. Dra. Karen Cherubini Pesquisadores Associados Ana Carolina Uchoa Vasconcelos Nível: Doutorado Foi aprovado pela Comissão Científica e de Ética da Faculdade de Odontologia da PUCRS em 07 de janeiro de 2009. Este projeto deverá ser imediatamente encaminhado ao CEUA/PUCRS 00 Prof. Dr. Eraldo Luiz Batista Júnior Presidente da Comissão Científica e de Ética da Faculdade de Odontologia da PUCRS Fone/Fax: (51) 3320-3538 Av. Ipiranga, 6681, Prédio 06 sala 209 Porto Alegre /RS – Brasil – Cx. Postal:1429 90619-900 e-mail: odontologia-pg@puers.br

#### **ANEXO D**



Pontifícia Universidade Católica do Rio Grande do Sul CEUAs PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO COMITÊ DE ÉTICA PARA O USO DE ANIMAIS



Ofício 043/09 - CEUA

Porto Alegre, 16 de abril de 2009.

Senhora Pesquisadora:

O Comitê de Ética para o Uso de Animais apreciou e aprovou seu protocolo de pesquisa, registro CEUA 09/00083, intitulado: "Avaliação macro e microscópica de lesões orais induzidas por procedimentos cirúrgicos em ratos sob terapia com bisfosfonatos".

Sua investigação está autorizada a partir da presente data.

Relatórios do andamento do projeto devem ser entregues a este Comitê.

Atenciosamente,

nand Profa. Dr. Anamaria Feijó Coordenadora do CEUA - PUCRS

Ilma. Sra. Profa. Dra. Karen Cherubini Faculdade de Odontologia N/Universidade

**Campus Central** Av. Ipiraga, 6690 – Prédio 60 sala 314 CEP: 90610-000 Fone/Fax: (51) 3320-3345 E-mail: <u>ceua@pucrs.br</u>