ORIGINAL ARTICLE



Something is not quite right: Effects of two land uses on anuran diversity in subtropical grasslands

Leonardo Felipe Bairos Moreira¹ \bigcirc | Henrique Zanette de Castilhos² | Santiago Castroviejo-Fisher²

¹Programa de Pós-Graduação em Ecologia e Conservação da Biodiversidade, Instituto de Biociências, UFMT–Universidade Federal de Mato Grosso, Cuiabá, Brazil

²PUCRS–Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre, Brazil

Correspondence

Leonardo Felipe Bairos Moreira, Instituto de Biociências, Universidade Federal de Mato Grosso, Cuiabá CEP-78060-900, Brazil. Email: leonardobm@gmail.com

Funding information

Conselho Nacional de Desenvolvimento Científico e Tecnológico, Grant/Award Number: 442987/20; Coordenação de Aperfeiçoamento de Pessoal de Nível Superior, Grant/Award Number: 20132816

Associate Editor: Ferry Slik Handling Editor: Ulmar Grafe

Abstract

Although habitat modification is considered one of the main causes of biodiversity loss, the relative contribution of different rural land uses to biodiversity conservation is far less known. Additionally, the realization of the multidimensionality of biodiversity demands studies integrating variation of functional traits and phylogenetic information as complements to address the effects of land use on the structure of animal communities. Herein, we investigated the effects of land use (i.e., intensive agricultural and extensive livestock rearing) on functional and phylogenetic diversity of anuran communities in farmland ponds from the Uruguayan savanna ecoregion, while considering the effects of local factors (i.e., water depth) on species composition. We surveyed adults and tadpoles in 22 ponds and quantified five traits related to tadpole feeding, habitat use, and predator avoidance. Tadpole identification was corroborated by DNA barcoding based on a fragment of the mitochondrial 16S rRNA gene. We observed a decline in phylogenetic mean nearest taxon distance associated with increase of surrounding agricultural land use. While land use intensification did not affect richness (functional or phylogenetic), ponds in livestock ranches hosted about four times more tadpoles than agricultural ponds. Functional evenness decreased with water depth, although such relationship disappeared when considering phylogenetic non-independence. Our results indicated that specific anuran clades were more sensitive to intensification in land use, reinforcing a recent view of phylogenetic homogenization following habitat conversion. Additionally, our study suggests that extensive cattle grazing over wide native pastures may provide an alternative more compatible with conservation than short-term crops in subtropical grasslands.

Abstract in Portuguese is available with online material.

KEYWORDS

agriculture, community phylogenetic, livestock, Pampa, tadpoles

© 2020 The Association for Tropical Biology and Conservation

1 | INTRODUCTION

Habitat modification is the primary driver of population declines and species extinctions (Kehoe et al., 2015), with food production through agriculture and cattle ranching—as one of the main causes of habitat change (Veach, Moilanen, & Di Minin, 2017). In landscapes where the conversion has already taken place, agricultural intensification may enhance biodiversity loss through increased mechanization, decreased crop diversity, input of hazardous pesticides, and grassland conversion to arable fields (Schiesari & Corrêa, 2016; Tscharntke, Klein, Kruess, Steffan-Dewenter, & Thies, 2005). After conversion to croplands, vegetation homogenization and a

Iterations in the hydrological regimes are the immediate impacts on freshwater communities. Thus, across a wide range of animal and plant taxa, it is unsurprising that pristine areas support greater levels of biodiversity than agricultural or pastoral lands (Flynn et al., 2009; Tscharntke et al., 2005).

When compared to intensively used agroecosystems, the relative contribution of non-intensively exploited areas to conservation is well acknowledged (Phillips, Newbold, & Purvis, 2017). Planted pastures may support moderate levels of biodiversity in relation to intensive crops, although pastures are often far less suitable than forested areas (Schiesari & Corrêa, 2016). However, exotic planted pastures occur by removing the original vegetation-often transforming forest and woodland ecosystems-and establishing exotic species (Moreira, Solino-Carvalho, Strüssmann, & Silveira, 2016). This is contrastingly different to using natural grasslands for moderate or extensive domestic animal grazing. Despite the importance of natural grasslands as ecosystems and their role in human economic activities, their relevance as economically productive systems that allow for the conservation of a more diverse biota, depending on management practices, has received little attention in the literature (Dotta, Phalan, Silva, Green, & Balmford, 2016; Isacch, Maceira, Bo, Demaría, & Peluc, 2005). Negative effects of grazing on freshwater communities result from direct nitrogenous waste input into ecosystems and changes in vegetation structure (Jansen & Healey, 2003; Schmutzer, Gray, Burton, & Miller, 2008). Recent reviews show no clear consensus about responses to livestock use of wetlands, although potential negative (even mixed or positive) effects seem dependent on grazing regimes and stocking densities (Howell et al., 2019; Schieltz & Rubenstein, 2016). Therefore, we identify a need for more integrative studies based on empirical data to understand how the biodiversity of open areas, in its different dimensions (e.g., functional and phylogenetic), copes with a changing habitat. More importantly, we should understand how different land uses modify natural grasslands-which uses have more pervasive impacts and which uses help to supplement the biodiversity conserved in protected areas.

The recognition that species trait diversity may contribute disproportionately to ecosystem functioning has led to new insights about ecological patterns and ecosystem health (Cadotte, Carscadden, & Mirotchnick, 2011). Organism's traits have direct and indirect consequences for its fitness, making functional traits a proxy

for understanding the environmental tolerances and habitat requirements (Mouillot, Graham, Villéger, Mason, & Bellwood, 2013). Levels of trait variation (i.e., functional richness) are expected to decrease with high disturbance levels, while functional evenness which incorporates both species traits and abundance—decreases even under moderate levels of disturbance (Mouillot et al., 2013; Villéger, Miranda, Hernández, & Mouillot, 2010). This happens because functional evenness reflects the balance between competitive interactions (at low disturbance levels) and trait filtering associated with the increase of disturbance intensity, while functional richness decreases with species extinctions with extreme traits (Mouillot et al., 2013).

Animal functional traits that describe food acquisition and habitat use may influence species composition in modified landscapes (Flynn et al., 2009; Trimble & van Aarde, 2014). Although trait-based approaches might be a valuable tool, trait data are not readily available or the investigated traits are hard to quantify. Methodological issues remain to establish the underlying mechanisms driving community assembly, including trait selection and types of functional diversity measures (Schmera, Heino, Podani, Erös, & Dolédec, 2017; Tsianou & Kallimanis, 2016). As an alternative, phylogenetic diversity metrics have been used as a surrogate for functional diversity (Gerhold, Cahill, Winter, Bartish, & Prinzing, 2015), based on the general assumption that proximity of common ancestry correlates with niche similarity. However, the generality of this assumption is contentious and commonly found processes in empirical studies, such as evolutionary convergence, ecological speciation, and phenotypic plasticity, will distort such associations (Cadotte, Davies, & Peres-Neto, 2017; Stroud & Losos, 2016).

While several studies have addressed the effects of land use intensification on functional and phylogenetic diversity (Lee & Martin, 2017; Ribeiro et al., 2017; Trimble & van Aarde, 2014), there are important knowledge gaps on the impacts of land use in grasslands and freshwater ecosystems. In particular for amphibians, phylogenetic homogenization following habitat conversion seems to be a global pattern (Nowakowski, Frishkoff, Thompson, Smith, & Todd, 2018), although local responses to conversion can vary widely in the functional context. Function-related responses to land use seems to be complex; modified habitats often show less functional groups, yet not all human land uses affect functional richness (Díaz-García, Pineda, López-Barrera, & Moreno, 2017; Riemann, Ndriantsoa, Rödel, & Glos, 2017; Trimble & van Aarde, 2014). In subtropical grasslands, biodiversity erosion has been documented under moderate levels of habitat loss. Such agricultural landscapes had fewer number of species, lower phylogenetic diversity, and higher nestedness (Saccol, Bolzan, & Santos, 2017; Staude et al., 2018), in which disturbance-tolerant species are favoured and non-random local extinctions occur in some evolutionary lineages.

Here, we investigated the relationship between anuran diversity and land use (intensive agricultural areas versus extensive livestock rearing) in farmland ponds from the Uruguayan savanna ecoregion. We considered two aspects of functional diversity (richness and evenness) and two aspects of phylogenetic diversity (richness

and divergence). Explicitly, we explored whether: (a) there are different amounts of evolutionary history represented by the anuran communities at each land use; (b) in a phylogenetic context, species within ponds surrounded by cultivated land would be more related than expected in a random assemblage, reflecting the role of environmental filters in selecting disturbance-tolerant species; (c) functional diversity patterns, based on tadpole traits related to food acquisition, habitat use, and predator avoidance would be linked to different land uses. We posit that functional evenness would decline with land use intensity, but without significant change in functional richness. Intensive agricultural areas are subject to physical (native vegetation removal and crop harvest) and chemical (pesticides and fertilizers) disturbances, whereas extensive cattle grazing leads to a change in grassland structure (Moreira, Solino-Carvalho, et al., 2016; Schiesari & Corrêa, 2016). In this way, extensive livestock ranches would support moderately higher levels of biodiversity in relation to more impoverish agricultural areas.

2 | METHODS

2.1 | Study area and data collection

We selected 22 ponds in southern Brazil (29.64°–30.10°S, 54.05°– 57.33°W), each with adjacent areas of intensive cultivated land (N = 11) or land used for extensive rearing livestock through grazing (N = 11). The Brazilian Pampa corresponds to the northern portion of the Uruguayan savanna ecoregion (Figure 1), which includes a mosaic of grasslands, scrub savannas, and gallery forests. Cattle grazing and fire have shaped this landscape for over 300 years (Bernardi, Holmgren, Arim, & Scheffer, 2016; Overbeck et al., 2007). But since the second half of the twentieth century, conversion of native grasslands to more profitable agricultural activities, mainly rice and soybean, have greatly expanded (Oliveira et al., 2017). Nowadays, livestock farming (65%) and intensive crop-based agriculture (20%) dominate the land use patterns (Modernel et al., 2016).

In the spring of 2015 (10-16 October), we sampled tadpoles in ponds located 5.5-316 km apart (Figure 1). Most anuran species in the Uruguayan savanna were spring breeders or overwinter as tadpoles (Maneyro & Carreira, 2012). We selected small ponds surrounded by similar land use up to 500 m (crop or livestock). Ponds were close to 0.15 ha (range 0.01-0.35), although one large pond was included in the sampling (0.76 ha). Sampled ranches raise cattle and sheep in natural grasslands (i.e., area for livestock grazing mostly formed by non-planted pastures with negligible amounts of external inputs). Stocking rates estimates range from 0.68 to 1.0 animal units/ha, although ranchers could adjust it according to the winter conditions and overgrazing may occur (Modernel et al., 2016). Intensive agricultural areas were used for short-term crops (sovbean or rice). Application of agrochemical, inorganic fertilizers, and glyphosate-based herbicides is concentrated in the initial growth stage (October-November). In order to ensure that the landscape composition surrounding each pond did not change over different scales, we also defined circular areas (1,000 m radius) and measured land-cover data (Table S1), using Qgis 2.18.16 (QGIS Development Team, 2018). We based analysis on Google Earth imagery, using a land-cover classification for 2015 produced within the scope of the MapBiomas initiative (Project MapBiomas, 2019). Other land uses (natural forests, forest plantation, and mosaics) showed small areas within buffers (Table S1), and there was a strong negative correlation between natural grasslands used for cattle ranching and intensive agricultural area (r = -.972). Thus, we used agricultural area as predictor variable inversely expressing livestock farming.

At each pond, we performed eight dip-net sweeps (30 cm diameter, 2 mm mesh), each covering approximately 1 m^2 and distributed at different pond depths (i.e., four sweeps near the edge of the pond



FIGURE 1 Location of Uruguayan savanna (dark gray) and the study sites in southern Brazil. Solid circles represent agricultural ponds and open circles livestock ranch ponds

and four sweeps in deep water near the center of the pond). Sweeps were pooled into one sample per pond (3.5 L plastic bucket), where the collected tadpoles were euthanized with a benzocaine solution and sorted into series based on their morphology (i.e., coloration, body shape, eyes position, tail fin proportion, and mouth position). From each series, tail samples were taken from three specimens to corroborate tadpole identification by DNA barcoding based on a fragment of the mitochondrial 16S rRNA gene (Grosjean, Ohler, Chuaynkern, Cruaud, & Hassanin, 2015). We also registered the presence of all observed adults, visually or acoustically, during tadpole sampling. Voucher specimens were deposited at the Amphibian Collection of the Museu de Ciências e Tecnologia da Pontifícia Universidade Católica do Rio Grande do Sul (MCT-PUCRS), Brazil.

Five functional traits were selected to reflect tadpole feeding, habitat use, and predator avoidance (Azizi, Landberg, & Wassersug, 2007: Strauss, Reeve, Randrianiaina, Vences, & Glos, 2010; Venesky, Wassersug, & Parris, 2010). To accommodate differences across tadpole developmental stages, all traits were ratios between morphological measures (Table 1). Each specimen was rinsed in distilled water and then measured for a set of eight morphological traits on an automated stereomicroscope-Leica M205A (Figure S1). Morphological terminology followed that of Altig and McDiarmid (1999), and only specimens without tail injuries between Gosner' stages 29 and 39 were measured. Phylogenetic relationships among species were obtained from a time-calibrated phylogenetic tree generated by Pyron (2014). We pruned the phylogeny to match the species pool of both land uses. When resolution for our taxa was not present in the phylogeny, we manually incorporated our species following a range of published sources for between-species relationships (Figure S2). Mean path length calibration was used to adjust branch lengths (Britton, Oxelman, Vinnersten, & Bremer, 2002).

-DIOTROPICA

2.2 | Diversity measures and phylogenetic signal

Some degree of phylogenetic signal in functional traits is common for taxa that share a common ancestor, so we quantified the phylogenetic signal in the traits using Blomberg's K statistic implemented in the phytools package (Revell, 2012) in R. To summarize anuran phylogenetic variation, we chose metrics related to richness and divergence: (a) Faith's phylogenetic diversity (PD) to represent the sum of accumulated phylogenetic differences (i.e., sum for a total branch lengths for the species occurring in each pond); (b) mean nearest taxon distance (MNTD) to represent the mean phylogenetic relatedness within an assemblage (i.e., mean of the branch lengths separating each species from its closest relative in each pond).

To quantify changes in functional trait variation across livestock ponds and crop ponds, we calculated two functional metrics: functional richness (FRic) and functional evenness (FEve) (Villéger, Mason, & Mouillot, 2008). The former describes a multidimensional trait space filled by the species assemblage, while the latter reflects the regularity of abundance distribution in the functional trait space. Functional richness is independent of species abundance, while FEve value is proportional to species relative abundance. Both functional metrics were calculated for all ponds with a minimum of three recorded species. To facilitate comparison between the different metrics (functional and phylogenetic), the magnitude of the differences was calculated based on standardized effect size (SES; Gotelli & McCabe, 2002). Standardized effects were derived from a null model that preserves regional species occurrence frequency and pond species richness (independent swap algorithm; Gotelli, 2000). Positive or negative SES. PD/FRic values indicate, respectively, phylogenetic/functional richness higher or lower than expected by the null model. For

Trait	Measure	Relevance
Body form	BL/TL	Swimming type and endurance. High values entail lateral bending during swimming, evading attacking predators. Low values entail little lateral flexion, enabling more endurance in higher flow velocity
Tail shape	(DTH + VTH)/TMH	Position in the water column. Deep tail fins may distract the predator, deflecting deadly strikes to the body wall
Tail position	TAL/BL	Acceleration and maneuverability. Larger tails enhance sprint swimming speed of tadpoles
Oral disk position	OD/BL	Food acquisition. Low values entail feeding mainly on midwater or surface, while high values entail feeding on pond bottom
Number of tooth rows	TR/OD	Food acquisition and substrate anchorage. Low values entail suspension feeders and midwater macrophagous tadpoles. More labial tooth rows enable tadpoles to feed on other resources by scraping or biting off material from substrate (macrophytes and carrion)

Abbreviations: BL, body length; DTH, dorsal tail fin height; OD, oral disk width; TAL, tail length; TL, total length; TMH, tail muscle height; TR, tooth rows number; VTH, ventral tail fin height.

 TABLE 1
 Traits and measurements

 used to calculate functional diversity
 indices

WILEY DIOTROPICA

	Сгор		Livestock ranch	
Species	Occurrence	Abundance	Occurrence	Abundance
Bufonidae				
Melanophryniscus atroluteus	1 ^a	7	1 ^a	
Rhinella diptycha			5ª	
Rhinella dorbignyi	3ª	1	3ª	1
Hylidae				
Boana pulchella	4	22	7 ^a	35
Dendropsophus minutus	1	3	4 ^a	27
Dendropsophus sanborni	1 ^a	1	3ª	4
Ololygon aromothyella			3ª	
Pseudis minuta	1 ^a	6	7 ^a	5
Scinax fuscovarius			3ª	
Scinax nasicus	1	9		
Scinax squalirostris	5ª	21	9 ^a	128
Leptodactylidae				
Leptodactylus gracilis	4 ^a	14	5ª	41
Leptodactylus latinasus	2 ^a	2	4 ^a	
Leptodactylus latrans			5ª	432
Physalaemus biligonigerus	4 ^a	15	1	6
Physalaemus cuvieri	3ª		2 ^a	
Physalaemus gracilis	4	4	2 ^a	1
Physalaemus henselii			2 ^a	
Physalaemus riograndensis	5ª	42	6 ^a	8
Pseudopaludicola falcipes	7 ^a	25	11 ^ª	19
Microhylidae				
Elachistocleis bicolor	2ª	2	5ª	8
Odontophrynidae				
Odontophrynus americanus			2	14

TABLE 2 Species occurrence and tadpole abundance registered in 22 ponds distributed in different land uses (crop N = 11, livestock N = 11) of the Uruguayan sayanna

^aSpecies also registered by calling activity of adults.

SES.MPD/FEve, positive significant values indicate clustering, and negative values indicate phylogenetic/functional overdispersion. Phylogenetic and functional diversity measures were computed using the picante (Kembel et al., 2010) and FD (Laliberté, Legendre, & Shipley, 2014) packages in R.

2.3 | Statistical analysis

We assessed sampling saturation overall and per land use with rarefaction and extrapolation curves with Hill numbers (Chao et al., 2014). We also estimated species richness per land use with two non-parametric richness estimators: one abundance-based (Chao 1) for tadpole data and one incidence-based (Chao 2) that include also anuran species identified from vocalizations. Sampling curves were generated using the iNext package (Hsieh, Ma, & Chao, 2016) in R.

Linear models with forward selection (Blanchet, Legendre, & Borcard, 2008) were used to evaluate the effect of pond characteristic (water depth) and the landscape effect (agricultural area) on SES.PD/MNTD or SES.FRic/FEve. Phylogenetic signal in traits may inflate type I error in the functional metrics used here (Diniz-filho, Cianciaruso, Rangel, & Bini, 2011; Duarte, Debastiani, Carlucci, & Diniz-Filho, 2018). So, we performed a phylogenetic correction for functional traits in two steps: (a) performing phylogenetic generalized least squares (PGLS) on each trait modeled only by its mean and variance; (b) taking the model normalized residuals as phylogeny-free estimates of trait variation (so-called S-component). These residuals were used to calculate phylogeny-free SES.FRic/FEve that was regressed on agricultural area and water depth. Finally, we performed a test for spatial autocorrelation in the residuals from all linear models, using Moran's I statistic implemented in the spdep package (Bivand & Piras, 2015) in R.

3 | RESULTS

Summarizing data over all sampled ponds, we found tadpoles of a total of 17 anuran species, representing 11 genera from five families (Table 2). Five additional anuran species were registered visually or acoustically as adults only (Ololygon aromothyella, Physalaemus cuvieri, P. henselii, Rhinella diptycha, and Scinax fuscovarius), bringing the total number of observed species to 22. Functional diversity indices were calculated from 294 individual measurements of five traits of the 17 species from the 22 ponds-note that many specimens had tail injuries and could not be included. We produced 67 barcodes (GenBank codes: MT460167-MT460233), corresponding to specimens of all species and tadpoles' series (MCP 13526-13602), that were used to confirm morphological identifications by comparing them with homologous fragments available in GenBank. Ponds in livestock ranches hosted about four times more tadpoles than agricultural ponds (Table 3). Across all study ponds, the most abundant species was Leptodactylus latrans, corresponding to 48% of all the collected individuals (Table 2). Even after the exclusion of L. latrans, livestock ranches hosted around two times more tadpoles than agricultural ponds. Species accumulation curves approached but did not reach an asymptote for anurans overall or any land use (Figure S3). Confidence interval of both indicators did not differ significantly between land uses (Table 3).

We observed a considerable degree of phylogenetic signal for most measured traits, indicating conserved evolution for three traits (Table S2). Overall, diversity metrics exhibited random distributions, with few values being significantly less than expected from the null model (Table S3). Only one cattle pond showed a significant low value of SES.FRic. Many crop ponds exhibited low values of SES.MNTD (N = 7), but only two ponds had significant low values, indicating environmental filtering. Regarding SES. FEve, one cattle pond showed functional clustering and one crop pond showed overdispersion (Table S3). Effect size of PD was not related to water depth or landscape, while MNTD was significantly related to land use intensification (Table 4). Average distances between each species and its nearest phylogenetic neighbor in the community decreased with agricultural area (Figure 2a). Variation in FRic was not explained by any of the measured variables while functional evenness was related to water depth (Figure 2b). However, when corrected by phylogenetic proximity, we found no significant relationship (Table 4). Considering that our estimate of FEve could be biased, due to the large abundance of L. latrans, we excluded this species and reanalyzed the data, but the

overall patterns remained (Figure S4). None of the model residuals showed evidence of spatial autocorrelation (Table 4).

4 | DISCUSSION

In line with accumulating evidence of ongoing biodiversity erosion in subtropical grasslands (Egorov et al., 2014; Saccol et al., 2017; Staude et al., 2018), our results showed that anuran communities respond negatively to land use intensification (i.e., conversion from native grasslands to intensive croplands). Although we found no relationship between richness (functional or phylogenetic) and land use, our results show that ponds surrounded by intensive crops contain species that are closely related to one another. This can be linked to phylogenetic homogenization, in which specific anuran clades were more sensitive to disturbances associated with agriculture. Functional evenness decreased with water depth, although this was probably a statistical artifact generated by phylogenetic non-independency among species.

In our study, ponds in the sampled properties harbored half of the anuran species richness normally found in conservation areas of the Brazilian Pampa (André, Cechin, & Santos, 2019; Bolzan, Saccol, & Santos, 2016) and about 30% of the species registered at the Uruguayan savanna ecoregion (Canavero et al., 2010; Maneyro, 2008). Such impoverishment could be due to a sampling effect, because of low detection probabilities of many anuran species in the region (Moreira, Moura, & Maltchik, 2016). However, our results agree with other estimates of total richness at Pampa farmlands that were not based on a single sampling period (Bolzan, Hartmann, & Hartmann, 2014; Machado & Maltchik, 2010; Moreira & Maltchik, 2015; Saccol et al., 2017). Together, these studies revealed a complementary pattern in relation to the composition of anuran assemblages in the Pampa biome. Many species that could be considered as generalists have unexpected gaps in their distribution. Such idiosyncrasies are commonly attributed to local vegetation variations associated with soil heterogeneity, topography, and land use (Bolzan et al., 2014; Lipinski & Santos, 2014). In spite of the extensive geographic extension of the Uruguayan savanna ecoregion, few areas within it are protected by conservation units. It is interesting to note that the species registered as adults only have the southern limit of their geographical distribution in the border between Brazil and Uruguay (P. cuvieri and S. fuscovarius; Maneyro, 2008; Maneyro & Carreira, 2012) or have low frequency of occurrence in the region (O. aromothyella, P. henselii, and R. diptycha; Bolzan et al., 2014; Bolzan et al., 2016; Santos, Kopp, Spies,

TABLE 3 Amphibian species richness(number in parentheses include audiosurveys), tadpole abundance, andabundance/incidence-based richnessestimators across different land uses

	Species registered	Total abundance	Chao 1 (95% CI)	Chao2 (95% Cl)
Total	17 (22)	903	17 (17–18.3)	22.1 (22-25.4)
Crop	15 (16)	174	16 (15.1–26)	21.7 (16.9–42)
Livestock	14 (21)	729	15 (14.1-28.1)	21.4 (21–26.8)

1292 WILEY DIOTROPICA

Coefficients Moran's I Adj. R² Variable $F(_{df})$ Area Depth (p-values) Phylogenetic diversity SES.PD ≈0 0.512(219) 0.93 (.16) SES.MNTD 0.20* -1.44 0.67 (.25) 6.811(2 19) Functional diversity SES.FRic 0.07 $1.674(_{217})$ -0.57 (.72) 0.593(2.17) SES.FRic (S-component) ≈0 0.7 (.24) 4.890(2.17) SES.FEve 0.17 -0.03 0.26 (.39) SES.FEve (S-component) ≈0 0.302(217) -0.44 (.67) TABLE 4Linear models relatingphylogenetic and functional diversity toagricultural area and pond depth in theUruguayan savanna, Brazil

Note: S-components control for phylogeny non-independency.

*p < .05.

Trevisan, & Cechin, 2008). Moreover, they display an explosive reproduction strategy and are usually recorded after heavy rainfall events (>50 mm/day).

The decline in MNTD associated with ponds inside a large matrix of croplands indicates that they tend to harbor a greater number of recently divergent species. This result reflects the greater occurrence of foam nest species (family Leptodactylidae). Indeed, it seems that leptodactylids are more tolerant to habitat conversion (Machado & Maltchik, 2010; Nowakowski et al., 2018). Reproductive modes associated with foam nests, floating on pond or inside subterranean constructed nests, enable adults and tadpoles to overcome several drawbacks that are common in open environments: desiccation risk, thermal damage, difficulty in oxygen exchange, and microbial colonization (Fleming, Mackenzie, Cooper, & Kennedy, 2009; Hissa et al., 2008; Zina, 2006). Moreover, foam nest has been pointed at as a key innovation in Leptodactylidae, increasing diversification rates in lineages with it (Pereira et al., 2017). Yet, not all leptodactylids respond similarly to land use constraints, and foam nest is only one of the various potentially relevant life history traits involved (Medina, Ponssa, & Aráoz, 2016). For example, the most abundant species in our sampled ponds (L. latrans) appears to be well adapted to habitat modification (Heyer et al., 2010). However, recent studies have also reported low occurrence of L. latrans in areas surrounded by agricultural fields (Moreira & Maltchik, 2015; Prado & Rossa-Feres, 2014). Tadpoles of this species have schooling behavior, and relative levels of water availability seems to be an important factor affecting tadpole survival-L. latrans tadpoles select deeper microhabitats covered by clay substrate (Melo, Garey, & Rossa-Feres, 2018). Agricultural runoff and changes in the water availability associated with crop cycle can be particularly challenging for this species and others with similar behavior. To elucidate why clades other than Leptodactylidae are less frequent in agricultural ponds is a tricky task, because of synergetic effects among various co-occurring stressors affecting amphibians. Toxic stress has consequences on amphibian survival and susceptibility to parasites (Hua et al., 2017; Peltzer et al., 2013), but there is no consensus about the existence of phylogenetic signal

in the impact of agrochemicals (Egea-Serrano, Relyea, Tejedo, & Torralva, 2012; Guenard, Carsten von der Ohe, Carlisle Walker, Lek, & Legendre, 2014).

Several studies have stressed the role of depth gradients enabling different guilds of anuran larvae to co-occur in the same habitat (Both, Cechin, Melo, & Hartz, 2011; Melo et al., 2018). Such link between tadpole guild and water depth is commonly invoked to explain responses to habitat alteration. For instance, Queiroz, da Silva, and Rossa-Feres (2015) suggested that some traits (triangular bodies, high dorsal and ventral fins, and the presence of flagella) enhanced tadpole performance in medium depth ponds (40-70 cm deep) in agricultural landscapes of the Cerrado ecoregion. Thus, nektonic species with such traits would be poorly adapted to shallow or deep waters in ponds. Although human induced changes are often correlated with amphibian functional diversity, decoupling observed patterns from phylogenetic autocorrelation is not a common practice. While other studies found anatomical modifications associated with some strata in the water column, we did not find a signal of changes in functional evenness when controlled for phylogenetic relatedness among species. In this sense, our results reinforce the idea that phylogenetic relationships have to be considered in analyses trying to correlate functional diversity with environmental variation or land use changes.

An important point here is that the presence of tadpoles depends on breeding-site preferences of adults (Both, Melo, Cechin, & Hartz, 2011). However, amphibian adults and tadpoles live in different contexts and land use may have distinct effects for each stage. Extensive cattle production of Uruguayan savanna is associated with native grasslands (Oliveira et al., 2017) and despite unsuitable management problems (i.e., overgrazing and erosion), traditional extensive livestock rearing have maintained many areas of Uruguayan savanna as grasslands with low level of disturbance (Oliveira et al., 2017; Overbeck et al., 2007). On the other hand, conversion to intensive agricultural lands is followed by a homogenization of plant communities and water physicochemical properties (Schiesari & Corrêa, 2016; Staude et al., 2018). Relative benefits and drawbacks of agroecosystems to amphibians are often crop dependent (Cosentino, Schooley, & Phillips, 2011), but common negative



FIGURE 2 Relationship between phylogenetic/functional measures and environmental descriptors. (a) Mean phylogenetic relatedness (SES MNTD: standard effect size for mean nearest taxon distance) and landscape agricultural area; (b) functional evenness (SES FEve: standard effect size for functional evenness) and pond water depth. Hatched lines represent the 95% confidence boundaries

Water depth (cm)

40

30

impacts result from the input of agrochemicals, periodic land preparation, and pond water consumption (Mann, Hyne, Choung, & Wilson, 2009; Schiesari & Corrêa, 2016). Thus, part of the lack of an influence of the land use on the tadpole functional metrics might be derived from constraints influencing juveniles and adults, because tadpoles do not choose the pond where they will live—only the microhabitat they use within a pond.

20

10

Studies that investigate community assembly in amphibians have observed divergent results according to the used null model (Both, Melo, et al., 2011; Moreira & Maltchik, 2012; Tsianou & Kallimanis, 2019). Our choice of null model was based on a specific biological hypothesis (i.e., habitat filtering) and statistical properties (i.e., low type I error rates). However, the performance of null models against different assembly processes is an ongoing debate and may depend on the choice of the metric (Miller, Farine, & Trisos, 2017). In this sense, we highlight that other mechanisms like negative biotic interaction may have contributed to our results. Two other subtle issues must be recognized here. First, it is possible that imperfect detectability could have influenced our results. For example, tadpoles that swim in head-up posture and form schools can be more easily sampled than solitary benthic species. Functional metrics seem to be robust to imperfect detection,

50

60

WILEY biotropica J Association for TROPICAL BIOLOGY

although detectability has the potential to bias trait-environment relationships (Roth, Allan, Pearman, & Amrhein, 2017). We were not able to correct functional composition without repeated surveys in the same pond. Yet, detection-adjusted occupancy models indicated a relationship between occupancy and agricultural activities and/or livestock management in the study area (Moreira et al., 2016). Second, the unavailability of information before conversion to productive lands is another limitation of this study. This issue is particularly hard to consider when working in the Brazilian Pampa, because untouched native grasslands and forests have been reduced by almost 90% (Cordeiro & Hasenack, 2009) and most sampling designs, such as the one used in our study, usually underestimate biodiversity losses (França et al., 2016).

In conclusion, this study showed that phylogenetic relatedness mediates most responses of anuran species to land use intensification in South American subtropical grasslands. Because tadpoles are often key consumers in freshwater wetlands, our results have direct effects on ecosystem structure. Under a scenario of land sharing (i.e., integrating biodiversity conservation and food production on the same land), our observations highlight that extensive cattle grazing over wide native pastures may provide an alternative more compatible with conservation than intensive short-term crops. However, reconciling biodiversity conservation with economic gains is a tough task, which minimally should involve strategies of land sharing and land sparing (Phalan, Onial, Balmford, & Green, 2011). We hope that our results contribute to the current debate about management practices that help maintaining biodiversity in productive areas.

ACKNOWLEDGMENTS

We thank D. Araújo for help with field sampling. L.F.B. Moreira is grateful to the *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior* (CAPES) for a postdoctoral fellowship (PNPD; process 20132816). H. Z. de Castilhos developed his work under a scholarship of *Iniciação Científica* funded by *Concelho Nacional de Desenvolvimento Científico e Tecnológico*-CNPq. This study is part of the Project "Influence of land use on tadpole diversity of grasslands at southern Brazil" and was supported by funds from CNPq, Brazil (grant number 442987/20). We thank P.I. Simões and three anonymous referees for suggestions on previous versions of the manuscript that increased its quality. We declare that the data collection complied with Brazilian current laws (Sistema de Autorização e Informação em Biodiversidade-SISBIO no. 41780).

CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTION

LFBM and SCF conceived the project and coordinated several aspects of the study. LFBM collected and conducted the specimen identification, analyzed the data, and participated in the drafting of the manuscript. HZC analyzed the data and drafted the manuscript. All authors read and approved the final version of the manuscript.

DATA AVAILABILITY STATEMENT

Data available from the Dryad Digital Repository: https:// doi.org/10.5061/dryad.tdz08kpx4 (Moreira, de Castilhos, & Castroviejo-Fisher, 2020).

ORCID

Leonardo Felipe Bairos Moreira D https://orcid. org/0000-0002-2753-9933

REFERENCES

- Altig, R., & McDiarmid, R. (1999). Body plan: Development and morphology. In R. McDiarmid, & R. Altig (Eds.), *Tadpoles: The biology of anuran larvae* (pp. 24–51). Chicago, IL: The University of Chicago Press.
- André, T. C., Cechin, S. Z., & Santos, T. G. (2019). Anurans in the Brazilian pampa biome: The first checklist from biological reserve of São Donato and a summary on actual knowledge of conservation units. *Herpetology Notes*, 12, 1095–1103.
- Azizi, E., Landberg, T., & Wassersug, R. J. (2007). Vertebral function during tadpole locomotion. *Zoology*, 110, 290–297. https://doi. org/10.1016/j.zool.2007.02.002
- Bernardi, R. E., Holmgren, M., Arim, M., & Scheffer, M. (2016). Why are forests so scarce in subtropical South America? The shaping roles of climate, fire and livestock. *Forest Ecology and Management*, 363, 212–217. https://doi.org/10.1016/j.foreco.2015.12.032
- Bivand, R., & Piras, G. (2015). Comparing implementations of estimation methods for spatial econometrics. *Journal of Statistical Software*, 63, 05–23. https://doi.org/10.18637/jss.v063.i18
- Blanchet, G., Legendre, P., & Borcard, D. (2008). Forward selection of spatial explanatory variables. *Ecology*, 89, 2623–2632. https://doi. org/10.1890/07-0986.1
- Bolzan, A. M. R., Hartmann, P. A., & Hartmann, M. T. (2014). Diversidade de anfíbios anuros de uma área de Pampa no município de São Gabriel, Rio Grande do Sul, Brasil. *Iheringia Série Zoologia*, 104, 277– 283. https://doi.org/10.1590/1678-476620141043277283
- Bolzan, A. M. R., Saccol, S. S. A., & Santos, T. G. (2016). Composition and diversity of anurans in the largest conservation unit in Pampa biome, Brazil. *Biota Neotropica*, 16, 1–14. https://doi. org/10.1590/1676-0611-BN-2015-0113
- Both, C., Cechin, S. Z., Melo, A. S., & Hartz, S. M. (2011). What controls tadpole richness and guild composition in ponds in subtropical grasslands? *Austral Ecology*, 36, 530–536. https://doi. org/10.1111/j.1442-9993.2010.02183.x
- Both, C., Melo, A. S., Cechin, S. Z., & Hartz, S. M. (2011). Tadpole co-occurrence in ponds: When do guilds and time matter? Acta Oecologica, 37(2), 140–145. https://doi.org/10.1016/j.actao.2011.01.008
- Britton, T., Oxelman, B., Vinnersten, A., & Bremer, K. (2002). Phylogenetic dating with confidence intervals using mean path lengths. *Molecular Phylogenetics and Evolution*, 24, 58–65. https://doi.org/10.1016/ S1055-7903(02)00268-3
- Cadotte, M. W., Carscadden, K., & Mirotchnick, N. (2011). Beyond species: Functional diversity and the maintenance of ecological processes and services. *Journal of Applied Ecology*, 48, 1079–1087. https://doi.org/10.1111/j.1365-2664.2011.02048.x
- Cadotte, M. W., Davies, T. J., & Peres-Neto, P. R. (2017). Why phylogenies do not always predict ecological differences. *Ecological Monographs*, 87, 535–551. https://doi.org/10.1002/ecm.1267
- Canavero, A., Brazeiro, A., Camargo, A., da Rosa, I., Maneyro, R., & Núñez, D. (2010). Amphibian diversity of Uruguay: Background knowledge, inventory completeness and sampling coverage. Boletín de la Sociedad Zoológica de Uruguay, 19, 1–19.
- Chao, A., Gotelli, N. J., Hsieh, T. C., Sander, E. L., Ma, K. H., Colwell, R. K., & Ellison, A. M. (2014). Rarefaction and extrapolation with Hill numbers: A framework for sampling and estimation in species

diversity studies. *Ecological Monographs*, 84, 45-67. https://doi. org/10.1890/13-0133.1

- Cordeiro, J. L. P., & Hasenack, H. (2009). Cobertura vegetal atual do Rio Grande do Sul. In V. P. Pillar, S. C. Müller, Z. M. S. Castilhos, & A. V. A. Jacques (Eds.), *Campos sulinos: Conservação e uso sustentável da biodiversidade* (pp. 285-299). Brasília: Ministério do Meio Ambiente.
- Cosentino, B. J., Schooley, R. L., & Phillips, C. A. (2011). Connectivity of agroecosystems: Dispersal costs can vary among crops. Landscape Ecology, 26, 371–379. https://doi.org/10.1007/s10980-010-9563-1
- Díaz-García, J. M., Pineda, E., López-Barrera, F., & Moreno, C. E. (2017). Amphibian species and functional diversity as indicators of restoration success in tropical montane forest. *Biodiversity and Conservation*, 26, 2569–2589. https://doi.org/10.1007/s10531-017-1372-2
- Diniz-filho, A. F., Cianciaruso, M. V., Rangel, T. F., & Bini, L. M. (2011). Eigenvector estimation of phylogenetic and functional diversity. *Functional Ecology*, 25, 735–744. https://doi. org/10.1111/j.1365-2435.2011.01836.x
- Dotta, G., Phalan, B., Silva, T. W., Green, R., & Balmford, A. (2016). Assessing strategies to reconcile agriculture and bird conservation in the temperate grasslands of South America. *Conservation Biology*, 30, 618–627. https://doi.org/10.1111/cobi.12635
- Duarte, L. S., Debastiani, V. J., Carlucci, M. B., & Diniz-Filho, J. A. F. (2018). Analyzing community-weighted trait means across environmental gradients: Should phylogeny stay or should it go? *Ecology*, 99, 385–398. https://doi.org/10.1002/ecy.2081
- Egea-Serrano, A., Relyea, R. A., Tejedo, M., & Torralva, M. (2012). Understanding of the impact of chemicals on amphibians: A meta-analytic review. *Ecology and Evolution*, 2, 1382–1397. https://doi. org/10.1002/ece3.249
- Egorov, E., Prati, D., Durka, W., Michalski, S., Fischer, M., Schmitt, B., ... Brändle, M. (2014). Does land-use intensification decrease plant phylogenetic diversity in local grasslands? *PLoS One*, *9*, 1–10. https://doi. org/10.1371/journal.pone.0103252
- Fleming, R. I., Mackenzie, C. D., Cooper, A., & Kennedy, M. W. (2009). Foam nest components of the túngara frog: A cocktail of proteins conferring physical and biological resilience. *Proceedings of the Royal Society B: Biological Sciences*, 276, 1787–1795. https://doi. org/10.1098/rspb.2008.1939
- Flynn, D. F. B., Gogol-Prokurat, M., Nogeire, T., Molinari, N., Richers, B. T., Lin, B. B., ... DeClerck, F. (2009). Loss of functional diversity under land use intensification across multiple taxa. *Ecology Letters*, 12, 22– 33. https://doi.org/10.1111/j.1461-0248.2008.01255.x
- França, F., Louzada, J., Korasaki, V., Griffiths, H., Silveira, J. M., & Barlow, J. (2016). Do space-for-time assessments underestimate the impacts of logging on tropical biodiversity? An Amazonian case study using dung beetles. *Journal of Applied Ecology*, 53, 1098–1105. https://doi. org/10.1111/1365-2664.12657
- Gerhold, P., Cahill, J. F., Winter, M., Bartish, I. V., & Prinzing, A. (2015). Phylogenetic patterns are not proxies of community assembly mechanisms (they are far better). *Functional Ecology*, 29, 600–614. https:// doi.org/10.1111/1365-2435.12425
- Gotelli, N. J. (2000). Null model analysis of species co-occurrence patterns. *Ecology*, *81*, 2606–2621. https://doi. org/10.1890/0012-9658(2000)081[2606:NMAOSC]2.0.CO;2
- Gotelli, N. J., & McCabe, D. J. (2002). Species co-occurrence: A meta-analysis of j. m. diamond's assembly rules model. *Ecology*, 83, 2091–2096. https://doi.org/10.1890/0012-9658(2002)083[2091:S-COAMA]2.0.CO;2
- Grosjean, S., Ohler, A., Chuaynkern, Y., Cruaud, C., & Hassanin, A. (2015). Improving biodiversity assessment of anuran amphibians using DNA barcoding of tadpoles. Case studies from Southeast Asia. *Comptes Rendus Biologies*, 338, 351–361. https://doi.org/10.1016/j. crvi.2015.03.015

- Guenard, G., Carsten von der Ohe, P., Carlisle Walker, S., Lek, S., & Legendre, P. (2014). Using phylogenetic information and chemical properties to predict species tolerances to pesticides. *Proceedings of the Royal Society B: Biological Sciences*, *281*, 20133239. https://doi. org/10.1098/rspb.2013.3239
- Heyer, R., Langone, J., La Marca, E., Azevedo-Ramos, C., diTada, I., Baldo, D., ... Hardy, J. (2010). *Leptodactylus latrans*. The IUCN Red List of Threatened Species 2010: e.T57151A11592655. Retrieved from https://www.iucnredlist.org/species/57151/11592655
- Hissa, D. C., Vasconcelos, I. M., Carvalho, A. F. U., Nogueira, V. L. R., Cascon, P., Antunes, A. S. L., ... Melo, V. M. M. (2008). Novel surfactant proteins are involved in the structure and stability of foam nests from the frog *Leptodactylus vastus*. *The Journal of Experimental Biology*, 211, 2707–2711. https://doi.org/10.1242/jeb.019315
- Howell, H. J., Mothes, C. C., Clements, S. L., Catania, S. V., Rothermel, B. B., & Searcy, C. A. (2019). Amphibian responses to livestock use of wetlands: New empirical data and a global review. *Ecological Applications*, 29, 1–15. https://doi.org/10.1002/eap.1976
- Hsieh, T. C., Ma, K. H., & Chao, A. (2016). iNEXT: An R package for rarefaction and extrapolation of species diversity (Hill numbers). *Methods in Ecology and Evolution*, 7, 1451–1456. https://doi. org/10.1111/2041-210X.12613
- Hua, J., Wuerthner, V. P., Jones, D. K., Mattes, B., Cothran, R. D., Relyea, R. A., & Hoverman, J. T. (2017). Evolved pesticide tolerance influences susceptibility to parasites in amphibians. *Evolutionary Applications*, 10, 802–812. https://doi.org/10.1111/eva.12500
- Isacch, J. P., Maceira, N. O., Bo, M. S., Demaría, M. R., & Peluc, S. (2005). Bird-habitat relationship in semi-arid natural grasslands and exotic pastures in the west pampas of Argentina. *Journal of Arid Environments*, 62, 267–283. https://doi.org/10.1016/j.jaridenv.2004.11.008
- Jansen, A., & Healey, M. (2003). Frog communities and wetland condition: Relationships with grazing by domestic livestock along an Australian floodplain river. *Biological Conservation*, 109, 207–219. https://doi.org/10.1016/S0006-3207(02)00148-9
- Kehoe, L., Kuemmerle, T., Meyer, C., Levers, C., Václavík, T., & Kreft, H. (2015). Global patterns of agricultural land-use intensity and vertebrate diversity. *Diversity and Distributions*, 21, 1308–1318. https:// doi.org/10.1111/ddi.12359
- Kembel, S. W., Cowan, P. D., Helmus, M. R., Cornwell, W. K., Morlon, H., Ackerly, D. D., ... Webb, C. O. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26, 1463–1464. https://doi. org/10.1093/bioinformatics/btq166
- Laliberté, E., Legendre, P., & Shipley, B. (2014). FD: Measuring functional diversity from multiple traits, and other tools for functional ecology. R package version 1.0-12. Retrieved from https://cran.r-project.org/ package=FD
- Lee, M.-B., & Martin, J. A. (2017). Avian species and functional diversity in agricultural landscapes: Does landscape heterogeneity matter? *PLoS* One, 12, e0170540. https://doi.org/10.1371/journal.pone.0170540
- Lipinski, V. M., & Santos, T. G. (2014). Estrutura e organização espacial de duas comunidades de anuros do bioma pampa. *Iheringia Serie Zoologia*, 104, 462–469. https://doi.org/10.1590/1678-4766201410 44462469
- Machado, I. F., & Maltchik, L. (2010). Can management practices in rice fields contribute to amphibian conservation in southern Brazilian wetlands? Aquatic Conservation: Marine and Freshwater Ecosystems, 20, 39–46. https://doi.org/10.1002/aqc.1070
- Maneyro, R. (2008). Check list of anurans (Amphibia, Anura) from "Campo del Abasto" and surroundings, Rivera department, Uruguay. *Boletin de la Sociedad Zoologica Del Uruguay*, 17, 34–41.
- Maneyro, R., & Carreira, S. (2012). *Guía de anfíbios del Urugay*. Montevideo, Uruguay: Ediciones de la Fuga.
- Mann, R. M., Hyne, R. V., Choung, C. B., & Wilson, S. P. (2009). Amphibians and agricultural chemicals: Review of the risks in a

complex environment. *Environmental Pollution*, 157(11), 2903–2927. https://doi.org/10.1016/j.envpol.2009.05.015

- Medina, R. G., Ponssa, M. L., & Aráoz, E. (2016). Environmental, land cover and land use constraints on the distributional patterns of anurans: *Leptodacylus* species (Anura, Leptodactylidae) from Dry Chaco. *PeerJ*, 4, e2605. https://doi.org/10.7717/peerj.2605
- Melo, L. S. O., Garey, M. V., & Rossa-Feres, D. C. (2018). Looking for a place: How are tadpoles distributed within tropical ponds and streams ? *Herpetology Notes*, 11, 379–386.
- Miller, E. T., Farine, D. R., & Trisos, C. H. (2017). Phylogenetic community structure metrics and null models: A review with new methods and software. *Ecography*, 40, 461–477. https://doi.org/10.1111/ ecog.02070
- Modernel, P., Rossing, W. A. H., Corbeels, M., Dogliotti, S., Picasso, V., & Tittonell, P. (2016). Land use change and ecosystem service provision in Pampas and Campos grasslands of southern South America. *Environmental Research Letters*, 11, 113002. https://doi. org/10.1088/1748-9326/11/11/113002
- Moreira, L. F. B., de Castilhos, H. Z., & Castroviejo-Fisher, S. (2020). Data from: Something is not quite right: Effects of two land uses on anuran diversity in subtropical grasslands. *Dryad Digital Repository*, https:// doi.org/10.5061/dryad.tdz08kpx4
- Moreira, L. F. B., & Maltchik, L. (2012). Assessing patterns of nestedness and co-occurrence in coastal pond anuran assemblages. *Amphibia-Reptilia*, 33, 261–271. https://doi.org/10.1163/15685 3812X641721
- Moreira, L. F. B., & Maltchik, L. (2015). Our time will come: Is anuran community structure related to crop age? *Austral Ecology*, 40, 827–835. https://doi.org/10.1111/aec.12260
- Moreira, L. F. B., Moura, R. G., & Maltchik, L. (2016). Stop and ask for directions: Factors affecting anuran detection and occupancy in Pampa farmland ponds. *Ecological Research*, 31, 65–74. https://doi. org/10.1007/s11284-015-1316-9
- Moreira, L. F. B., Solino-Carvalho, L. A., Strüssmann, C., & Silveira, R. M. L. (2016). Effects of exotic pastures on tadpole assemblages in Pantanal floodplains: Assessing changes in species composition. *Amphibia Reptilia*, 37, 179–190. https://doi.org/10.1163/15685381-00003043
- Mouillot, D., Graham, N. A. J., Villéger, S., Mason, N. W. H., & Bellwood, D. R. (2013). A functional approach reveals community responses to disturbances. *Trends in Ecology and Evolution*, 28, 167–177. https:// doi.org/10.1016/j.tree.2012.10.004
- Nowakowski, A. J., Frishkoff, L. O., Thompson, M. E., Smith, T. M., & Todd, B. D. (2018). Phylogenetic homogenization of amphibian assemblages in human-altered habitats across the globe. Proceedings of the National Academy of Sciences of the United States of America, 115, E3454–E3462. https://doi.org/10.1073/PNAS.1714891115
- Oliveira, T. E. D., Freitas, D. S. D., Gianezini, M., Ruviaro, C. F., Zago, D., Mércio, T. Z., ... Barcellos, J. O. J. (2017). Agricultural land use change in the Brazilian Pampa Biome: The reduction of natural grasslands. *Land Use Policy*, *63*, 394–400. https://doi.org/10.1016/j.landu sepol.2017.02.010
- Overbeck, G., Muller, S., Fidelis, A., Pfadenhauer, J., Pillar, V., Blanco, C., ... Forneck, E. (2007). Brazil's neglected biome: The south Brazilian campos. Perspectives in Plant Ecology, Evolution and Systematics, 9, 101–116. https://doi.org/10.1016/j.ppees.2007.07.005
- Peltzer, P. M., Lajmanovich, R. C., Attademo, A. M., Junges, C. M., Cabagna-Zenklusen, M. C., Repetti, M. R., ... Beldoménico, H. (2013). Effect of exposure to contaminated pond sediments on survival, development, and enzyme and blood biomarkers in veined treefrog (*Trachycephalus typhonius*) tadpoles. *Ecotoxicology and Environmental Safety*, 98, 142–151. https://doi.org/10.1016/j.ecoenv.2013.09.010
- Pereira, E. B., Pinto-ledezma, J. N., Freitas, C. G. D. E., Villalobos, F., Collevatti, R. G., & Maciel, N. M. (2017). Evolution of the anuran foam nest : Trait conservatism and lineage diversification. *Biological Journal* of the Linnean Society, 122, 814–823.

- Phalan, B., Onial, M., Balmford, A., & Green, R. E. (2011). Reconciling food production and biodiversity conservation: Land sharing and land sparing compared. *Science*, 333, 1289–1291. https://doi. org/10.1126/science.1208742
- Phillips, H. R. P., Newbold, T., & Purvis, A. (2017). Land-use effects on local biodiversity in tropical forests vary between continents. *Biodiversity* and Conservation, 26, 2251–2270. https://doi.org/10.1007/s1053 1-017-1356-2
- Prado, V. H. M., & Rossa-Feres, D. C. (2014). Multiple determinants of anuran richness and occurrence in an agricultural region in Southeastern Brazil. *Environmental Management*, 53, 823–837. https://doi. org/10.1007/s00267-014-0241-y
- Project MapBiomas (2019). Collection 3.0 of Brazilian land cover & use map series. Retrieved from https://mapbiomas.org
- Pyron, R. A. (2014). Biogeographic analysis reveals ancient continental vicariance and recent oceanic dispersal in amphibians. Systematic Biology, 63, 779–797. https://doi.org/10.1093/sysbio/syu042
- QGIS Development Team (2018). QGIS geographic information system.
- Queiroz, C. S., da Silva, F. R., & Rossa-Feres, D. C. (2015). The relationship between pond habitat depth and functional tadpole diversity in an agricultural landscape. *Royal Society Open Science*, *2*, 150165. https:// doi.org/10.1098/rsos.150165
- Revell, L. J. (2012). phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3, 217–223. https://doi.org/10.1111/j.2041-210X.2011.00169.x
- Ribeiro, J., Colli, G. R., Caldwell, J. P., Ferreira, E., Batista, R., & Soares, A. (2017). Evidence of neotropical anuran community disruption on rice crops: A multidimensional evaluation. *Biodiversity and Conservation*, 26, 3363–3383. https://doi.org/10.1007/s10531-017-1410-0
- Riemann, J. C., Ndriantsoa, S. H., Rödel, M. O., & Glos, J. (2017). Functional diversity in a fragmented landscape – Habitat alterations affect functional trait composition of frog assemblages in Madagascar. *Global Ecology and Conservation*, 10, 173–183. https://doi.org/10.1016/j. gecco.2017.03.005
- Roth, T., Allan, E., Pearman, P. B., & Amrhein, V. (2017). Functional ecology and imperfect detection of species. *Methods in Ecology and Evolution*, 9, 917–928. https://doi.org/10.1111/2041-210X.12950
- Saccol, S. S. A., Bolzan, A. M. R., & Santos, T. G. (2017). In the shadow of trees: Does *Eucalyptus* afforestation reduce herpetofaunal diversity in southern Brazil? *South American Journal of Herpetology*, 12, 42–56. https://doi.org/10.2994/SAJH-D-16-00028.1
- Santos, T. G., Kopp, K., Spies, M. R., Trevisan, R., & Cechin, S. Z. (2008). Distribuição temporal e espacial de anuros em área de Pampa, Santa Maria, RS. *Iheringia Série Zoologia*, 4, 244–253. https://doi. org/10.1590/S0073-47212008000200013
- Schieltz, J. M., & Rubenstein, D. I. (2016). Evidence based review: Positive versus negative effects of livestock grazing on wildlife. What do we really know? Environmental Research Letters, 11, 113003. https://doi. org/10.1088/1748-9326/11/11/113003
- Schiesari, L., & Corrêa, D. T. (2016). Consequences of agroindustrial sugarcane production to freshwater biodiversity. *Global Change Biology Bioenergy*, 8, 644–657. https://doi.org/10.1111/gcbb.12279
- Schmera, D., Heino, J., Podani, J., Erös, T., & Dolédec, S. (2017). Functional diversity: A review of methodology and current knowledge in freshwater macroinvertebrate research. *Hydrobiologia*, 787, 27-44. https://doi.org/10.1007/s10750-016-2974-5
- Schmutzer, A. C., Gray, M. J., Burton, E. C., & Miller, D. L. (2008). Impacts of cattle on amphibian larvae and the aquatic environment. *Freshwater Biology*, 53, 2613–2625. https://doi. org/10.1111/j.1365-2427.2008.02072.x
- Staude, I. R., Vélez-Martin, E., Andrade, B. O., Podgaiski, L. R., Boldrini, I. I., Mendonça, M., ... Overbeck, G. E. (2018). Local biodiversity erosion in South Brazilian grasslands under moderate levels of landscape habitat loss. *Journal of Applied Ecology*, 55, 1241–1251. https://doi. org/10.1111/1365-2664.13067

- Strauss, A., Reeve, E., Randrianiaina, R.-D., Vences, M., & Glos, J. (2010). The world's richest tadpole communities show functional redundancy and low functional diversity: Ecological data on Madagascar's stream-dwelling amphibian larvae. BMC Ecology, 10, 12. https://doi. org/10.1186/1472-6785-10-12
- Stroud, J. T., & Losos, J. B. (2016). Ecological opportunity and adaptive radiation. Annual Review of Ecology, Evolution, and Systematics, 47, 507–532. https://doi.org/10.1146/annurev-ecolsys-121415-032254
- Trimble, M. J., & van Aarde, R. J. (2014). Amphibian and reptile communities and functional groups over a land-use gradient in a coastal tropical forest landscape of high richness and endemicity. *Animal Conservation*, 17, 441–453. https://doi.org/10.1111/acv.12111
- Tscharntke, T., Klein, A. M., Kruess, A., Steffan-Dewenter, I., & Thies, C. (2005). Landscape perspectives on agricultural intensification and biodiversity – Ecosystem service management. *Ecology Letters*, 8, 857–874. https://doi.org/10.1111/j.1461-0248.2005.00782.x
- Tsianou, M. A., & Kallimanis, A. S. (2016). Different species traits produce diverse spatial functional diversity patterns of amphibians. *Biodiversity and Conservation*, 25, 117–132. https://doi.org/10.1007/ s10531-015-1038-x
- Tsianou, M. A., & Kallimanis, A. S. (2019). Trait selection matters! A study on European amphibian functional diversity patterns. *Ecological Research*, 34, 225–234. https://doi.org/10.1111/1440-1703.1002
- Veach, V., Moilanen, A., & Di Minin, E. (2017). Threats from urban expansion, agricultural transformation and forest loss on global conservation priority areas. *PLoS One*, 12, 1–14. https://doi.org/10.1371/journ al.pone.0188397
- Venesky, M. D., Wassersug, R. J., & Parris, M. J. (2010). How does a change in labial tooth row number affect feeding kinematics and

foraging performance of a ranid tadpole (*Lithobates sphenocephalus*)? *Biological Bulletin*, 218, 160–168. https://doi.org/10.1645/GE-2353.1

- Villéger, S., Mason, H., & Mouillot, D. (2008). New multidimensional functional diversity indices for a multifaceted framework in fucntional ecology. *Ecology*, 89, 2290–2301.
- Villéger, S., Miranda, J. R., Hernández, D. F., & Mouillot, D. (2010). Contrasting changes in taxonomic vs. functional diversity of tropical fish communities after habitat degradation. *Ecological Applications*, 20, 1512–1522. https://doi.org/10.1890/09-1310.1
- Zina, J. (2006). Communal nests in *Physalaemus pustulosus* (Amphibia: Leptodactylidae): Experimental evidence for female oviposition preferences and protection against desiccation. *Amphibia-Reptilia*, 27, 148–150. https://doi.org/10.1163/156853806776052092

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Moreira LFB, de Castilhos HZ, Castroviejo-Fisher S. Something is not quite right: Effects of two land uses on anuran diversity in subtropical grasslands. *Biotropica*. 2020;52:1286–1297. <u>https://doi.org/10.1111/</u> btp.12836