

Modulatory potential of resveratrol during lung inflammatory disease



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ABSTRACT

Neutrophils are the first cells to achieve the sites of infection or inflammation in the lungs. The massive accumulation of these cells is associated with acute and chronic lung injury. Therefore, they have been implicated in the pathogenesis of many lung diseases through the release of reactive oxygen intermediates, proteolytic enzymes and Neutrophil Extracellular Traps (NETs). The excessive and continuous release of NETs, fibers composed by decondensed chromatin coated with neutrophil proteins, are associated to the impairment of lung function in different pathological settings.

Flavonoids inhibit the respiratory burst of neutrophils in mammals. However, one of these flavonoids, resveratrol has a particular chemical property. It reduce Cu(II) to Cu(I) form with concomitant formation of reactive oxygen species, which can produce DNA breakage as reported in several in vitro models.

We hypothesize that direct resveratrol administration in lungs can cleave DNA in NETs, improving lung function during acute airway infections or chronic inflammatory lung diseases.

If the hypothesis is correct, the control of NET formation can be used to reduce the inflammatory environment in lung after neutrophil stimuli. Additionally, the production of proinflammatory cytokines by neutrophils could be also diminished by resveratrol administration. In this sense, this flavonoid provides a multifaceted opportunity for treatment of lung diseases with strong or chronic neutrophil activation.

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Introduction

Neutrophils constitute the first line of defense against infections by eliminating phagocytosed pathogenic bacteria, fungi and viruses [1–5]. Generally, neutrophils are present in pulmonary capillaries in higher numbers compared to systemic blood even in the absence of inflammatory stimuli [6]. This phenomenon allows neutrophils to migrate into the lungs as a response to an inflammatory insult. During inflammation, neutrophils are activated upon stimulation and migrate into the lungs [7].

Neutrophils recruited to the lungs engulf microbes into a phagocytic vacuole that fuse with intracellular granules [8]. These granules contain an overlapping set of antimicrobial proteins, such as gelatinase, lipocalin, and lysozyme among other proteins [9]. More recently, neutrophils have been shown to possess an alternative mechanism of pathogen killing, designated as Neutrophil Extracellular Traps (NETs). NETs are extracellular structures composed by decondensed chromatin complexed with granule and cytoplasmic proteins [10–12]. These traps bind and kill pathogens

by juxtaposing microbes with neutrophil antimicrobial proteins and histones [10,11]. NET formation is accompanied by neutrophil death (NETosis), a non-conventional form of cell death [11–13]. Besides being expressed on NETs, neutrophil elastase and myeloperoxidase also regulate NET formation [14]. Furthermore, histone deimination by peptidylarginine deiminase 4 (PAD4) is a central step to NET formation [15]. Additionally, reactive oxygen species produced by the assembly and activation of NADPH oxidase are also required for NET release [13,16].

Although NETs are released to kill microorganisms and prevent microbial spreading, the excessive NET formation leads to tissue injury. Massive NET production has been demonstrated to be involved in the pathogenesis of several diseases, such as acute respiratory distress syndrome (ARDS), viral infections, cystic fibrosis, asthma and chronic obstructive pulmonary disease (COPD) [6,17–19]. In an inflamed tissue, prolonged or excessive release of cytotoxic molecules anchored on NETs, accompanied by delayed apoptosis of neutrophils, amplify inflammation, increasing tissue damage [20] and reduction of lung function [21–23], as is the case in severe influenza pneumonia, where NETs contribute to lung tissue injury [24].

Current development of therapies to target NETs in inflammatory lung diseases include treatment with recombinant human

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DNase (dornase alfa/pulmozyme) [25,26], anti-histone antibodies [27,28] and antiproteases [29]. However, these therapies require the use of recombinant DNA technologies with high-quality purification. Therefore, the need for alternative approaches to dismantle NETs structure is clear.

Flavonoids, a group of plant secondary metabolites, affect inflammation and particularly neutrophil activity [30]. This group have been attributed to the capacity of plant phenols to prevent activation of nuclear factor-kappa B (NF- κ B) and the subsequent overexpression of pro-inflammatory mediators (cytokines, adhesion molecules, cyclooxygenase-2, 5-lipoxygenase, myeloperoxidase or inducible nitric oxide synthase), and to inhibit neutrophil apoptosis [20]. However, a flavonoid member, resveratrol has an additional property; it is capable of inducing oxidative damage in DNA in the presence of certain transition metal ions [31–33]. This characteristic can be used to develop chemical modulators of NETs.

Hypothesis

We hypothesize that resveratrol (Rsv) could act as a DNA cleavage agent of NETs, produced by inflammatory lung diseases and microbial infections improving lung function. Our hypothesis is represented in Fig. 1.

For an easier analysis of our hypothesis, we support in two interrelated points based on Rsv properties: DNA-damaging activity and bioavailability in the lungs.

DNA-damaging activity of resveratrol

Rsv (3,4',5-trihydroxy-*trans*-stilbene) is a phytoalexin polyphenolic found in grape skins, peanuts, and red wine, has been reported to have a wide range of biological and pharmacological properties [34–36]. These chemical properties facilitate their permeability across cellular membranes to interact with multiple protein targets [37], binding to DNA and chelating metal ions. These properties combined with their pro-oxidant activity convert this polyphenol in a multifaceted component with great therapeutic potential. Rsv forms a complex with intracellular copper metal [Cu(II)], leading to its reduction [Cu(I)] with in situ formation of reactive oxygen species (ROS), which induce DNA breakage. Several experiments showed DNA damage mediated by the combined reaction Rsv \pm Cu(II) in different DNA substrates (bacteriophage DNA, plasmids, calf thymus DNA and intracellular DNA of human peripheral lymphocytes) [31–33].

Fukuhara and Miyata proposed that Cu(II)-dependent DNA damage by Rsv is caused predominantly by a copper \pm peroxide complex with diffusible oxygen species production [38]. These authors have further suggested that the reaction occurs without oxygenated transformation of benzene nuclei. However, Ahmad et al. [31] indicate that “transformed species” of Rsv, possibly in an oxidized form, are produced in the presence of Cu(II). It is possible that more than one mechanism of DNA cleavage is involved, but in consensus Cu ions are essential to oxidative reaction to account. Other metal ions such as [Co(II), Fe(II), Mn(II), Mg(II) and Ni(II)] do not produce DNA cleavage reaction as they do not interact with Rsv [31]. A summary of putative prooxidant reaction

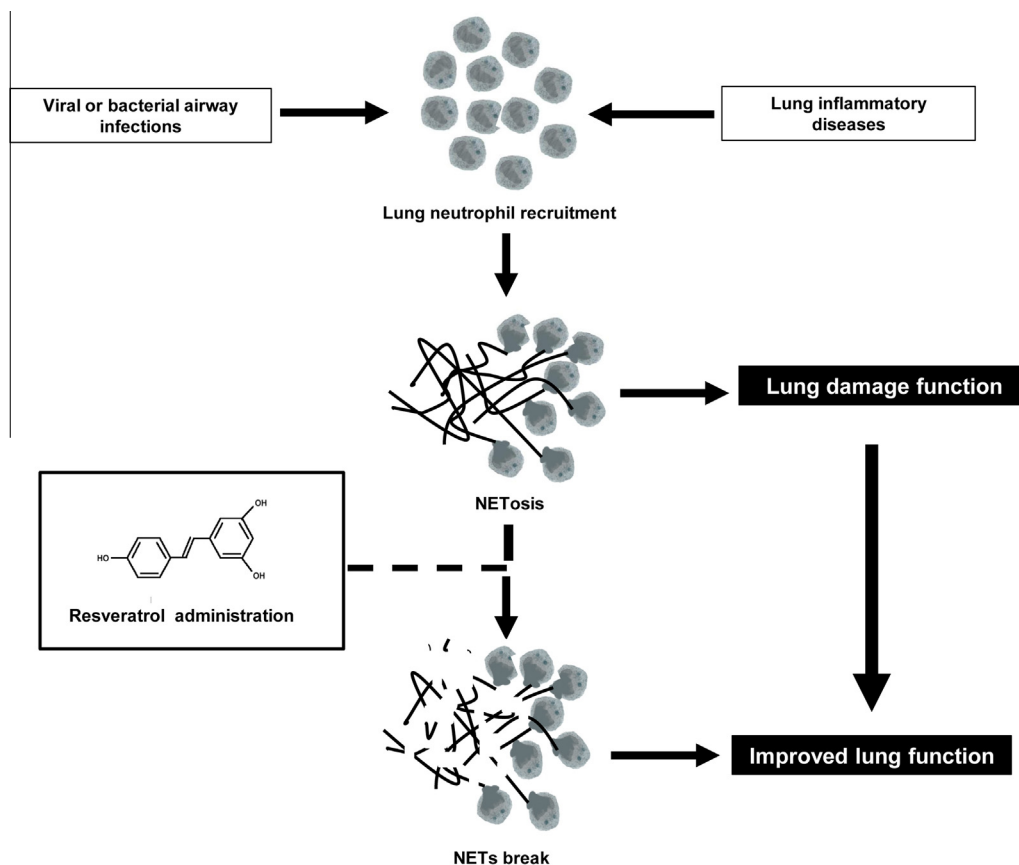


Fig. 1. Schematic representation of hypothesis for resveratrol action on NETs. Viral or bacterial airway infections or inflammatory diseases induce neutrophil recruitment and activation in the lungs. These stimuli can promote NETosis, which leads to lung tissue damage and impair lung function. Resveratrol administration can cleave NETs and induce the improvement of lung function.

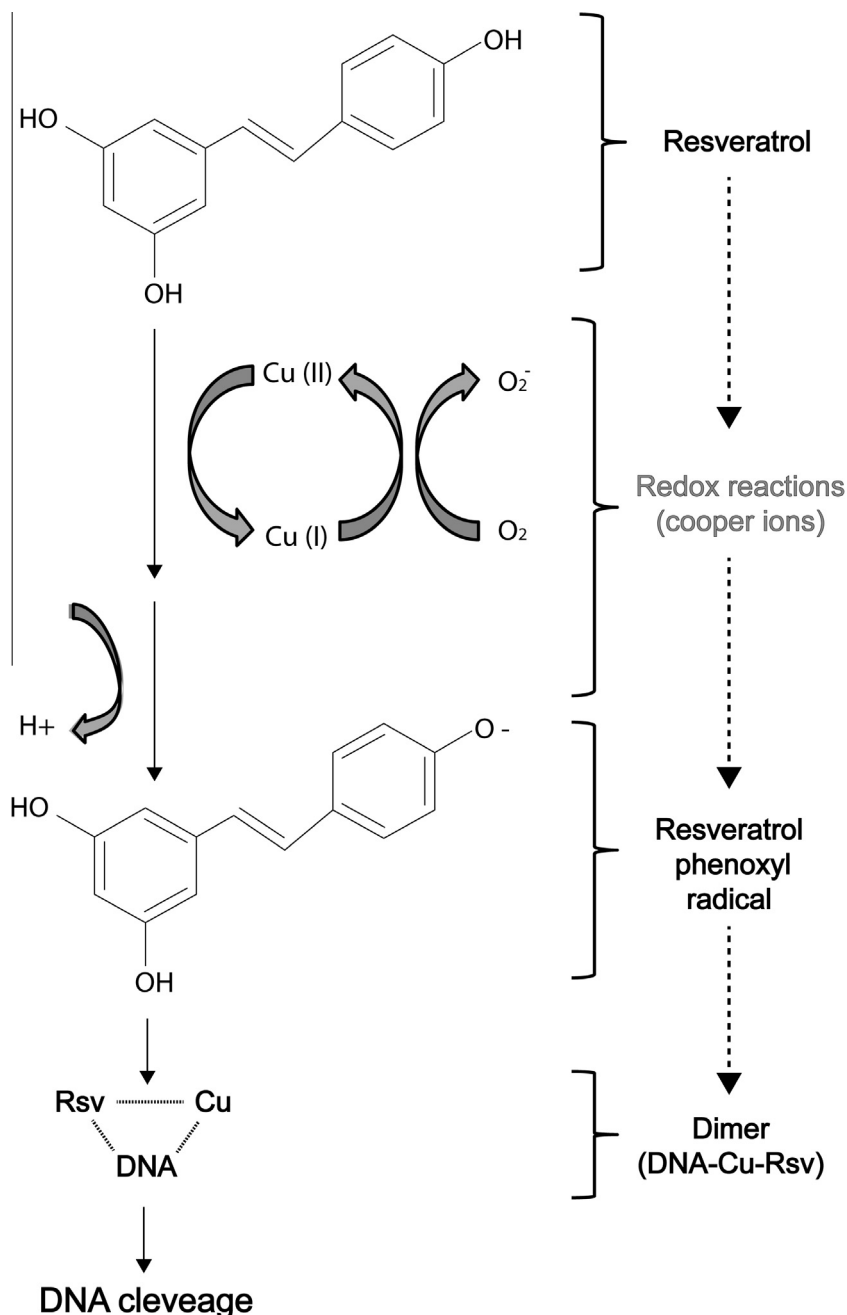


Fig. 2. Putative mechanism showing mobilization of endogenous copper ions mediated by resveratrol. This figure was adapted from Villegas et al. [63].

mediated by Rsv in presence of copper ions is represented in the Fig. 2.

Copper metal bioavailability becomes a limiting factor for Rsv ± Cu(II) reaction to induce DNA breakage. The real quantification of cellular copper concentration is imprecise. It is due to the non-existence of free copper in the nature [39,40]. Extracellular and intracellular copper is bound to cellular components (i.e., copper chaperones) [40,41]. Independent of these difficulties, cellular copper concentration could be sufficient to complex to Rsv to catalyze DNA damage reaction, which could support our hypothesis.

Copper ions are closely associated with bases of DNA being a normal component of chromatin. Different levels of copper promote structural and functional modifications of DNA and induce chromatin condensation in a concentration-dependent manner

[42]. For this reason, copper is one of the etiological factors most described in neurodegenerative diseases (Alzheimer's disease, Parkinson, Huntington and familial amyotrophic lateral sclerosis), where an abnormal metal accumulation is associated directly to the pathological state [43].

In situ endogenous copper ions are mobilized easily from DNA by chelating agents and the consequent induction of pro-oxidant reactions [31,44]. Gutteridge showed that copper is loosely bound copper to DNA, which is available for binding to the chelating agent, as 1,10-phenanthroline that cause internucleosomal DNA fragmentation [31,45]. Rsv would react in a similar manner that 1,10-phenanthroline with the intracellular copper [33]. It is possible that such loosely bound copper of NETs can also be mobilized by Rsv and cleave reaction can account. So far, to our knowledge, any study has shown metal ions associations to NETs. However,

it is possible that NETosis process could maintain copper ions associated to chromatin after neutrophil death in inflammatory environments.

In the lungs, an alternative mechanism should compensate the possible low ion concentration of copper in NETs. For example, a redox cycle of this metal ion should result in increased rate of DNA hydrolysis in situ on NETs in the presence of Rsv. Ahmed showed that Cu(I) recycled in the reaction [31,46]. This study suggests that “oxidized species” of Rsv are also able to catalyze the reduction of recycled copper ions [Cu(II)]. It facilitates to obtain copper concentration necessary to DNA break to takes place.

Other alternative is the administration of Rsv together with Cu (II) ions to the lungs, but it should be performed with precautions, since high levels of copper can be toxic to the respiratory tract [47].

Resveratrol bioavailability in the lungs

The bioavailability of a chemical compound is defined by the capacity to become available to the target tissue after its administration [48]. Current knowledge on metabolism and bioavailability of Rsv has been described in humans, dependent on route of administration. The oral bioavailability of Rsv is very low due to its rapid and extensive metabolism, elimination through ABC transporters and incomplete intestinal absorption [49–51]. It is important to note that lungs (along with gut and liver) are third in a series of three putative metabolism sites, which orally ingested Rsv must cross before entering the circulation. First step metabolism by gut, liver and lungs can synergistically promote total body clearance of Rsv [52]. Considering our hypothesis, oral administration of this polyphenol is a problem for therapeutic use. In addition, metabolism of Rsv would produce unexpected difficulties. The metabolism of this polyphenol involves complex pathways that produce alterations on its structure through glucuronides and sulfates conjugations [53]. In in vitro experiments, Rsv induces DNA damage in a non-metabolized form (above-discussed in previous section). Chemical modifications on Rsv could be important determinants of its pharmacological activity [54]. However, other study suggest that resveratrol's glucuronide or sulfate conjugates may be unconjugated at the target sites of action, thereby releasing to elicit his biological activity [55]. In addition, was described that resveratrol's glucuronide has comparable or same degree of activity that non-metabolized Rsv depending on the test model [56]. In this sense, more studies will be required to understand its effects on the DNA damage.

Aerosol version of Rsv for inhalation probably would be a better option. Ulterior work showed that atomization inhaled Rsv could alleviate rat COPD lung injury inducing an amelioration of pathological changes [57]. Considering our hypotheses, this route of administration offer two major advantages. Firstly, it is a non-invasive manner to deliver drugs or compounds rapidly and directly into the airways. In a second point, aerosol Rsv could increase directly the polyphenol-DNA interactions (without metabolized intermediate, simulating in vitro experiments). In our case, Rsv would interact with extracellular DNA derived from NETosis process. In addition, several approaches to improve inhaled versions of Rsv using vehicles were developed, ie Rsv-loaded gelatin nanoparticles used to treat lung cancer [58] or inhalable microparticles containing budesonide and Rsv to reduce alveolar inflammation [59]. However, the delivery of Rsv directly to the lungs continues to be a challenge.

It is noteworthy that we could not discard the use of other routes of Rsv administration. Rsv administered by an intraperitoneal (IP) route, inhibits airway inflammation, and airway remodeling which are the main contributors to airway hyper reactivity

and irreversible lung function loss in rats [60]. However, this route of Rsv delivery has similar availability limitations of oral administration in humans. Additionally, abdominal pain is a common and undesired consequence of the IP route of drug delivery, as described for chemotherapy treatments [61].

We focused our hypothesis to improve lung function using Rsv during Netosis promoted during inflammatory lung diseases. However, other stilbenoid, piceatannol, was described with similar properties (DNA breakage in presence of copper ions) [62]. Given that the biological properties of Rsv has been more studied, we decided to explore Rsv for this hypothesis, but we do not discard the potential of piceatannol to improve lung function.

Conclusion

Resveratrol could act as a DNA cleavage agent of NETs, produced by inflammatory lung diseases or viral and microbial infections. In this sense, this study could help researchers to develop chemical modulators based on resveratrol to improve lung function.

Conflict of interest statement

The authors declare that no competing interests exist.

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References

- [1] Rosen H. Bacterial responses to neutrophil phagocytosis. *Curr Opin Hematol* 2004;11:1–6.
- [2] Rubin-Bejerano I, Abejón C, Magnelli P, Crisafi P, Fink GR. Phagocytosis by human neutrophils is stimulated by a unique fungal cell wall component. *Cell Host Microbe* 2007;2:55–67.
- [3] Hashimoto Y, Moki T, Takizawa T, Shiratsuchi A, Nakanishi Y. Evidence for phagocytosis of influenza virus-infected, apoptotic cells by neutrophils and macrophages in mice. *J Immunol* 2007;178:2448–57.
- [4] Craig A, Mai J, Cai S, Jeyaseelan S. Neutrophil recruitment to the lungs during bacterial pneumonia. *Infect Immun* 2009;77:568–75.
- [5] Gazendam RP, van de Geer A, Roos D, van den Berg TK, Kuijpers TW. How neutrophils kill fungi. *Immunol Rev* 2016;273:299–311.
- [6] Cheng OZ, Palaniyar N. NET balancing: a problem in inflammatory lung diseases. *Front Immunol* 2013;4:1.
- [7] Ley K, Laudanna C, Cybulsky MI, Nourshargh S. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat Rev Immunol* 2007;7:678–89.
- [8] Balamayoran G, Batra S, Fessler MB, Happel KI, Jeyaseelan S. Mechanisms of neutrophil accumulation in the lungs against bacteria. *Am J Respir Cell Mol Biol* 2010;43:5–16.
- [9] Urban CF, Lourido S, Zychlinsky A. How do microbes evade neutrophil killing? *Cell Microbiol* 2006;8:1687–96.
- [10] Young RL, Malcolm KC, Kret JE, et al. Neutrophil extracellular trap (NET)-mediated killing of *Pseudomonas aeruginosa*: evidence of acquired resistance within the CF airway, independent of CFTR. *PLoS One* 2011;6:e23637.
- [11] Mesa MA, Vasquez G. NETosis. *Autoimmune Dis* 2013;2013:651497.
- [12] Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science* 2004;303:1532–5.
- [13] Brinkmann V, Zychlinsky A. Beneficial suicide: why neutrophils die to make NETs. *Nat Rev Microbiol* 2007;5:577–82.
- [14] Papayannopoulos V, Metzler KD, Hakkim A, Zychlinsky A. Neutrophil elastase and myeloperoxidase regulate the formation of neutrophil extracellular traps. *J Cell Biol* 2010;191:677–91.
- [15] Li P, Li M, Lindberg MR, et al. PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. *J Exp Med* 2010;207:1853–62.
- [16] Munoz-Caro T, Lendner M, Dausgschies A, Hermsilla C, Taubert A. NADPH oxidase, MPO, NE, ERK1/2, p38 MAPK and Ca²⁺ influx are essential for *Cryptosporidium parvum*-induced NET formation. *Dev Comp Immunol* 2015;52:245–54.
- [17] Cortjens B, de Boer OJ, de Jong R, et al. Neutrophil extracellular traps cause airway obstruction during respiratory syncytial virus disease. *J Pathol* 2016;238:401–11.

- [18] Wright TK, Gibson PG, Simpson JL, McDonald VM, Wood LG, Baines KJ. Neutrophil extracellular traps are associated with inflammation in chronic airway disease. *Respirology* 2016;21:467–75.
- [19] Grabcanovic-Musija F, Obermayer A, Stoiber W, et al. Neutrophil extracellular trap (NET) formation characterises stable and exacerbated COPD and correlates with airflow limitation. *Respir Res* 2015;16:59.
- [20] Jancinova V, Perecko T, Harmatha J, Nosal R, Drabikova K. Decreased activity and accelerated apoptosis of neutrophils in the presence of natural polyphenols. *Interdiscip Toxicol* 2012;5:59–64.
- [21] Sagel SD, Wagner BD, Anthony MM, Emmett P, Zemanick ET. Sputum biomarkers of inflammation and lung function decline in children with cystic fibrosis. *Am J Respir Crit Care Med* 2012;186:857–65.
- [22] Tate MD, Deng YM, Jones JE, Anderson GP, Brooks AG, Reading PC. Neutrophils ameliorate lung injury and the development of severe disease during influenza infection. *J Immunol* 2009;183:7441–50.
- [23] Redding GJ. Current concepts in adult respiratory distress syndrome in children. *Curr Opin Pediatr* 2001;13:261–6.
- [24] Narayana Moorthy A, Narasaraju T, Rai P, et al. In vivo and in vitro studies on the roles of neutrophil extracellular traps during secondary pneumococcal pneumonia after primary pulmonary influenza infection. *Front Immunol* 2013;4:56.
- [25] Shak S, Capon DJ, Hellmiss R, Marsters SA, Baker CL. Recombinant human DNase I reduces the viscosity of cystic fibrosis sputum. *Proc Natl Acad Sci USA* 1990;87:9188–92.
- [26] Hakkim A, Furnrohr BG, Amann K, et al. Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc Natl Acad Sci USA* 2010;107:9813–8.
- [27] Xu J, Zhang X, Pelayo R, et al. Extracellular histones are major mediators of death in sepsis. *Nat Med* 2009;15:1318–21.
- [28] Semeraro F, Ammolto CT, Morrissey JH, et al. Extracellular histones promote thrombin generation through platelet-dependent mechanisms: involvement of platelet TLR2 and TLR4. *Blood* 2011;118:1952–61.
- [29] Greene CM, McElvaney NG. Proteases and antiproteases in chronic neutrophilic lung disease – relevance to drug discovery. *Br J Pharmacol* 2009;158:1048–58.
- [30] Ciz M, Denev P, Kratchanova M, Vasicek O, Ambrozova G, Lojek A. Flavonoids inhibit the respiratory burst of neutrophils in mammals. *Oxid Med Cell Longev* 2012;2012:181295.
- [31] Ahmad A, Farhan Asad S, Singh S, Hadi SM. DNA breakage by resveratrol and Cu(II): reaction mechanism and bacteriophage inactivation. *Cancer Lett* 2000;154:29–37.
- [32] Zheng LF, Wei QY, Cai YJ, et al. DNA damage induced by resveratrol and its synthetic analogues in the presence of Cu(II) ions: mechanism and structure-activity relationship. *Free Radical Biol Med* 2006;41:1807–16.
- [33] Hadi SM, Ullah MF, Azmi AS, et al. Resveratrol mobilizes endogenous copper in human peripheral lymphocytes leading to oxidative DNA breakage: a putative mechanism for chemoprevention of cancer. *Pharm Res* 2010;27:979–88.
- [34] Frémont FL. Minireview: biological effects of resveratrol. *Life Sci* 2000;66:663–73.
- [35] Vargas JE, Filippi-Chiela EC, Suhre T, Kipper FC, Bonatto D, Lenz G. Inhibition of HDAC increases the senescence induced by natural polyphenols in glioma cells. *Biochem Cell Biol = Biochimie et biologie cellulaire* 2014;92:297–304.
- [36] Zamin LL, Filippi-Chiela EC, Dillenburg-Pilla P, Horn F, Salbego C, Lenz G. Resveratrol and quercetin cooperate to induce senescence-like growth arrest in C6 rat glioma cells. *Cancer Sci* 2009;100:1655–62.
- [37] Brittes J, Lucio M, Nunes C, Lima JL, Reis S. Effects of resveratrol on membrane biophysical properties: relevance for its pharmacological effects. *Chem Phys Lipids* 2010;163:747–54.
- [38] Fukuhara K, Miyata N. Resveratrol as a new type of DNA-cleaving agent. *Bioorg Med Chem Lett* 1998;8:3187–92.
- [39] Rae TD, Schmidt PJ, Pufahl RA, Culotta VC, O'Halloran TV. Undetectable intracellular free copper: the requirement of a copper chaperone for superoxide dismutase. *Science* 1999;284:805–8.
- [40] Kaplan JH, Lutsenko S. Copper transport in mammalian cells: special care for a metal with special needs. *J Biol Chem* 2009;284:25461–5.
- [41] Markossian KA, Kurganov BI. Copper chaperones, intracellular copper trafficking proteins. Function, structure, and mechanism of action. *Biochemistry, Biokhimiia* 2003;68:827–37.
- [42] Govindarajua M, Shekarb HS, Sateeshac SB, et al. Copper interactions with DNA of chromatin and its role in neurodegenerative disorders. *J Pharm Anal* 2013;3:354–9.
- [43] Rouault TA. Systemic iron metabolism: a review and implications for brain iron metabolism. *Pediatr Neurol* 2001;25:130–7.
- [44] Azmi AS, Bhat SH, Hanif S, Hadi SM. Plant polyphenols mobilize endogenous copper in human peripheral lymphocytes leading to oxidative DNA breakage: a putative mechanism for anticancer properties. *FEBS Lett* 2006;580:533–8.
- [45] Gutteridge JM. Copper-phenanthroline-induced site-specific oxygen-radical damage to DNA. Detection of loosely bound trace copper in biological fluids. *Biochem J* 1984;218:983–5.
- [46] Bhat R, Hadi SM. DNA breakage by tannic acid and Cu(II): sequence specificity of the reaction and involvement of active oxygen species. *Mutat Res* 1994;313:39–48.
- [47] Nemery B. Metal toxicity and the respiratory tract. *Eur Respir J* 1990;3:202–19.
- [48] Wenzel E, Somoza V. Metabolism and bioavailability of trans-resveratrol. *Mol Nutr Food Res* 2005;49:472–81.
- [49] van de Wetering K, Burkon A, Feddema W, et al. Intestinal breast cancer resistance protein (BCRP)/Bcrp1 and multidrug resistance protein 3 (MRP3)/Mrp3 are involved in the pharmacokinetics of resveratrol. *Mol Pharmacol* 2009;75:876–85.
- [50] Juan ME, Gonzalez-Pons E, Planas JM. Multidrug resistance proteins restrain the intestinal absorption of trans-resveratrol in rats. *J Nutr* 2010;140:489–95.
- [51] Planas JM, Alfaras I, Colom H, Juan ME. The bioavailability and distribution of trans-resveratrol are constrained by ABC transporters. *Arch Biochem Biophys* 2012;527:67–73.
- [52] Sharan S, Nagar S. Pulmonary metabolism of resveratrol: in vitro and in vivo evidence. *Drug Metab Dispos* 2013;41:1163–9.
- [53] Andreadi C, Britton RG, Patel KR, Brown K. Resveratrol-sulfates provide an intracellular reservoir for generation of parent resveratrol, which induces autophagy in cancer cells. *Autophagy* 2014;10:524–5.
- [54] Ladurner A, Schachner D, Schueller K, et al. Impact of trans-resveratrol-sulfates and -glucuronides on endothelial nitric oxide synthase activity, nitric oxide release and intracellular reactive oxygen species. *Molecules* 2014;19:16724–36.
- [55] Shankar S, Singh G, Srivastava RK. Chemoprevention by resveratrol: molecular mechanisms and therapeutic potential. *Front Biosci* 2007;12:4839–54.
- [56] Lu DL, Ding DJ, Yan WJ, et al. Influence of glucuronidation and reduction modifications of resveratrol on its biological activities. *Chemochem* 2013;14:1094–104.
- [57] Zhou M, He JL, Yu SQ, et al. Effect of resveratrol on chronic obstructive pulmonary disease in rats and its mechanism. *Yao Xue Xue Bao* 2008;43:128–32.
- [58] Karthikeyana S, Rajendra Prasada N, Ganamanib A, Balamurugana E. Anticancer activity of resveratrol-loaded gelatin nanoparticles on NCI-H460 non-small cell lung cancer cells. *Biomed Preventive Nutr* 2013;3:64–73.
- [59] Trotta V, Lee WH, Loo CY, Young PM, Traini D, Scalia S. Co-spray dried resveratrol and budesonide inhalation formulation for reducing inflammation and oxidative stress in rat alveolar macrophages. *Eur J Pharm Sci* 2016;86:20–8.
- [60] Royce SG, Dang W, Yuan G, et al. Resveratrol has protective effects against airway remodeling and airway hyperreactivity in a murine model of allergic airways disease. *Pathobiol Aging Age Relat Dis* 2011;1.
- [61] Zunino SJ, Storms DH, Newman JW, Pedersen TL, Keen CL, Ducore JM. Dietary resveratrol does not delay engraftment, sensitize to vincristine or inhibit growth of high-risk acute lymphoblastic leukemia cells in NOD/SCID mice. *Int J Oncol* 2012;41:2207–12.
- [62] Li Z, Yang X, Dong S, Li X. DNA breakage induced by piceatannol and copper(II): Mechanism and anticancer properties. *Oncol Lett* 2012;3:1087–94.
- [63] de la Lastra CA, Villegas I. Resveratrol as an antioxidant and pro-oxidant agent: mechanisms and clinical implications. *Biochem Soc Trans* 2007;35:1156–60.